

Role of the Microbiota in Immunity and Inflammation

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The microbiota plays a fundamental role on the induction, training, and function of the host immune system. In return, the immune system has largely evolved as a means to maintain the symbiotic relationship of the host with these highly diverse and evolving microbes. When operating optimally, this immune system-microbiota alliance allows the induction of protective responses to pathogens and the maintenance of regulatory pathways involved in the maintenance of tolerance to innocuous antigens. However, in high-income countries, overuse of antibiotics, changes in diet, and elimination of constitutive partners, such as nematodes, may have selected for a microbiota that lack the resilience and diversity required to establish balanced immune responses. This phenomenon is proposed to account for some of the dramatic rise in autoimmune and inflammatory disorders in parts of the world where our symbiotic relationship with the microbiota has been the most affected.

"The states of health or disease are the expressions of the success or failure experienced by the organism in its efforts to respond adaptively to environmental challenges."—Rene Dubos, 1965

Introduction

Multicellular organisms exist as meta-organisms comprised of both the macroscopic host and its symbiotic commensal microbiota. With an estimated composition of 100 trillion cells, human symbionts outnumber host cells by at least a factor of 10 and express at least 10-fold more unique genes than their host's genome (Ley et al., 2006a). These complex communities of microbes that include bacteria, fungi, viruses, and other microbial and eukaryotic species provide a tremendous enzymatic capability and play a fundamental role in controlling many aspects of host physiology. Over the past few years, the field of immunology has been revolutionized by the growing understanding of the fundamental role of the microbiota in the induction, education, and function of the mammalian immune system.

The immune system is composed of a complex network of innate and adaptive components endowed with an extraordinary capacity to adapt and respond to highly diverse challenges. Collectively, this cellular network acts as a formidable regulator of host homeostasis that operates to sustain and restore tissue function in the context of microbial and environmental encounters. The development of defined arms of the immune system—and, more particularly, those associated with adaptive immunity—has coincided with the acquisition of a complex microbiota, supporting the concept that a large fraction of this machinery has evolved as a means to maintain a symbiotic rela-

tionships with these highly diverse microbial communities. In turn, the microbiota promote and calibrate multiple aspects of the immune system.

When operating optimally, the immune system-microbiota alliance interweaves the innate and adaptive arms of immunity in a dialog that selects, calibrates, and terminates responses in the most appropriate manner. However, both the acquisition of a complex immune system and its reliance on the microbiota came at a price. Pathologies that increasingly affect humans, such as allergies, autoimmune, and inflammatory disorders, all arise from a failure to control misdirected immune responses against self, microbiota-derived, or environmental antigens. Further, alteration of the composition and function of the microbiota as a result of antibiotic use, diet evolution, and recent elimination of constitutive partners such as helminth worms has transformed our microbial allies into potential liabilities. Although members of the microbiota are often referred to as commensals, symbiosis between the microbiota and its mammalian host encompasses various forms of relationship, including mutualistic, parasitic, or commensal. However, how defined members of the microbiota interact with their host can be highly contextual, with the same microbe developing as mutualist or parasite according to the nutritional, coinfection, or genetic landscape of its host. Over the past decade, exploration of optimal and dysregulated partnerships between the microbiota and its mammalian host has taken center stage in the field of immunology and has led to the rediscovery of a more holistic view of host physiology. Indeed, the notion that microbial partners can promote human health is not a recent concept and was originally proposed by the seminal work of Döderlein (1892) and his understanding of the role of lactobacilli



as gatekeepers of the vaginal ecosystem as well as the observation of Metchnikoff associating prolonged life with fermented milk products. Recent sequencing efforts of the human metagenome have changed our understanding of the microbiome and how variations in these populations can contribute to disease states. In this Review, we will discuss some of the major concepts that have emerged from the recent dialog between immunologists, geneticists, microbiologists, and clinicians that highlight the complex role of the microbiota on the immune system in health and diseases.

Microbiota-Immune System Interaction during Development

Under normal conditions, the fetal gastrointestinal tract is believed to be sterile, with the first exposure of the immune system to commensals occurring during the passage through the birth canal. These early interactions are considered to set the tone of the mucosal and systemic immune system for the long term. The mechanism by which neonate tissues adapt to the formidable challenge of microbial colonization remains incompletely understood, but factors contained in maternal milk are believed to define some of these early responses to commensals. Indeed, colostrum and breast milk contain live microbes, metabolites, IgA, and immune cells as well as cytokines. These factors synergize to shape the breast-fed infant microbiota and the response of the host to these microbes. For instance, maternal IgA restricts immune activation and microbial attachment by binding nutritional and microbial antigens, and the presence of metabolites, including oligosaccharides in mother's milk, promotes the expansion of defined constituents of the microbiota such as Bifidobacterium (Marcobal et al., 2010; Marcobal and Sonnenburg, 2012). Bacterial translocation from the mouse gut is increased during pregnancy and lactation, and bacterially loaded dendritic cells in the milk have been proposed to contribute to neonatal immune imprinting (Perez et al., 2007).

The capacity to accept the microbiota can also be explained by the relative immaturity of the neonate immune system at birth and the tolerogenic environment that defines early mammalian life. Indeed, the developing immune system is characterized by blunted inflammatory cytokine production and skewed T and B cell development in favor of regulatory responses (PrabhuDas et al., 2011; Siegrist, 2001). Although a consequence of this blunted immune response is high susceptibility to infections, this regulatory environment ensures that the establishment of the microbiota occurs without overt inflammation. Recent reports reveal that a defined population of erythroid cells enriched in neonates contributes to the maintenance of this immunoregulatory environment and limits mucosal inflammation following colonization with the microbiota (Elahi et al., 2013). Early exposure of the host to commensals can also repress cells involved in the induction of inflammatory responses such as invariant natural killer T (iNKT) cells, an effect that has long-term consequences for the host capacity to develop inflammatory diseases (Olszak et al., 2012). A recent report proposed that this control can be mediated by the direct interaction, early in life, of unique inhibitory commensal-derived sphingolipids with iNKT cells (An et al., 2014).

One of the primary modes of dialog between the host and the microbiota is mediated by the recognition of conserved microbial-associated molecular patterns (MAMPs). The neonate innate immune system integrates these signals in a unique way to promote healthy microbial colonization. For instance, although neonate innate cells express Toll-like receptor (TLR) ligands, their response to microbial ligands is distinct from that of adult cells, with notable impairment in the production of inflammatory mediators such as oxygen radicals and heightened production of regulatory cytokines such as IL-10 (Kollmann et al., 2012). Part of this phenomenon results from the action of the microbiota itself. Indeed, early responses to microbial ligands such as LPS, the endotoxin found in the outer membrane of Gram-negative bacterial walls, condition gut epithelial cells to become hypo-responsive to subsequent TLR stimulation (Chassin et al., 2010; Lotz et al., 2006). How the innate immune system integrates microbial-derived signals remains unclear, but recent findings support the idea that expression of epigenomemodifying enzymes by epithelial cells may be required for the coordination of commensal dependent intestinal homeostasis (Alenghat et al., 2013).

Commensals also contribute to the postnatal development of the immune system that, in turn, contributes to their containment. Studies performed in animals raised in the absence of live microbes referred to as germ-free (GF), revealed that the microbiota plays a critical role in secondary lymphoid structure development. This is particularly evident in the gastrointestinal tract, where germ-free mice are characterized by smaller Peyer's patch size and a reduced number of CD4⁺T cells and IgA-producing plasma cells (Bauer et al., 1963; Hamada et al., 2002; Macpherson et al., 2001: Mazmanian et al., 2005: Smith et al., 2007; Talham et al., 1999). In the intestine, tertiary lymphoid structures such as isolated lymphoid follicle or crytopatches are induced after birth as a result of commensal exposure (Bouskra et al., 2008; Ohnmacht et al., 2011). As further discussed below, commensals can also contribute to the fortification of the intestinal barrier by various mechanisms, including the promotion of epithelial cell maturation and angiogenesis (Hooper et al., 2001; Stappenbeck et al., 2002).

When operating properly, the highly regulatory tone of the neonate immune system and the action of commensals in the development and training of this system lead to the establishment of a durable and homeostatic host/commensal relationship. These primary encounters between the host immune system and the microbiota have profound and long-term implications for human health. Indeed, epidemiological observations revealed that alteration of the microbiota in mothers or in neonates may predispose to diseases associated with dysregulated barrier responses, such as asthma (Ege et al., 2011).

Containing the Microbiota

An important point to consider when exploring the role of the microbiota on the immune system is that pathogenicity is, in most cases, a contextual state. Indeed, the capacity of a given microbe, including those composing the microbiota, to trigger or promote disease is highly dependent on the state of activation of the host, the host's genetic predisposition, and the localization of the particular microbe. As such, the mechanisms utilized by

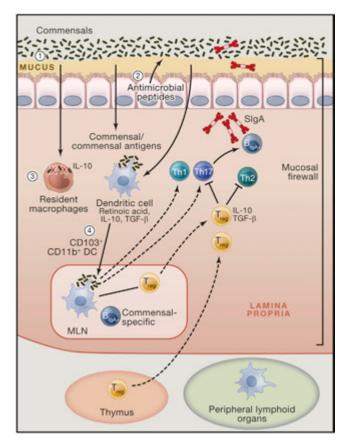


Figure 1. The Mucosal Firewall

(1) The mucus represents the primary barrier limiting contact between the microbiota and host tissue preventing microbial translocation. (2) Epithelial cells produce antimicrobial peptides that also play a significant role in limiting exposure to the commensal microbiota. (3) Translocating commensals are rapidly eliminated by tissue-resident macrophages. (4) Commensals or commensal antigens can also be captured by CD103⁺ CD11b⁺ DCs that traffic to the mLN from the lamina propria but do not penetrate further. Presentation of commensal antigens by these DCs leads to the differentiation of commensal-specific regulatory cells (T_{reg}), Th₁₇ cells, and IgA-producing B cells. Commensal-specific lymphocytes traffic to the lamina propria and Peyer's patches. In the Peyer's patches, T_{reg} can further promote class switching and IgA generation against commensals. The combination of the epithelial barrier, mucus layer, IgA, and DCs and T cells comprises the "mucosal firewall," which limits the passage and exposure of commensals to the gut-associated lymphoid tissue, preventing untoward activation and pathology.

the immune system to maintain its relationship with the microbiota are highly analogous to the ones that are used to constrain organisms with pathogenic potential.

An enormous fraction of the immune system's constitutive function is aimed at controlling our relationship with the microbiota. As such, the highest number of immune cells in the body are resident at sites colonized by commensals such as the skin or the GI tract. In turn, to protect their ecological niche, a dominant action of the healthy microbiota on the immune system is aimed at reinforcing barrier immunity and therefore their own containment. A central strategy utilized by the host to maintain its homeostatic relationship with the microbiota is to minimize contact between microorganisms and the epithelial cell surface, thereby limiting tissue inflammation and microbial translocation. In the gastrointestinal tract, home to the largest density of commensals, this segregation is accomplished by the combined action of epithelial cells, mucus, IgA, antimicrobial peptides, and immune cells. Collectively, these structural and immunological components have been referred to as the "mucosal firewall" (Figure 1) (Macpherson et al., 2009).

The mucus represents the primary shield limiting contact between the microbiota and host tissue and preventing microbial translocation (McGuckin et al., 2011). In addition to the production of mucus by goblet cells, all intestinal epithelial cell lineages can produce antimicrobial peptides that play a significant role in limiting exposure to the commensal microbiota (Hooper and Macpherson, 2010). These proteins can exert antimicrobial functions resulting from enzymatic attack of the bacterial cell wall or by disrupting the bacterial inner membrane (Hooper and Macpherson, 2010). Some of these molecules, such as α -defensins, are constitutively expressed by epithelial cells, whereas in other cases, engagement of pattern recognition receptors (PRRs) by commensally derived products is required (Hooper and Macpherson, 2010). One of the best-characterized mucosal antimicrobial peptides is RegIII_Y, which is expressed soon after birth or following colonization of germ-free mice (Cash et al., 2006). Production of this lectin is tightly controlled by the flora in a MyD88-dependent manner and has a direct microbicidal effect on Gram-positive bacteria (Brandl et al., 2007; Cash et al., 2006; Ismail et al., 2011; Mukherjee et al., 2009). Accumulation of antimicrobial peptides such as RegIII_Y in the mucus contributes to the maintenance of the segregation between the microbiota and the host intestine, creating a physical separation referred to as the "demilitarized zone" (Vaishnava et al., 2011).

Compartmentalization of intestinal bacteria also depends on secreted immunoglobulin A (IgA). IgA specific for commensals is produced with the help of intestinal dendritic cells that sample commensals associated with the epithelium and interact with B and T cells in the Peyer's patches to produce IgA specific for commensal-derived antigens (Macpherson and Uhr, 2004). Further, commensals that translocate across the intestinal epithelial cell barrier can be rapidly engulfed and eliminated by macrophages that reside in the lamina propria or carried alive by dendritic cells (DC) (Kelsall, 2008; Macpherson and Uhr, 2004). The bacteria loaded DC traffic to the mesenteric lymph node via the intestinal lymphatics but do not penetrate further, allowing the induction of a mucosal-compartmentalized IgA response (Macpherson and Uhr, 2004). IgA⁺B cells migrate to the intestinal lamina propria and secrete IgA that are subsequently transcytosed across epithelial cells. These transcytosed IgAs control host commensal interaction by both impacting commensal gene expression (Peterson et al., 2007) and preventing adhesion of commensal bacteria to the epithelial surfaces (Figure 1) (Boullier et al., 2009; Fernandez et al., 2003; Hornquist and Lycke, 1995; Martinoli et al., 2007; Wade and Wade, 2008; Wei et al., 2011). Mucosal IgA responses lack classical memory characteristics and are able to respond to flux in commensal microbiota composition. Indeed, established IgA-producing clones are outcompeted by novel antibacterial responses, allowing the mucosal immune system to respond to a constantly changing microbiota (Hapfelmeier et al., 2010).

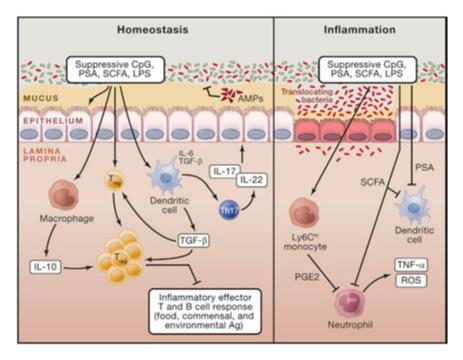


Figure 2. Promotion of Immune Regulation by the Microbiota during Steady State and Inflammation

(Left) Commensals promote the induction of regulatory T cells via direct sensing of microbial products or metabolites by T cells or dendritic cells. Further commensals promote the induction of Th17 cells that can regulate the function and homeostasis of epithelial cells. In the context of inflammation, similar mechanisms may account for the regulatory role of the microbiota. (Right) Commensal-derived metabolites can also have a local and systemic effect on inflammatory cells. For example, SCFA can inhibit neutrophil activation. Upon entrance in the tissue, inflammatory monocytes can also respond to microbial-derived ligands by producing mediators such as PGE₂ that limit neutrophil activation and tissue damage.

cells referred to as group 3 innate lymphoid cells (ILCs). Although some reports propose that the development of these cells is independent of signals derived from the microbiota, their phenotype and functional capacity can evolve to accommodate physiologic alterations

Most activated or memory T cells reside in tissues that are constitutively colonized by commensals such as the skin and the GI tract. Notably, at steady state, most IL-17 (Th17) and IFN_γ-producing (Th1) T cells are found in the GI tract and develop from signals that are derived from the microbiota (Gaboriau-Routhiau et al., 2009; Ivanov et al., 2008; Macpherson and Harris, 2004). The current view is that constitutive sensing of commensals plays an important homeostatic role, whereas active responses against the flora are believed to be associated with pathogenesis. However, this distinction is clearly not absolute and needs to be revisited in light of the observation that healthy human serum normally contains antibodies and T cells specific to commensals (Ergin et al., 2011; Macpherson et al., 1996), suggesting that a certain degree of commensal recognition is a common occurrence and, in most circumstances, is not associated with pathogenic responses. Although tissue-derived cues can dictate the induction and maintenance of Th17 cells irrespective of antigen specificity, we could speculate that, in a similar manner to that proposed for Treg cells, antigenic specificities of tissue-resident effector T cells are highly enriched for commensal antigens. Notably, Th17 cells produce cytokines such as IL-17 and, more particularly, IL-22, which contribute to the homeostatic dialog with commensals via the capacity of these cytokines to act on epithelial cell function. Failure to maintain the Th17 lineage in the gut, as observed during HIV or SIV infection, is associated with microbial translocation that contributes to dissemination of the virus (Klatt et al., 2013). The action of IL-22 on the mucosal immune system is highly pleiotropic and promotes the production of antimicrobial peptides, enhances of epithelial regeneration, increases mucus production, and regulates wound repair (Wolk and Sabat, 2006; Zenewicz and Flavell, 2011). This cytokine can also be produced by other cell lineages and, more particularly, by a population of gut-resident innate

in the intestinal environment following microbial colonization at birth (Satoh-Takayama et al., 2008). Production of IL-22 by ILC promotes the containment of specific members of the microbiota community and, more particularly, microbes that reside in mucosal lymphoid structures, such as bacteria of the *Alcaligenes* genus (Qiu et al., 2013; Sonnenberg et al., 2012). Thus, in addition to broad and nonspecific modes of commensal containment, discrete pathways may have evolved to promote the selective containment of communities of microbes residing in unique ecological niches.

Induction of Regulatory Responses by the Microbiota

Maintenance of tissue homeostasis is imperative for host survival. This fundamental process relies on a complex and coordinated set of innate and adaptive responses that selects and calibrates responses against self, food, commensals, and pathogens in the most appropriate manner. To this end, specialized populations of cells have to integrate local cues such as defined metabolites, cytokines, or hormones, allowing the induction of responses in a way that preserves the physiological and functional requirements of each tissue. As such, the regulatory pathways that are involved in the maintenance of a homeostatic relationship with the microbiota are likely to be tissue specific. However, most of our current understanding of commensaldependent regulatory pathways relates to the gastrointestinal environment. In the gut, the formidable challenge represented by the exposure to the microbiota, food-derived antigens, metabolites, and pathogens requires a highly complex network of regulatory pathways that is only beginning to be understood (Figure 2). Failure to regulate these responses can lead to severe pathological outcomes ranging from inflammatory bowel diseases (IBD) to allergies or, as further discussed, metabolic syndromes.

Commensals are a critical and active inducer of regulatory responses. Notably, the establishment of tolerance-the active suppression of inflammatory responses to food and other orally ingested antigens-could not be induced in the absence of gut flora-derived signals (Kiyono et al., 1982; Sudo et al., 1997; Wannemuehler et al., 1982; Weiner et al., 2011). Although immunological tolerance is likely to be achieved via multiple and redundant mechanisms (Weiner et al., 2011), over the past few years, Foxp3+ regulatory T (T_{reg}) cells have taken central stage in our understanding of this process. These cells maintain both peripheral and mucosal homeostasis throughout the lifespan of the host, and disruption of the homeostasis of these cells results in loss of oral tolerance and development of aberrant effector responses in the gut (Josefowicz et al., 2012a; Mucida et al., 2005; Worbs et al., 2006). Although Foxp3⁺ T_{reg} can arise as differentiated cells in the thymus, the gastrointestinal tract environment is a privileged site for the induction of $\mathsf{T}_{\mathsf{reg}}$ cells in response to oral antigens (Coombes et al., 2007; Mucida et al., 2005; Sun et al., 2007). A current consensus is that optimal maintenance of tolerance to commensal and environmental antigens requires the combined effect of both thymically and GI-induced T_{reg} (Cebula et al., 2013; Josefowicz et al., 2012a; Lathrop et al., 2011). This specialized property of the gut to induce T_{reg} can be, at least in part, explained by the presence of specialized populations of antigen-presenting cells, such as the CD103⁺CD11b⁺ DC. Notably, these gut-resident dendritic cells are endowed with the capacity to produce factors involved in the induction of T_{reg}, such as the cytokine TGF- β and the vitamin A metabolite retinoic acid (RA) (Coombes et al., 2007; Mucida et al., 2007; Sun et al., 2007). Tissue-specific factors such as vitamin A and MUC2, a mucus glycoprotein produced by intestinal goblet cells, contribute to the regulatory specialization of mucosal dendritic cells (Klebanoff et al., 2013; Shan et al., 2013). The importance of this pathway for the control of mucosal homeostasis is highlighted by the finding that a proportion of induced T_{req} in the colonic tissue is specific for antigens derived from the commensal microbiota (Lathrop et al., 2011). Induction of T_{req} cells is proposed as one of the mechanisms of action of probiotics-defined bacteria known to confer a health benefit to the host. Indeed, some of the regulatory effects of probiotics in the context of inflammatory diseases and atopic eczema in neonates and infants is believed to be associated with the induction or expansion of T_{reg} (Di Giacinto et al., 2005; Feleszko et al., 2007; Karimi et al., 2009; Zoumpopoulou et al., 2008) and the manipulation of mucosal DCs toward a proregulatory function (Foligne et al., 2007; Smits et al., 2005). Commensals can also control oral antigen sampling by mucosal DCs and promote the induction of lamina propriaresident macrophages associated with local expansion of Trea cells (Chieppa et al., 2006; Niess and Adler, 2010). Aside from the direct influence of the microbiota on the immune machinery associated with the induction of oral tolerance, commensalspecific T_{req} can promote class switching to IgA in an antigenspecific manner (Cong et al., 2009; Tsuji et al., 2009), thereby controlling the host relationship with the microbiota via multiple mechanisms (Peterson et al., 2007; Suzuki et al., 2004) (Figure 2).

The first demonstration that a unique symbiont molecule could promote regulatory responses was provided by the identification of the polysaccharide A (PSA), which is produced by a prominent human symbiont Bacteroides fragilis (Mazmanian et al., 2005). B. fragilis, via PSA expression, can protect mice from experimental colitis induced by Helicobacter hepaticus, a commensal bacterium with pathogenic potential (Mazmanian et al., 2008). This protective activity was associated with the capacity of PSA to induce and expand IL-10-producing Treg cells (Mazmanian et al., 2008; O'Mahony et al., 2008; Ochoa-Repáraz et al., 2010a). B. fragilis was able to promote Treg cell function and induction via engagement of the microbial-derived PSA with TLR2 expressed by T cells, a phenomenon associated with the capacity of this bacterium to also limit Th17 responses (Round et al., 2011). The discovery of a link between defined members of the microbiota and the induction of regulatory cells able to limit mucosal inflammation and promote tolerance led to a rational approach for the identification of the next generation of probiotics with superior capacity to induce Treg cells. Induction of Treg cells is not restricted to B. fragilis, as the presence of an indigenous Clostridium species also promotes Treg cell accumulation via, at least in part, its capacity to create a TGF- β rich environment (Atarashi et al., 2011). Of note, optimal induction of T_{rea} in the colonic environment relies on the synergistic effect of a consortium of Clostridia strains, whereas individual species have a modest effect on the immune system (Atarashi et al., 2013). Based on the fundamental role of $T_{\rm reg}$ in maintaining mucosal homeostasis, it is likely that, rather than being restricted to defined commensals, a large fraction of any given microbiota or microbiota metabolism may have evolved to favor this aspect of the regulatory network. Indeed, recent findings have shown that many organisms can increase the frequency of $T_{\mbox{\scriptsize regs}}$ in the colon (Faith et al., 2014; Geuking et al., 2011).

Mammals rely on bacteria to break down undigestible dietary components such as fiber (Ley et al., 2006a). One dominant metabolite resulting from this process is short-chain fatty acids (SCFA), ubiquitous bacterial fermentation products that are highly enriched in the colonic environment (Cummings et al., 1987). Although a role for SCFA in controlling various aspects of immune responses has been long recognized (Meijer et al., 2010), their link to lymphocyte function has only recently been appreciated. Notably, SCFA and, in particular, butyrate regulate the size and function of the regulatory T cell network by promoting the induction and fitness of regulatory T cells in the colonic environment (Arpaia et al., 2013; Furusawa et al., 2013; Smith et al., 2013). Butyrate is well known to regulate gene expression epigenetically by inhibiting histone deacetylases (HDACs) (Davie, 2003), and this property is currently proposed as an underlying mechanism for enhanced T_{req} generation in the gut. The action of SCFA likely results from the manipulation of various cells involved in the induction of regulatory responses, and indeed, the effect of SCFA on both T cell and dendritic cells has been linked to this process (Arpaia et al., 2013; Furusawa et al., 2013; Smith et al., 2013).

Altogether, these results reveal a major role for the microbiota in shaping the repertoire, number, and activation of tissue-resident T_{reg} cells and in the maintenance of host-microbe mutualism at barrier sites. Based on the abundance and complexity of the flora, one could also speculate that opportunity for cross reactivity between commensals, pathogenic, and environmental

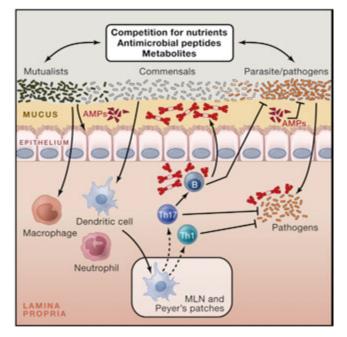


Figure 3. Promotion of Protective Immunity by the Microbiota

The symbiosis between the microbiota and its mammalian host encompasses multiple relationships, including mutualistic, parasitic, and commensal. The capacity of a given microbe, including those composing the microbiota, to trigger or promote disease is highly contextual, and some microbes can shift from mutualist to commensal to parasite according to the state of activation of the host, co-infection, or localization. Commensals can control microbes with pathogenic potential (as normal constituent of the microbiota or acquired) via distinct mechanisms. Commensals can compete for nutrients and produce of pathogens. Commensals can promote the production of antimicrobial peptides by epithelial cells and reinforce tight junctions. Finally, commensals can modulate the function of dendritic cells and other innate cells both locally and systemically in a manner that promotes the induction of effector T and B cells responses against pathogens. When uncontrolled, this adjuvant property of the microbiota can promote inflammatory and autoimmune disorders.

antigens is high. Thus, microbial pressure in the gut could lead to the induction and maintenance of a pool of activated T_{reg} cells that may not only contribute to the maintenance of a mutualistic relationship with the microbiota, but also to the systemic control of immune responses.

One of the first demonstrations of the protective role of commensals during acute injury was provided by the observation that, in the gut, TLR activation by commensals was required to promote tissue repair and host survival (Rakoff-Nahoum et al., 2004). Part of the protective effect of the microbiota in the context of inflammation relates to its capacity to sustain the aforementioned regulatory network (Arpaia et al., 2013; Bouladoux et al., 2012; Furusawa et al., 2013; Smith et al., 2013). Commensal-derived products can also act by controlling directly or indirectly the function of inflammatory cells. For instance, recognition of the commensal-derived metabolites SCFA by innate immune cells is critical for the regulation of inflammation in response not only to intestinal injury, but also in models of arthritis and allergy (Maslowski et al., 2009). Furthermore, commensals can also tune the function of inflammatory monocytes, a population of cells involved in the control of pathogens

(Figure 2). During acute mucosal infection, encounter of inflammatory monocytes with the microbiota in the gastrointestinal tract promotes their production of the lipid mediator PGE₂ that, in turn, limits the level of activation of tissue-damaging neutrophils (Grainger et al., 2013). Although most of what is known today about the regulatory properties of the microbiota arises from the exploration of the bacterial component of the flora, other microbes such as fungi and virus are likely to promote similar or complementary aspects of the regularly network. In the gastrointestinal tract, interaction of commensal fungi with the C-type lectin receptor Dectin 1 was able to prevent inflammation in the context of acute mucosal injury (liev et al., 2012).

Induction of Protective Responses by Commensals

Tissues that are natural habitats of the microbiota such as the skin, the GI tract, or the lung are also the portals by which pathogens access the host and, often, the primary site of infections. This implies that the initial encounter of pathogens with the immune system occurs in an environment conditioned and regulated by its endogenous microbiota. As such, the fate of commensals and pathogens (as well as their classification) is highly interdependent (Figure 3). Notably, commensals can directly and dynamically interact with pathogens and immune cells, and the results of this interaction can define the pathogenesis and outcome of a given infection.

Protection of the host from exogenous pathogens by commensal bacteria, a phenomenon referred to as colonization resistance, was described more than five decades ago (Buffie and Pamer, 2013; van der Waaij et al., 1971). One of the major forms of interaction between the microbiota and invading microbes relates to the need for both forms of organisms to compete for the same ecological niche. Consequently, commensals have been shown to limit pathogen colonization through competition for defined metabolites in a process referred to as colonization resistance (Kamada et al., 2013). Alteration of nutrient availability by the host microbiota can also have profound consequences on the expression of virulence genes and growth rate of pathogens such as enterohemorragic E. coli or Clostridium difficile (Kamada et al., 2013; Ng et al., 2013; Pacheco et al., 2012). Manipulation of microbial virulence can also occur as a consequence of commensal metabolism. In some instances, the very same metabolites involved in the manipulation of the immune system, such as SCFA, can also directly act on pathogens by downregulating the expression of virulence genes such as those encoding the type 3 secretion system in Salmonella enterica and typhimurium (Gantois et al., 2006). Commensals can also promote the establishment of an environment hostile for pathogen establishment, with the best illustration in the vaginal environment in which lactobacilli can protect from pathogenic colonization via reduction of the local pH (Turovskiy et al., 2011). Finally, commensals can also produce antimicrobial peptides that directly affect pathogen growth or survival (Hammami et al., 2013). For instance, E. coli produces bacteriocins-proteinaceous toxins that specifically inhibit the growth of the same or similar bacterial strains-thus impairing the growth of the related pathogen, enterohemorrhagic E. coli. Additionally, the dominant skin commensal Staphylococcus epidermidis produces several antimicrobial proteins and proteases that can limit the biofilm formation of *Staphylococcus aureus* (lwase et al., 2010; Schamberger and Diez-Gonzalez, 2002) (Figure 3).

In tandem with its direct action on invasive microbes, the microbiota's capacity to control infection is also associated with its remarkable ability to promote and calibrate both innate and adaptive immunity (Molloy et al., 2012). Indeed, commensals play a fundamental role in both the training of the immune system and its functional tuning, thereby acting as adjuvants to the immune system as a whole. As previously discussed, commensals can reinforce defined components of barrier immunity, an effect that has the dual function of promoting their own containment and limiting pathogen invasion. For example, the commensaldependent antimicrobial peptide RegIII_Y not only contributes to maintenance of segregation between the microbiota and host epithelial cells, but also promotes protection against defined pathogens, such as vancomycin-resistant Enterococcus faecalis (Brandl et al., 2008; Vaishnava et al., 2008, 2011). An important role of the microbiota is associated with its capacity to condition cells to respond to infectious challenge both systemically and locally. For instance, commensals can tune innate cells in a way that allows them to rapidly respond to pathogen encounters. An example of this control is provided by the capacity of the gut microbiota to control the production of IL-1 ß, a cytokine that is involved in host defense. The microbiota contributes to the homeostatic production of pro-IL-1ß by intestinal resident macrophages in a MyD88-dependent manner, thereby priming these cells to respond rapidly to enteric infections by conversion of pro-IL-1β to mature active IL1β (Franchi et al., 2012).

Some of the stimulatory effects of the microbiota can be attributed to dominant microbial-derived signals. Indeed, commensals and pathogenic microbes interact with the host immune system through conserved ligands that are immutable features of microorganisms (Sansonetti and Di Santo, 2007). Many of these ligands signal through the Toll-like and Nod-like families of receptors (Sansonetti and Di Santo, 2007; Takeda et al., 2003). One example of this is flagellin, a structural protein that forms the main portion of flagella and promotes bacterial chemotaxis, adhesion, and invasion of host tissue by pathogenic bacteria. Flagellin is also expressed by a large number of commensal bacteria, and several lines of evidence indicate that interaction of commensal flagellin with Toll-like receptor 5, in particular in the context of barrier breach, plays an important role in the promotion of mucosal immunity. Notably, dendritic cells that reside in the lamina propria of the intestine are poised to respond to flagellin by rapidly expressing chemokines and antimicrobial peptides, as well as cytokines involved in the initiation of immune responses (Kinnebrew et al., 2012; Uematsu et al., 2008; Uematsu et al., 2006). In response to flagellin, CD103⁺ DC produce IL-23, in part, to induce IL-22 by innate lymphoid cells (ILCs), thereby promoting epithelial-mediated host defense (Kinnebrew et al., 2012). Unmethylated cytosine phosphate guanosine (CpG) dinucleotides, which are abundant within the prokaryotic DNA of intestinal flora, can contribute to intestinal homeostasis under steady-state conditions (Hall et al., 2008), and constitutive interaction between CpG-expressing commensal DNA and TLR9 can act as a local adjuvant of immune responses (Hall et al., 2008). Nonetheless, under homeostatic conditions, both inflammatory and regulatory signals are constantly integrated-the sum of these signals leading to the establishment of an immune tone compatible with tissue immunity.

The capacity of the microbiota to stimulate innate responses translates into its important role in the induction of adaptive immunity (Figure 3). Indeed, early studies have identified significantly impaired host immune responses to pathogens in mice treated with antibiotics or raised under germ-free conditions (Cebra, 1999; Hall et al., 2008; Ivanov et al., 2009; Mazmanian et al., 2005). One of the first demonstrations of this "adjuvant" effect was revealed in a parasitic model of small intestine infection with Encephalitozoon cuniculi in which protective Th1 and Th17 responses were severely compromised in the absence of commensals (Hall et al., 2008). Similarly, protective Th17 responses failed to develop in response to Citrobacter rodentium, a model of attaching and effacing infection that primarily affects the large intestine (lvanov et al., 2009). The adjuvant property of the microbiota has also been revealed in a model of oral vaccination (Hall et al., 2008), an observation that may help to explain some of the failures of oral vaccination in developing countries in which malnutrition and infection have profoundly affected the microbiota (Korpe and Petri, 2012).

In addition to the pleiotropic effects of conserved microbial ligands or metabolites in the education and function of the immune system, it is now becoming clear that unique microbes or groups of bacteria can dominantly influence immune system development and function under steady-state and inflammatory conditions. In the language of ecological systems, these organisms of paramount importance are termed "keystone species." The prototype of a keystone species in the GI tract is represented by the interaction between segmented filamentous bacteria (SFB) and the immune system. Under steady-state conditions, this spore-forming Gram-positive anaerobic bacteria colonizes the terminal ileum of mice. SFB has a dominant effect on the mucosal immune system by promoting the accumulation of both Th17 and Th1 cells in the small intestine and driving the production of IqA (Gaboriau-Routhiau et al., 2009; Ivanov et al., 2009; Umesaki et al., 1999). In contrast to most commensals that reside outside of the "demilitarized zone," SFB interacts closely with the mucosal tissue via tight adhesion to Peyer's patches and epithelial cells, inducing cytoskeletal reorganization in these cells at the site of contact (Ivanov et al., 2008; Sczesnak et al., 2011). This intimate contact with epithelial cells, a property shared by a select minority of commensal organisms, is believed to account for the capacity of SFB to heighten tissue immunity and promote protective responses to pathogens (lvanov et al., 2009). The presence of bacteria with a keystone role on the immune system raises an intriguing possibility. Although optimal control of host metabolism and physiology may rely on complex and redundant populations of microbes, we could speculate that a more limited number of microbes may act as adjuvants of immune responses. Indeed, overpopulation of the GI tract with bacteria that have enhanced inflammatory potential can have local and systemic pathological consequences. Under steadystate conditions, these ambivalent members of the microbiota, such as E. coli or SFB, are maintained in check by the immune system and coopted by the host to control invasive microbes.

This protective effect of the microbiota has been revealed in clinical and experimental settings in which broad antibiotic

treatment can allow the domination of intestinal microbiota with drug-resistant microbes such as vancomycin-resistant enterococcus (VRE), a pathogen that causes bloodstream infections in immunocompromised patients (Buffie and Pamer, 2013; Murray, 2000). Infections caused by multidrug-resistant organisms are on the rise and have developed into endemic and epidemic situations worldwide (Gupta et al., 2011). Harnessing the microbiota to combat these infections represents an important therapeutic avenue, with the most spectacular results obtained thus far in the context of Clostridium difficile colitis (van Nood et al., 2013). During this recurrent infection, transfer of a microbiota from a healthy donor eradicated the infection with a remarkable efficiency (van Nood et al., 2013). Again highlighting the concept of defined microbes endowed with superior adjuvant capacity, the protective effect of the microbiota in VRE-infected patients was highly dependent on the presence of the commensal Barnesiella (Ubeda et al., 2013).

Thus, the microbiota is a required component of the effector response of the host. Although currents studies are attempting to link defined microbes to unique immunological states in human, the microbiota is a highly dynamic and complex composite of microbes all expressing a large number of potential ligand and metabolites. Under homeostatic conditions, both inflammatory and regulatory signals are constantly integrated—the sum of these signals leading to the establishment of a controlled inflammation compatible with tissue immunity. Therefore, under most settings, changes in inflammatory state associated with the microbiota are unlikely due to a single microbial product or metabolite but may result from a shift in the balance of signals.

Systemic Control of Protective Immunity

Though it is readily accepted that shifts in the gut microbiota composition and density can affect local immune responses, it is becoming apparent that these changes can also alter immunity and inflammation in organs distal from the intestine (Belkaid and Naik, 2013). For instance, reduction of out commensals via broad-spectrum antibiotic treatment results in blunted T and B cell response against intranasal infection with influenza (Ichinohe et al., 2011). This effect of the microbiota can be linked to its capacity to promote the inflammasome-mediated induction of IL-1 β and IL-18 secretion (Ichinohe et al., 2011). In this setting, rectal administration of TLR agonists restored protective immune responses, indicating that either the microbial products are capable of diffusing systemically or that inflammasome activation does not need to occur at the site of infection (Ichinohe et al., 2011). Antibiotic treatment also impaired adaptive and innate antiviral responses following exposure to systemic lymphocytic choriomeningitis virus (LCMV) and musosal influenza virus (Abt et al., 2012). Genome-wide transcriptional profiling of macrophages from antibiotic-treated mice revealed a broad decrease of genes associated with antiviral immunity (Abt et al., 2012). This bystander control of peripheral responses can be, at least in part, explained by the unique requirement of the GI tract for absorption resulting in the constant diffusion of low-level microbial products such as TLR or NOD ligands and metabolites into the bloodstream. For instance, commensalderived peptidoglycan can be found in the serum and has been shown to improve the killing of Streptococcus pneumonia and *Staphylococcus aureus* by bone-marrow-derived neutrophils in a NOD1-dependent manner (Clarke et al., 2010). Experimental evidence suggests that the tonic sensing of commensal products or metabolites present in the blood stream also contributes to steady-state hematopoiesis and monocyte egress from the bone marrow (Maslowski et al., 2009; Shi et al., 2011).

Recent evidence implies that the capacity of commensals to calibrate systemic immunity has profound consequences in the context of immunotherapy. Total body irradiation, used in defined settings of immunotherapy and bone marrow transplantation, is associated with gut damage and microbial translocation, providing an adjuvant effect to the transferred anti-tumoral T cells (Paulos et al., 2007). A similar mechanism is proposed to explain the protective role of the microbiota in the context of chemotherapy. Cyclophosphamide, a clinically important cancer drug, also leads to intestinal damage, bacterial dysbiosis, and translocation and induction of anti-commensal Th17 responses that collectively contribute to the anti-tumoral response (Viaud et al., 2013). Some of the effect of the microbiota can also occur independently of gut damage. Indeed, disruption of the gut flora via antibiotic treatment or in germ-free mice impairs the capacity of the host to control subcutaneous tumors during immunotherapy (lida et al., 2013). In these experimental settings, the protective effect results from the capacity of the commensalderived ligands to control the status of activation of tumor myeloid cells and, more particularly, their level of TNF- α and reactive oxygen species, both associated with optimal tumor control (lida et al., 2013). Remarkably, tumor control was associated with the presence of defined commensal species such as Alistipes shahii (lida et al., 2013). Thus, commensals and more particularly defined member of the microbiota can control various aspects of immunity associated with anti-tumoral responses, an effect that has profound clinical implications.

Altogether, these data reveal that exposure to microbial ligands shapes systemic immunity at both the steady state and in the context of inflammation. The basal tuning of the immune system associated with constant sensing of microbial products or ligands implies that subtle changes in this conditioning may have long-term consequences on the capacity of the host to mount systemic immune responses and develop inflammatory diseases. The mechanism underlying this phenomenon remains incompletely understood, but we could speculate that commensal bacteria-derived signals can influence gene expression profiles of immune cells via epigenetic modifications of genes involved in innate responses, thus creating a transcriptional state that enables basal expression of host-defense factors and rapid responses upon encounter with a pathogen. How permanent the effects of microbial sensing are and to what extent tissue and hematopoetic stem cells are permanently influenced by this tonic sensing remain to be explored.

Compartmentalized Control of Tissue Immunity

Although the above mentioned observations propose that gut commensals can control the systemic threshold of activation of innate and adaptive cells, these studies do not exclude a direct role for commensals residing in the lung, skin, or other barrier sites in the control of local immunity. Indeed, microbial surveys unveiled the remarkable partitioning of commensals within the human body, with each tissue and microenvironment hosting unique microbial communities (Belkaid and Naik, 2013). Thus, each barrier tissue is a complex and in some cases unstable composite of microbes and host structural, hormonal, nervous, and immunological networks, with each of these systems potentially controlled by resident microbiota.

The skin, the largest organ of the body, represents a critical interface between the host and the environment. Unbiased microbial sequencing has shown the presence of highly diverse and specific commensal niches along distinct topographical sites of the skin (Costello et al., 2009; Grice et al., 2009). Although the skin is a rather inhospitable environment-poor in nutrients and moisture-up to 1 billion bacteria inhabit a typical square centimeter of human skin, covering the surface and extending down into the sebaceous glands and hair follicles (Grice et al., 2008). In contrast to the known role of the gut microbiota in promoting the gastrointestinal-associated lymphoid tissue (GALT) development, skin commensals are not required for the development of associated lymphoid tissue (Naik et al., 2012). The skin-resident bacteria, such as Staphylococcus epidermidis, can control fundamental aspects of local immunity and tissue repair (Lai et al., 2009; Naik et al., 2012). Skin commensals do not affect the capacity of T cells to be primed or to migrate to the skin but modulate dermal T cell function by tuning the cutaneous inflammatory milieu and, more particularly, the production of IL-1 α that, in turn, directly controls the capacity of dermal resident T cells to produce inflammatory cytokines such as IFN- γ and IL-17A (Naik et al., 2012). Thus, in contrast to the role of the gut microbiota, the action of skin commensals on the local immune system is discrete and highly compartmentalized.

The oral cavity also harbors a unique and complex microbial community accumulating on both the hard and soft oral tissues in sessile biofilms (Avila et al., 2009). One of the proposed roles of the oral microbiome on the immune system is associated with its capacity to promote inflammasome activity. leading to the local increase of the inflammatory cytokine IL1 β (Dixon et al., 2004). At other sites, such as the lung or vaginal mucosa, the role of commensals on tissue immunity remains largely unknown. In the absence of commensals, the number of infiltrating Th2 lymphocytes and eosinophils was elevated, and the composition and status of activation of lung dendritic cells were altered during airway inflammation (Herbst et al., 2011). Additionally, intranasal priming of mice with live or heat-inactivated Lactobacillus can dampen local responses, thereby protecting against lethal sequelae infection with the virulent pathogen pneumonia virus of mice (PVM) (Gabryszewski et al., 2011; Garcia-Crespo et al., 2012). These results are consistent with the notion that commensal bacteria, in most tissues, can establish a threshold of activation and regulation required for immune fitness. Nonetheless and despite the growing number of studies that associate commensal dysbiosis at diverse tissue sites with specific pathologies such as psoriasis, atopic dermatitis, and asthma (Abreu et al., 2012; Belda-Ferre et al., 2012; Hilty et al., 2010; Kong et al., 2012; Srinivasan et al., 2010), little is known about the effect of these unique microbial communities in the control of tissue-specific immunity. Based on our understanding of tissue specialization, we could postulate that these unique microbial communities have coevolved with their host to finely tune the unique environment of each tissue site.

Role of the Microbiota in the Pathogenesis of Infection: The Accidental Pathogen

As previously discussed, pathogenicity of most microbes is a contextual state. Owing to the adjuvant capacity of the microbiota, infections that occur at sites colonized by commensals can be, in some cases, considered as co-infections, with normal constituents of the microbiota being a major culprit of tissue damage and pathogen transmission. Although tissue-resident symbionts can provide an immunological and ecological shield against pathogen invasion, these microbes can in some instances be coopted by pathogens for optimal transmission. One of the first illustrations of the positive effect of the flora on pathogen development and survival was revealed in a model of Trichuris muris nematode infection (Hayes et al., 2010). In this model of infection, egg hatching in the large intestine only occurred upon tight contact with bacteria, suggesting that the microflora provide critical cues for the appropriate establishment of the life cycle of gut tropic nematodes (Hayes et al., 2010). This pro-infection role of the microbiota represents a novel paradigm for the transmission of various pathogens, including viruses. Poliovirus relies on the microbiota for efficient replication, an effect that can be at least in part associated with the capacity of the virus to bind to cardinal microbial products such as LPS (Kuss et al., 2011). Similarly, the capacity of MMTV to bind to commensally derived LPS favors mucosal transmission of the virus via the induction of the regulatory cytokine IL-10. Such effects lead to a state of immunological unresponsiveness toward viral antigen that favors transmission of the virus (Kane et al., 2011). In these studies, the elements accounting for the pathogen transmission and/or virulence-promoting effect of the flora are highly represented microbial products such as LPS, suggesting that both virus and nematodes may have evolved to bypass commensal population shifts by thriving on ubiquitous microbial derived components. This would suggest that, although manipulating the flora may represent an efficient way of altering immunity to pathogens, this strategy is unlikely to have global consequences on pathogen transmission. Based on the pleitropic effect of the flora in induction of regulatory pathways, host metabolism, and tissue-resident function, one would expect that a high number of pathogens transmitted via mucosa or using commensal-rich habitats have evolved to benefit from the complex interaction of the host with its microbiota.

Infections represent highly volatile situations for barrier tissues, with pathogens, commensals, and environmental antigens transiently sharing the same inflamed environment. In westernized countries, it is estimated that a child will suffer ten diarrheal episodes on average before the age of 5, a number that can be dramatically increased in the developing world (Kosek et al., 2003; Vernacchio et al., 2006). Thus, when added together with skin and lung infections, the immune system has ample opportunity to be exposed to commensals during inflammation, a situation that has the potential to disrupt our homeostatic relationship with these microbes. In the gastrointestinal tract, acute mucosal infections are characterized by dysbiosis associated with significant shifts in the microbiota and dominance of

bacteria, with enhanced invasive and inflammatory properties that can directly exacerbate inflammation and tissue damage (Egan et al., 2012; Heimesaat et al., 2006; Lupp et al., 2007; Stecher et al., 2007). In particular, γ -proteobacteria dominance has emerged as a hallmark of acute mucosal infections and enhanced pathology (Benson et al., 2009; Craven et al., 2012; Egan et al., 2012; Heimesaat et al., 2006; Lupp et al., 2007; Molloy et al., 2013; Raetz et al., 2013; Stecher et al., 2007). Various mechanisms could contribute to proteobacterial "blooms," including the capacity of these commensals to thrive on metabolites derived from inflammatory setting such as nitrates and benefit from the selective death of Paneth cells (Raetz et al., 2013; Winter et al., 2013). Because of the pathogenic role of these bacteria, the immune system may have evolved specific modes of control. Indeed, in the context of acute mucosal infection, both neutrophils and monocytes can exit the lamina propria and enter the gut lumen, thereby creating a containment structure referred to as intraluminal cast, which limits epithelial contact with these invasive microbes and translocation (Molloy et al., 2013).

In a number of models of gastrointestinal infections, such as Toxoplasma gondii and Yersinia pseudotuberculosis, immunopathology can also induce the translocation of commensal bacteria (Brenchley and Douek, 2012; Brenchley et al., 2006; Estes et al., 2010; Hand et al., 2012; Heimesaat et al., 2006; Jung et al., 2012; Meinzer et al., 2012). A consequence of infection that results from the induction of inflammation and enhanced microbial translocation is that, in contrast to steady-state responses, commensal-specific T cells, much like pathogen-specific cells, become activated and differentiate to an inflammatory phenotype (Hand et al., 2012). In the GI tract, these commensalspecific T cells form memory cells that are phenotypically and functionally indistinguishable from pathogen-specific T cells (Hand et al., 2012). Because of the extraordinary number of potential antigens expressed by the host microbiota, this implies that a significant fraction of memory cells are expected to be commensal specific and will develop over time in response to successive infections and/or various barrier breaches. Thus, primary exposure to a pathogen in the skin, lung, and GI tract is likely to occur in the context of a much broader recall response against commensal bacteria. Much like pathogen-specific CD4 T cells, commensal-specific memory T cells declined steadily over time (Hand et al., 2012; Homann et al., 2001; Pepper et al., 2010). As CD4 T cells carry out the complex task of remembering and responding to pathogenic and commensal organisms in a constantly changing environment, perhaps development of CD4 memory reflects this necessity for flexibility. An evolving pool of specificities within the regulatory and effector CD4 T cell compartment may allow for the maintenance of tolerance and barrier function in the face of variable commensal populations and intermittent infection.

The physiological consequence of long-term T cell memory against commensals remains to be addressed. One possible consequence may be the induction of heterologous memory, wherein antigen-specific responses against commensal bacteria could drive the rapid production of inflammatory cytokines upon secondary infection, leading to increased protection against both pathogens and commensal bacteria. In support of this hypothesis, healthy human serum contains antibodies that are specific to the skin and intestinal microbiota (Haas et al., 2011). In addition, recent studies suggest that CD4 T cell clones that are cross-reactive to commensals and viruses are common in healthy individuals (Su et al., 2013). Conversely, aberrant accumulation of commensal-specific T cells may lead to the development of inflammation and IBD (Sartor, 2006). A further exploration of antigen-specific memory T cells residing at all barrier sites and their regulation would inform us of the potential of impact of effector responses against commensals at steady state or in the context of inflammation.

Role of the Microbiota in Chronic Diseases

Optimal microbiota host interaction implies that balance between stimulatory and regulatory signals would allow the development of immunity without compromising the capacity of the host to maintain tolerance to innocuous antigens. Yet, in westernized countries, the overuse of antibiotics, changes in the diet, and elimination of chronic parasitic infections, such as those caused by helminths, including roundworms, hookworms, and whipworms, may have selected for a microbiota that lacks the resilience required for the establishment of balanced immune responses. For instance, as recently as 1940, the prevalence of helminthic worm infection in children in some rural areas of the United States was as high as 70% (Johnston et al., 2014; Weinstock et al., 2004). Further, the use of antibiotics and changes in diet have caused the chronic disappearance of potentially critical components of the human microbiota (Blaser and Falkow, 2009). These profound changes in the microbiota and, as a direct result, the immune system are now believed to contribute to the dramatic and rapid increase in chronic inflammatory and autoimmune disorders seen in high-income countries. Indeed, though each inflammatory disease is associated with unique genetic and biological mechanisms, a unifying trend seems to be that many inflammatory diseases are associated with significant shifts in the resident microbiota from a "healthy" to "diseased" state. One tantalizing hypothesis is that the partially penetrant nature of genetic polymorphisms associated with many complex diseases is due to the necessity of changes in the microbiota for the onset of pathology. An alarming consequence of this hypothesis is that susceptibility to some diseases is partly due to the stochastic acquisition of specific commensal organisms. Interestingly, similar "opportunists" are associated with multiple disease states. Therefore, some bacteria may be particularly adept at surviving in and contributing to inflammation, and the recent acquisition of a particular set of inflammatory clades of bacteria may be an important contributor to the etiology of inflammatory and autoimmune diseases. Below, we will discuss a number of diseases and their associations with the microbiota.

Role of the Microbiota in IBD and Autoimmune Disease

The gastrointestinal tract is home to the largest population of commensal organisms in the human body and, as such, is home to a unique set of immunoregulatory mechanisms that prevent the unnecessary activation of the immune system against innocuous antigens, including those expressed by the microbiota. A breakdown in these overlapping regulatory mechanisms results in a set of chronic inflammatory conditions that are collectively known as inflammatory bowel disease (IBD). The relationship between mucosal immune dysfunction and IBD is illustrated by the fact that both Crohn's disease (CD) and ulcerative colitis (UC) are associated with genes that are critical in maintenance of the epithelial barrier and the regulation of innate and adaptive immune responses (Rivas et al., 2011). The etiology of IBD is complex and is believed to be the consequence of genetic factors, the host immune system, and environmental factors, such as the microbiota (Maloy and Powrie, 2011). Stressors such as defined infections have also been proposed to contribute to the induction of these disorders (Cadwell et al., 2010).

IBD is associated with profound changes in the composition of the intestinal microbiota, highlighting the importance of the microbiota in disease etiology. Notably, numerous studies have shown that both CD and UC are associated with a reduced complexity of the commensal microbiota and consistent shifts in the consortia to a dysbiotic state. For example, in a similar manner to what is observed during acute mucosal infections, both CD and UC are characterized by the outgrowth of the phyla proteobacteria and, in particular, the Enterobacteriaceae family (Frank et al., 2007). Notably Crohn's disease has been associated with commensals that are intrinsically inflammatory and blur the line between commensal and pathogen. For example, adherent-invasive E. coli, Yersinia, and Clostridium difficile are much more common in patients suffering from Crohn's disease than in healthy individuals and in some mouse models have been shown to be key contributors to IBD (Issa et al., 2008; Lamps et al., 2003; Rolhion and Darfeuille-Michaud, 2007). In mice deficient in IL-10 or in IL-10 and TGF- β and therefore prone to develop colitis, defined commensals such as Helicobacter hepaticus or commensal Bacteroides are sufficient to induce disease (Bloom et al., 2011; Cahill et al., 1997; Kullberg et al., 2002). It is hypothesized that these commensal bacteria with enhanced inflammatory potential contribute to disease via their capacity to invade and promote innate and adaptive immune responses to otherwise benign food antigens and commensal organisms (Elson and Cong, 2012; Packey and Sartor, 2008). Indeed, increased serum antibody responses against antigens derived from the microbiota are characteristic of Crohn's disease patients (Dubinsky et al., 2006; Lodes et al., 2004). One simple explanation as to why particular commensal organisms are able to "bloom" during inflammation is that these organisms are closely related to obligate pathogens and therefore possess modules that allow for survival under the harsh conditions of immune activation. Indeed, commensal E. coli, which is benign under homeostatic conditions, is capable of using inflammatory nitric oxides generated by the immune response as an energy source, conferring a significant growth advantage (Winter et al., 2013). Other organisms such as Bilophila wadsworthia can take advantage of dietary-induced bile acids to dominate the intestine, contributing to disease in mouse strains prone to colitis, providing a glimpse at how diet may trigger IBD via its profound impact on the microbiota (Devkota et al., 2012). An intriguing hypothesis that can be derived from these studies is that IBD is caused or exacerbated by a positive feedback loop, where host mutation leads to dysregulated immune responses in the GI tract, which drive the outgrowth of inflammatory bacteria that in turn promote more inflammation. One striking experimental example of the influence of the flora on IBD is that of mice deficient in both T-bet and adaptive immunity that develop a spontaneous and transferrable form of ulcerative colitis (TRUC mice) (Garrett et al., 2007). Deficiency of T-bet in the innate immune system leads to exaggerated tumor necrosis factor a (TNF- α) production by dendritic cells, which together with the absence of T_{reg} , creates a chronic inflammatory state that modulates the composition of the microflora and eventually leads to the development of colorectal cancer (Garrett et al., 2007; Garrett et al., 2009). Interestingly, transfer of the microbiota from TRUC mice into wild-type recipients also transfers the colitis. Defined species of commensals such as Proteus mirabilis, Klebsiella pneumoniae, or Helicobacter typhlonius are found at increased frequency in TRUC mice and can transfer disease in wild-type mice (Garrett et al., 2010; Powell et al., 2012). Further demonstration that colitis can, at least in experimental settings, become "contagious" was revealed in a number of mouse models in which innate immune defects in components of the inflammasome or IL-22 led to aberrant responses to commensals (Elinav et al., 2011; Zenewicz et al., 2013). Deficiency of the NLRP6 component of the inflammasome from colonic cells results in reduced IL-18 levels and shifts in microbiota composition characterized by expansion of the bacterial phyla Bacteroidetes (in particular, the family Prevotellaceae) and TM7. Consequent to the microbial shift, these mice spontaneously develop intestinal hyperplasia and are more susceptible to chemically induced colitis and colonic cancer (Elinav et al., 2011; Hu et al., 2013). As for the TRUC mice, the colitic phenotype can be transferred by the microbiota to neonatal or adult wild-type mice that have no innate immune defect, though it should be noted that colitis is less severe in these animals (Garrett et al., 2007). How these infectious and dysbiotic microbiota contribute to human disorders remains to be addressed but is of critical importance if we are to understand the recent rise of IBD incidence.

Perhaps equally important to the outgrowth of inflammatory components of the microbiota in the induction of IBD is the loss of symbiotic commensal organisms that rely upon fermentation as their energy source. Sequencing studies have shown that the bacteria of the phylum Firmicutes and, in particular, the class Clostridia are reduced in patients suffering IBD (Frank et al., 2007). This is potentially important because of the effect that bacteria within this class can have on GI-resident T_{reg} cells. Clostridia has been shown to directly induce colonic T_{reas} that can oppose colitis induction (Atarashi et al., 2011, 2013). Additionally and as previously discussed, single-chain fatty acids (SCFA) have been shown to limit GI inflammation both by the induction of T_{req} and by the direct inhibition of macrophage and neutrophil activation (Arpaia et al., 2013; Furusawa et al., 2013; Maslowski et al., 2009; Smith et al., 2013). The enzymatic processes necessary for the fermentation of fiber to SCFA are largely dependent upon bacteria within the Clostridia class (Pryde et al., 2002). Thus, the mucosa-associated immune system is shaped by commensal dysbiosis in at least two distinct ways. First, the outgrowth of opportunistic clades of bacteria drives increased inflammation. And second, the loss of benign fermenting bacteria that produce "keystone" metabolites results in reduced immunoregulation.

The microbiome may also contribute to autoimmune disease at other barrier and systemic sites. Though it was initially thought to be largely sterile, the resident microbiota of the respiratory tract has recently been described (Beck et al., 2012) and is a likely contributor to the etiology of asthma and other airway inflammatory disorders, such as cystic fibrosis (CF). Cystic fibrosis is caused by inactivating mutations in the cystic fibrosis transmembrane conductance regulator (CFTR) gene that is required for the homeostatic control of chloride ions in the lung. The expression of mutated CFTR proteins leads to the build-up of thick mucus on the surface of the airway that cannot be easily removed by the action of the cilia. The build-up of mucus on the surface of the lung allows inflammatory constituents of the lung microbiota to gain a foothold and grow out, leading to an influx of neutrophils and macrophages and immune-mediated damage to the airway that severely limits pulmonary function. The bacteria that are most commonly associated with the disease, Pseudomonas and Staphylococcus, are often constituents of the healthy lung microbiota (Blainey et al., 2012), and therefore CF is perhaps the simplest example of a genetic disease whose mechanism is the induction of dysbiosis within the microbiota and subsequent activation of the immune system, leading to uncontrolled inflammation.

Given the microbiota's role in setting the systemic immune tone, a number of groups have investigated the role of the commensal microbiota in animal models of autoimmune disease. Rheumatoid arthritis is an autoimmune disease that is characterized by the invasion of joints by immune cells and, in particular, immune complexes formed from autoreactive antibodies. In a murine model of rheumatoid arthritis, disease is significantly reduced in the absence of commensal bacteria. with diminished disease associated with a reduction in the proinflammatory Th17 response (Wu et al., 2010). Arthritic phenotypes were completely restored when germ-free animals were monocolonized with SFB, which drives the differentiation and/ or dissemination of both intestinal and splenic Th17 cells (Wu et al., 2010). Analysis of the microbiome of patients suffering RA revealed a surprising relationship between the disease and a potentially inflammatory clade of Prevotellaceae, but how the microbiota contribute to immune activation and dissemination of T cells and antibodies to the joints in RA is unknown (Scher et al., 2013).

Multiple sclerosis is an autoimmune disease characterized by the invasion of the central nervous system by immune cells, resulting in the demyelination of neurons and subsequent pathology. The incidence of disease in the mouse model of multiple sclerosis, experimental autoimmune encephalitis (EAE), is also dependent upon the commensal microbiota (Berer et al., 2011; Lee et al., 2011; Ochoa-Repáraz et al., 2009, 2010a, 2010b). These data are in agreement with what had been known anecdotally for many years-that disease varied among genetically identical animals housed in different settings. Multiple studies have now shown that the activation of myelin-specific B cells and the invasion of the neurons by effector T cells require the commensal microbiota (Berer et al., 2011; Ochoa-Repáraz et al., 2009). It should be noted that the role of the microbiota in inducing EAE is contextual. For example, the commensal microbiota may be protective in this disease, as PSA of B. fragilis can ameliorate symptoms via the induction of IL-10-producing regulatory T cells (Lee et al., 2011).

How the microbiota is acting to drive disease in these models is not completely understood. It does seem likely that, in individuals prone to autoimmunity, the intrinsic inflammatory nature of defined commensal bacteria could lead to an increase in the systemic threshold of activation. Further, the average microbiota possesses millions of potential antigens, and as recently highlighted, the possibility of dangerous cross-reactivity between commensals, pathogens, and self is higher than previously understood (Su et al., 2013).

As alluded to previously, the commensal microbiota can provide protection against some inflammatory disorders. For instance, incidence of disease in the nonobese diabetic (NOD) mouse model of type 1 diabetes (T1D) is maximal in germ-free animals, and the presence of SFB within the microbiota segregates with protection from the disease (Kriegel et al., 2011; Pozzilli et al., 1993). Moreover, MyD88 knockout mice bred to the pro-diabetic background are also protected from disease due to the possession of a "protective" microbiota that can be transmitted from mother to child (Wen et al., 2008). The mouse model of diabetes, but not the human disease, has a significant preference for females, which is in concert with most other autoimmune disorders. Interestingly, this sexual dimorphism is also dependent upon the microbiota, as it is not seen in germ-free mice, and the transfer of a male microbiota to females equalizes the incidence of the disease as a result of changes in the expression of sex hormones (Markle et al., 2013; Yurkovetskiy et al., 2013). How the microbiota shapes the immune system and which bacterial clades are important in T1D are still not known, but this research may lead to an understanding of factors contributing to the initiation of T1D, a disease that has been increasing in prevalence worldwide without a known stressor. Additionally, how the microbiota may shape host physiology, such as sex hormones, resulting in modification of the immune response adds an additional layer of complexity that will be an important component of future studies.

Role of the Microbiota in Cancer

Perhaps the first indication that the commensal microbiota could drive cancer was provided by the observation that stomach ulcers and subsequent stomach cancer were caused by the presence of a single type of bacteria, *Helicobacter pylori* (Marshall and Warren, 1984). The vast majority of people carrying *H. pylori* are asymptomatic, and thus, as for most host-microbe interactions, the host response to the colonization determines whether chronic inflammation and carcinogenesis occurs. Similar phenomena seem to be important to the etiology of cancers at other barrier surfaces—most notably, the intestine. On the other hand, *H. pylori* is associated with protection from other types of cancer (Atherton and Blaser, 2009), further highlighting the functional contextuality of commensal.

Intestinal cancer is inextricably linked to inflammation in the GI tract, and commensal-driven immune activation is an important contributing factor to the etiology of these diseases (Grivennikov and Karin, 2010). Notably, IBD and, in particular, CD are associated with the development of colon cancer (Elinav et al., 2013). Liver cancer has also been linked to the microbiota via induction

of specific bile acids that damage DNA within the liver (Yoshimoto et al., 2013). A number of studies have attempted to link signaling via innate immune receptors in the GI tract and disease in mouse models of colorectal cancer; however, the results have been disparate, perhaps due to the confounding factor of the microbiota (Elinav et al., 2013; Fukata et al., 2007; Lowe et al., 2010; Rakoff-Nahoum and Medzhitov, 2007; Salcedo et al., 2010). Indeed, dysbiosis of the microbiota seems to be an important factor in determining whether the immune system contributes or protects against the initiation of cancer. For example, in the NLRP6-deficient mice that lack a key component of the inflammasome and TRUC mice, susceptibility to colorectal cancer is associated with dysbiosis, which is directly related to immune dysfunction (Garrett et al., 2009; Hu et al., 2013). Moreover, as a direct illustration of the importance of the microbiota in colon cancer, disease susceptibility can be transferred to healthy mice via the commensal microbiota (Garrett et al., 2009; Hu et al., 2013). The microbiome of human patients suffering from colorectal cancer is also modestly disrupted, with an increased representation of Prevotella (Sobhani et al., 2011). Additionally, mouse models of colorectal cancer have been shown to be exacerbated by specific strains of bacteria. For example, E. coli that express the polyketide synthase genotoxic island can promote tumorigenesis in defined models of colon cancer, and E. coli that express the same gene module are found in patients afflicted with colon cancer (Arthur et al., 2012). Bacteria of the phylum Fusobacteria are also found at a high frequency residing within the tumor tissue and are important in inducing the traffic of tumor-promoting myeloid cells (Kostic et al., 2012, 2013; Rubinstein et al., 2013). In general, commensal-driven infiltration of intestinal tumors by immune cells is believed to be an important driving factor in tumorigenesis. Studies have shown that colonic tumors have a reduced mucus layer and are susceptible to bacterial translocation and the influx of Th17 cells (Grivennikov et al., 2012). Related to this, in the multiple intestinal neoplasia mouse model, chronic colonization with the Th17inducing bacteria enterotoxigenic Bacteroides fragilis leads to increased tumorigenesis (Wu et al., 2009). Therefore, colon cancer, much like IBD and stomach cancer, may be caused or exacerbated by an over-exuberant immune response against a dysbiotic commensal microbiota. However, it should be noted that commensal-induced inflammation is a "double-edged sword" for the host. As previously discussed, the microbiota is also an integral part of the capacity of the host to control tumor in the context of immunotherapy and chemotherapy. Thus, the role of the microbiota in tumor development and control is highly contextual and very much dependent upon the exact constituents of the microbiota and the state of the host immune system.

Role of the Microbiota in Type II Diabetes and Metabolic Syndrome

Metabolic syndrome is a set of risk factors for heart disease, stroke, and diabetes that are related to increased adiposity and sedentary lifestyles. Type II diabetes (T2D) is an inflammatory condition in which increased adiposity and metabolic defects drive chronic production of inflammatory cytokines such as TNF- α and IL-1 (Gregor and Hotamisligil, 2011). These inflammatory cytokines inhibit insulin signaling, leading to insulin resis-

tance and increased blood sugar (Osborn and Olefsky, 2012). The microbiota has been proposed to play a prominent role in this process. Indeed, the commensal microbiota of obese individuals is significantly altered such that the ratio of firmicutes is increased in relation to bacteroidetes, a shift that is associated with increased nutrient production for the host (Turnbaugh et al., 2006). Importantly, these changes are transferrable, as the microbiota of genetically obese mice leads to increased adiposity among healthy cage mates (Turnbaugh et al., 2006). Significant changes to the commensal microbiota have been observed in T2D as well and, interestingly, are associated with a reduction in SCFA-producing strains of Clostridia and increases in E. coli (Karlsson et al., 2013; Qin et al., 2012). Serum LPS levels are also increased as a result of T2D, indicating that increased translocation of bacterial products may be taking place; in mouse models, high-fat diets have been associated with increased translocation of commensal E. coli (Amar et al., 2011; Jayashree et al., 2013). Finally, there is some evidence that susceptibility to obesity itself is regulated by the immune system. Mice that are deficient in TLR5 develop exacerbated metabolic syndrome and grow obese faster than wild-type controls (Vijay-Kumar et al., 2010). The transfer of the microbiota alone is sufficient to transfer the susceptibility to metabolic syndrome, and tellingly, these mice are particularly susceptible to the outgrowth of E. coli (Carvalho et al., 2012; Vijay-Kumar et al., 2010). Therefore, in T2D as in IBD, the disease can be exacerbated by complex interactions between the immune system and the microbiota, leading to chronic inflammation.

Interdependence of Diet, Commensals, and the Immune System

The immune system is not only controlled by its symbiotic relationship with the microbiota but is also exquisitely sensitive to the nutritional status of the host. Evidence now exists for a multidirectional interaction between the diet, immune system, and commensal microflora. The intestine serves as the primary site of nutrient absorption in the body. As such, the immune system and commensal microbiota are sensitive to changes in diet. Dietary control of immune cells is mediated both by metabolic requirements and direct sensing of food-derived metabolites. For instance, metabolite of vitamin A, retinoic acid, which is highly enriched in the gut, can directly control the capacity of lymphocytes to respond to antigen and to migrate to the gastrointestinal tract (Hall et al., 2011a, 2011b). In addition, dietary-derived AHR ligands from cruciferous vegetables are important signals for intestinal immune development and immune responses (Kiss et al., 2011; Li et al., 2011). The concept that defined metabolites could control unique aspects of the immune system is believed to be a highly generalizable observation extending to all micronutrients explored thus far, including iron, folate, zinc, selenium, and vitamins C, D, and E (Mora et al., 2008; Savy et al., 2009; von Essen et al., 2010). However, the diet has a profound and highly dynamic impact on the composition and function of the microbiota. Therefore, an understanding of the role of diet in immune responses requires integration of the commensal microbiota as well (Figure 4). As an example, mice fed a high-fat diet have altered commensal communities that not only effect energy harvest but that can also alter immunity and increase the severity

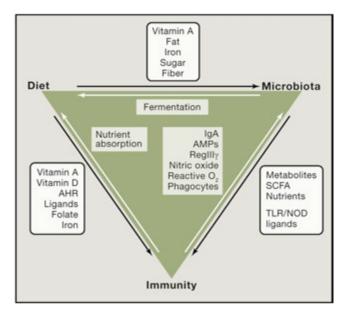


Figure 4. Interdependence of Diet, Immune, and Microbiota Interactions

Evidence now exists for bidirectional communication between the three key factors in the GI tract: diet, immune system, and commensal microflora. Diet can have a profound influence on the immune system (e.g., vitamin A, vitamin D, AHR), and the immune system can also affect nutrient uptake. Diet also has dominant influence on the composition and metabolic capacity of commensal bacteria, while this, in return, influences nutrient absorption. The immune system is able to exert control over both commensal composition and localization, whereas commensal signals are critical for development and function of the immune system.

of disease states such as colitis (Devkota et al., 2012; Gregor and Hotamisligil, 2011; Ley et al., 2006b; Monira et al., 2011). Defined dietary deficiency can also alter commensal communities. For example, the essential micronutrient vitamin A, which is required for the induction of protective immunity, has also profound effects on the composition of the microbiota (Cha et al., 2010). Recent findings have also shown that the immune system may also be able to directly affect dietary intake. For instance, mice lacking B cells have deficiencies in dietary absorption (Shulzhenko et al., 2011). Conversely and as mentioned, mice that are deficient in the receptor for bacterial flagellin TLR5 have a dysbiotic commensal microflora that leads to increased weight gain and the development of metabolic syndrome (Vijay-Kumar et al., 2010). Thus, immunodeficiency can alter the gut flora's composition and thereby the metabolic capacity of both the flora and the host. This idea may have important consequences for human health not only in the developed world, where metabolic syndrome is epidemic, but also in the developing world. Children with severe malnutrition represent an example of both immunodeficiency and dietary deficiency coinciding in a clinically relevant way. Malnourished children have dramatically altered commensal communities, likely with diminished metabolic capacity and impaired ability to promote host energy absorption-a problem that can persist even after dietary intervention (Gordon et al., 2012; Monira et al., 2011; Yatsunenko et al., 2012).

Fundamental changes in cultural traditions and socioeconomic status have affected the nutritional status of humans worldwide, altering microbial communities in unpredictable and possibly harmful ways (Lozupone et al., 2012). Considering the interdependency of the diet, immune system, and microbiota, an important challenge over the next few years will be to develop experimental strategies aimed at defining critical dietary regulators of this complex and interdependent network.

Concluding Remarks

Autoimmune and inflammatory diseases, all associated with dysregulated immune responses, have been rising dramatically over the past few decades. The field of immunology is at a fascinating crossroad. In recent years, immunological research has evolved from a lymphoid-tissue-centric view of the immune system to the understanding of tissue microenvironments as a fundamental determinant of immune responses. This area of research has led to the integration of the microbiota as an intrinsic regulator of all immune responses. There is today an explosion of discoveries associated with the role of communities of microbes, keystone bacterial species, commensalderived products, or metabolites and disease states in humans. This provides scientists and clinicians with a unique opportunity to develop an integrated exploration of human health that includes ecologists, nutritionists, geneticists, microbiologists, biochemists, and immunologists. In this multidisciplinary area of research lies the key to fundamental discoveries aimed at manipulating or restoring defined aspects of the immune system-microbiota dialog in order to promote or restore the health of the human meta-organism.

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REFERENCES

Abreu, N.A., Nagalingam, N.A., Song, Y., Roediger, F.C., Pletcher, S.D., Goldberg, A.N., and Lynch, S.V. (2012). Sinus microbiome diversity depletion and Corynebacterium tuberculostearicum enrichment mediates rhinosinusitis. Sci. Transl. Med. *4*, 151ra124.

Abt, M.C., Osborne, L.C., Monticelli, L.A., Doering, T.A., Alenghat, T., Sonnenberg, G.F., Paley, M.A., Antenus, M., Williams, K.L., Erikson, J., et al. (2012). Commensal bacteria calibrate the activation threshold of innate antiviral immunity. Immunity *37*, 158–170.

Alenghat, T., Osborne, L.C., Saenz, S.A., Kobuley, D., Ziegler, C.G., Mullican, S.E., Choi, I., Grunberg, S., Sinha, R., Wynosky-Dolfi, M., et al. (2013). Histone deacetylase 3 coordinates commensal-bacteria-dependent intestinal homeostasis. Nature *504*, 153–157.

Amar, J., Serino, M., Lange, C., Chabo, C., Iacovoni, J., Mondot, S., Lepage, P., Klopp, C., Mariette, J., Bouchez, O., et al.; D.E.S.I.R. Study Group (2011). Involvement of tissue bacteria in the onset of diabetes in humans: evidence for a concept. Diabetologia *54*, 3055–3061.

An, D., Oh, S.F., Olszak, T., Neves, J.F., Avci, F.Y., Erturk-Hasdemir, D., Lu, X., Zeissig, S., Blumberg, R.S., and Kasper, D.L. (2014). Sphingolipids from a

symbiotic microbe regulate homeostasis of host intestinal natural killer T cells. Cell *156*, 123–133.

Arpaia, N., Campbell, C., Fan, X., Dikiy, S., van der Veeken, J., deRoos, P., Liu, H., Cross, J.R., Pfeffer, K., Coffer, P.J., and Rudensky, A.Y. (2013). Metabolites produced by commensal bacteria promote peripheral regulatory T-cell generation. Nature *504*, 451–455.

Arthur, J.C., Perez-Chanona, E., Mühlbauer, M., Tomkovich, S., Uronis, J.M., Fan, T.J., Campbell, B.J., Abujamel, T., Dogan, B., Rogers, A.B., et al. (2012). Intestinal inflammation targets cancer-inducing activity of the microbiota. Science 338, 120–123.

Atarashi, K., Tanoue, T., Shima, T., Imaoka, A., Kuwahara, T., Momose, Y., Cheng, G., Yamasaki, S., Saito, T., Ohba, Y., et al. (2011). Induction of colonic regulatory T cells by indigenous Clostridium species. Science 331, 337–341.

Atarashi, K., Tanoue, T., Oshima, K., Suda, W., Nagano, Y., Nishikawa, H., Fukuda, S., Saito, T., Narushima, S., Hase, K., et al. (2013). Treg induction by a rationally selected mixture of Clostridia strains from the human microbiota. Nature *500*, 232–236.

Atherton, J.C., and Blaser, M.J. (2009). Coadaptation of Helicobacter pylori and humans: ancient history, modern implications. J. Clin. Invest. *119*, 2475–2487.

Avila, M., Ojcius, D.M., and Yilmaz, O. (2009). The oral microbiota: living with a permanent guest. DNA Cell Biol. *28*, 405–411.

Bauer, H., Horowitz, R.E., Levenson, S.M., and Popper, H. (1963). The response of the lymphatic tissue to the microbial flora. Studies on germfree mice. Am. J. Pathol. *42*, 471–483.

Beck, J.M., Young, V.B., and Huffnagle, G.B. (2012). The microbiome of the lung. Transl. Res. *160*, 258–266.

Belda-Ferre, P., Alcaraz, L.D., Cabrera-Rubio, R., Romero, H., Simón-Soro, A., Pignatelli, M., and Mira, A. (2012). The oral metagenome in health and disease. ISME J. 6, 46–56.

Belkaid, Y., and Naik, S. (2013). Compartmentalized and systemic control of tissue immunity by commensals. Nat. Immunol. *14*, 646–653.

Benson, A., Pifer, R., Behrendt, C.L., Hooper, L.V., and Yarovinsky, F. (2009). Gut commensal bacteria direct a protective immune response against Toxoplasma gondii. Cell Host Microbe *6*, 187–196.

Berer, K., Mues, M., Koutrolos, M., Rasbi, Z.A., Boziki, M., Johner, C., Wekerle, H., and Krishnamoorthy, G. (2011). Commensal microbiota and myelin autoantigen cooperate to trigger autoimmune demyelination. Nature 479, 538–541.

Blainey, P.C., Milla, C.E., Cornfield, D.N., and Quake, S.R. (2012). Quantitative analysis of the human airway microbial ecology reveals a pervasive signature for cystic fibrosis. Sci. Transl. Med. *4*, 153ra130.

Blaser, M.J., and Falkow, S. (2009). What are the consequences of the disappearing human microbiota? Nat. Rev. Microbiol. 7, 887–894.

Bloom, S.M., Bijanki, V.N., Nava, G.M., Sun, L., Malvin, N.P., Donermeyer, D.L., Dunne, W.M., Jr., Allen, P.M., and Stappenbeck, T.S. (2011). Commensal Bacteroides species induce colitis in host-genotype-specific fashion in a mouse model of inflammatory bowel disease. Cell Host Microbe *9*, 390–403.

Bouladoux, N., Hall, J.A., Grainger, J.R., dos Santos, L.M., Kann, M.G., Nagarajan, V., Verthelyi, D., and Belkaid, Y. (2012). Regulatory role of suppressive motifs from commensal DNA. Mucosal Immunol. *5*, 623–634.

Boullier, S., Tanguy, M., Kadaoui, K.A., Caubet, C., Sansonetti, P., Corthésy, B., and Phalipon, A. (2009). Secretory IgA-mediated neutralization of Shigella flexneri prevents intestinal tissue destruction by down-regulating inflammatory circuits. J. Immunol. *183*, 5879–5885.

Bouskra, D., Brézillon, C., Bérard, M., Werts, C., Varona, R., Boneca, I.G., and Eberl, G. (2008). Lymphoid tissue genesis induced by commensals through NOD1 regulates intestinal homeostasis. Nature 456, 507–510.

Brandl, K., Plitas, G., Schnabl, B., DeMatteo, R.P., and Pamer, E.G. (2007). MyD88-mediated signals induce the bactericidal lectin RegIII gamma and protect mice against intestinal Listeria monocytogenes infection. J. Exp. Med. *204*, 1891–1900. Brandl, K., Plitas, G., Mihu, C.N., Ubeda, C., Jia, T., Fleisher, M., Schnabl, B., DeMatteo, R.P., and Pamer, E.G. (2008). Vancomycin-resistant enterococci exploit antibiotic-induced innate immune deficits. Nature *455*, 804–807.

Brenchley, J.M., and Douek, D.C. (2012). Microbial translocation across the GI tract. Annu. Rev. Immunol. *30*, 149–173.

Brenchley, J.M., Price, D.A., Schacker, T.W., Asher, T.E., Silvestri, G., Rao, S., Kazzaz, Z., Bornstein, E., Lambotte, O., Altmann, D., et al. (2006). Microbial translocation is a cause of systemic immune activation in chronic HIV infection. Nat. Med. *12*, 1365–1371.

Buffie, C.G., and Pamer, E.G. (2013). Microbiota-mediated colonization resistance against intestinal pathogens. Nat. Rev. Immunol. *13*, 790–801.

Cadwell, K., Patel, K.K., Maloney, N.S., Liu, T.C., Ng, A.C., Storer, C.E., Head, R.D., Xavier, R., Stappenbeck, T.S., and Virgin, H.W. (2010). Virus-plussusceptibility gene interaction determines Crohn's disease gene Atg16L1 phenotypes in intestine. Cell *141*, 1135–1145.

Cahill, R.J., Foltz, C.J., Fox, J.G., Dangler, C.A., Powrie, F., and Schauer, D.B. (1997). Inflammatory bowel disease: an immunity-mediated condition triggered by bacterial infection with Helicobacter hepaticus. Infect. Immun. *65*, 3126–3131.

Carvalho, F.A., Koren, O., Goodrich, J.K., Johansson, M.E., Nalbantoglu, I., Aitken, J.D., Su, Y., Chassaing, B., Walters, W.A., González, A., et al. (2012). Transient inability to manage proteobacteria promotes chronic gut inflammation in TLR5-deficient mice. Cell Host Microbe *12*, 139–152.

Cash, H.L., Whitham, C.V., Behrendt, C.L., and Hooper, L.V. (2006). Symbiotic bacteria direct expression of an intestinal bactericidal lectin. Science *313*, 1126–1130.

Cebra, J.J. (1999). Influences of microbiota on intestinal immune system development. Am. J. Clin. Nutr. 69, 1046S–1051S.

Cebula, A., Seweryn, M., Rempala, G.A., Pabla, S.S., McIndoe, R.A., Denning, T.L., Bry, L., Kraj, P., Kisielow, P., and Ignatowicz, L. (2013). Thymus-derived regulatory T cells contribute to tolerance to commensal microbiota. Nature 497, 258–262.

Cha, H.R., Chang, S.Y., Chang, J.H., Kim, J.O., Yang, J.Y., Kim, C.H., and Kweon, M.N. (2010). Downregulation of Th17 cells in the small intestine by disruption of gut flora in the absence of retinoic acid. J. Immunol. *184*, 6799–6806.

Chassin, C., Kocur, M., Pott, J., Duerr, C.U., Gütle, D., Lotz, M., and Hornef, M.W. (2010). miR-146a mediates protective innate immune tolerance in the neonate intestine. Cell Host Microbe *8*, 358–368.

Chieppa, M., Rescigno, M., Huang, A.Y., and Germain, R.N. (2006). Dynamic imaging of dendritic cell extension into the small bowel lumen in response to epithelial cell TLR engagement. J. Exp. Med. *203*, 2841–2852.

Clarke, T.B., Davis, K.M., Lysenko, E.S., Zhou, A.Y., Yu, Y., and Weiser, J.N. (2010). Recognition of peptidoglycan from the microbiota by Nod1 enhances systemic innate immunity. Nat. Med. *16*, 228–231.

Cong, Y., Feng, T., Fujihashi, K., Schoeb, T.R., and Elson, C.O. (2009). A dominant, coordinated T regulatory cell-IgA response to the intestinal microbiota. Proc. Natl. Acad. Sci. USA *106*, 19256–19261.

Coombes, J.L., Siddiqui, K.R., Arancibia-Carcamo, C.V., Hall, J., Sun, C.M., Belkaid, Y., and Powrie, F. (2007). A functionally specialized population of mucosal CD103+ DCs induces Foxp3+ regulatory T cells via a TGF-beta and retinoic acid-dependent mechanism. J. Exp. Med. *204*, 1757–1764.

Costello, E.K., Lauber, C.L., Hamady, M., Fierer, N., Gordon, J.I., and Knight, R. (2009). Bacterial community variation in human body habitats across space and time. Science *326*, 1694–1697.

Craven, M., Egan, C.E., Dowd, S.E., McDonough, S.P., Dogan, B., Denkers, E.Y., Bowman, D., Scherl, E.J., and Simpson, K.W. (2012). Inflammation drives dysbiosis and bacterial invasion in murine models of ileal Crohn's disease. PLoS ONE 7, e41594.

Cummings, J.H., Pomare, E.W., Branch, W.J., Naylor, C.P., and Macfarlane, G.T. (1987). Short chain fatty acids in human large intestine, portal, hepatic and venous blood. Gut *28*, 1221–1227.

Davie, J.R. (2003). Inhibition of histone deacetylase activity by butyrate. J. Nutr. Suppl. 133, 2485S-2493S.

Devkota, S., Wang, Y., Musch, M.W., Leone, V., Fehlner-Peach, H., Nadimpalli, A., Antonopoulos, D.A., Jabri, B., and Chang, E.B. (2012). Dietary-fatinduced taurocholic acid promotes pathobiont expansion and colitis in II10-/- mice. Nature 487, 104–108.

Di Giacinto, C., Marinaro, M., Sanchez, M., Strober, W., and Boirivant, M. (2005). Probiotics ameliorate recurrent Th1-mediated murine colitis by inducing IL-10 and IL-10-dependent TGF-beta-bearing regulatory cells. J. Immunol. *174*, 3237–3246.

Dixon, D.R., Reife, R.A., Cebra, J.J., and Darveau, R.P. (2004). Commensal bacteria influence innate status within gingival tissues: a pilot study. J. Periodontol. 75, 1486–1492.

Döderlein, A. (1892). Das Scheidensekret und seine Bedeutung für das Puerperalfieber (Leipzig: E. Basold).

Dubinsky, M.C., Lin, Y.C., Dutridge, D., Picornell, Y., Landers, C.J., Farrior, S., Wrobel, I., Quiros, A., Vasiliauskas, E.A., Grill, B., et al.; Western Regional Pediatric IBD Research Alliance (2006). Serum immune responses predict rapid disease progression among children with Crohn's disease: immune responses predict disease progression. Am. J. Gastroenterol. *101*, 360–367.

Egan, C.E., Cohen, S.B., and Denkers, E.Y. (2012). Insights into inflammatory bowel disease using Toxoplasma gondii as an infectious trigger. Immunol. Cell Biol. *90*, 668–675.

Ege, M.J., Mayer, M., Normand, A.C., Genuneit, J., Cookson, W.O., Braun-Fahrländer, C., Heederik, D., Piarroux, R., and von Mutius, E.; GABRIELA Transregio 22 Study Group (2011). Exposure to environmental microorganisms and childhood asthma. N. Engl. J. Med. *364*, 701–709.

Elahi, S., Ertelt, J.M., Kinder, J.M., Jiang, T.T., Zhang, X., Xin, L., Chaturvedi, V., Strong, B.S., Qualls, J.E., Steinbrecher, K.A., et al. (2013). Immunosuppressive CD71+ erythroid cells compromise neonatal host defence against infection. Nature *504*, 158–162.

Elinav, E., Strowig, T., Kau, A.L., Henao-Mejia, J., Thaiss, C.A., Booth, C.J., Peaper, D.R., Bertin, J., Eisenbarth, S.C., Gordon, J.I., and Flavell, R.A. (2011). NLRP6 inflammasome regulates colonic microbial ecology and risk for colitis. Cell *145*, 745–757.

Elinav, E., Nowarski, R., Thaiss, C.A., Hu, B., Jin, C., and Flavell, R.A. (2013). Inflammation-induced cancer: crosstalk between tumours, immune cells and microorganisms. Nat. Rev. Cancer *13*, 759–771.

Elson, C.O., and Cong, Y. (2012). Host-microbiota interactions in inflammatory bowel disease. Gut Microbes 3, 332–344.

Ergin, A., Syrbe, U., Scheer, R., Thiel, A., Adam, T., Büssow, K., Duchmann, R., Zeitz, M., and Sieper, J. (2011). Impaired peripheral Th1 CD4+ T cell response to Escherichia coli proteins in patients with Crohn's disease and ankylosing spondylitis. J. Clin. Immunol. *31*, 998–1009.

Estes, J.D., Harris, L.D., Klatt, N.R., Tabb, B., Pittaluga, S., Paiardini, M., Barclay, G.R., Smedley, J., Pung, R., Oliveira, K.M., et al. (2010). Damaged intestinal epithelial integrity linked to microbial translocation in pathogenic simian immunodeficiency virus infections. PLoS Pathog. 6, e1001052.

Faith, J.J., Ahern, P.P., Ridaura, V.K., Cheng, J., and Gordon, J.I. (2014). Identifying gut microbe-host phenotype relationships using combinatorial communities in gnotobiotic mice. Sci. Transl. Med. *6*, 220ra211.

Feleszko, W., Jaworska, J., Rha, R.D., Steinhausen, S., Avagyan, A., Jaudszus, A., Ahrens, B., Groneberg, D.A., Wahn, U., and Hamelmann, E. (2007). Probiotic-induced suppression of allergic sensitization and airway inflammation is associated with an increase of T regulatory-dependent mechanisms in a murine model of asthma. Clin. Exp. Allergy *37*, 498–505.

Fernandez, M.I., Pedron, T., Tournebize, R., Olivo-Marin, J.C., Sansonetti, P.J., and Phalipon, A. (2003). Anti-inflammatory role for intracellular dimeric immunoglobulin a by neutralization of lipopolysaccharide in epithelial cells. Immunity *18*, 739–749.

Foligne, B., Zoumpopoulou, G., Dewulf, J., Ben Younes, A., Chareyre, F., Sirard, J.C., Pot, B., and Grangette, C. (2007). A key role of dendritic cells in probiotic functionality. PLoS ONE *2*, e313.

Franchi, L., Kamada, N., Nakamura, Y., Burberry, A., Kuffa, P., Suzuki, S., Shaw, M.H., Kim, Y.G., and Núñez, G. (2012). NLRC4-driven production of IL-1β discriminates between pathogenic and commensal bacteria and promotes host intestinal defense. Nat. Immunol. *13*, 449–456.

Frank, D.N., St Amand, A.L., Feldman, R.A., Boedeker, E.C., Harpaz, N., and Pace, N.R. (2007). Molecular-phylogenetic characterization of microbial community imbalances in human inflammatory bowel diseases. Proc. Natl. Acad. Sci. USA *104*, 13780–13785.

Fukata, M., Chen, A., Vamadevan, A.S., Cohen, J., Breglio, K., Krishnareddy, S., Hsu, D., Xu, R., Harpaz, N., Dannenberg, A.J., et al. (2007). Toll-like receptor-4 promotes the development of colitis-associated colorectal tumors. Gastroenterology *133*, 1869–1881.

Furusawa, Y., Obata, Y., Fukuda, S., Endo, T.A., Nakato, G., Takahashi, D., Nakanishi, Y., Uetake, C., Kato, K., Kato, T., et al. (2013). Commensal microbe-derived butyrate induces the differentiation of colonic regulatory T cells. Nature *504*, 446–450.

Gaboriau-Routhiau, V., Rakotobe, S., Lécuyer, E., Mulder, I., Lan, A., Bridonneau, C., Rochet, V., Pisi, A., De Paepe, M., Brandi, G., et al. (2009). The key role of segmented filamentous bacteria in the coordinated maturation of gut helper T cell responses. Immunity *31*, 677–689.

Gabryszewski, S.J., Bachar, O., Dyer, K.D., Percopo, C.M., Killoran, K.E., Domachowske, J.B., and Rosenberg, H.F. (2011). Lactobacillus-mediated priming of the respiratory mucosa protects against lethal pneumovirus infection. J. Immunol. *186*, 1151–1161.

Gantois, I., Ducatelle, R., Pasmans, F., Haesebrouck, F., Hautefort, I., Thompson, A., Hinton, J.C., and Van Immerseel, F. (2006). Butyrate specifically down-regulates salmonella pathogenicity island 1 gene expression. Appl. Environ. Microbiol. *72*, 946–949.

Garcia-Crespo, K.E., Chan, C.C., Gabryszewski, S.J., Percopo, C.M., Rigaux, P., Dyer, K.D., Domachowske, J.B., and Rosenberg, H.F. (2012). Lactobacillus priming of the respiratory tract: Heterologous immunity and protection against lethal pneumovirus infection. Antiviral Res. *97*, 270–279.

Garrett, W.S., Lord, G.M., Punit, S., Lugo-Villarino, G., Mazmanian, S.K., Ito, S., Glickman, J.N., and Glimcher, L.H. (2007). Communicable ulcerative colitis induced by T-bet deficiency in the innate immune system. Cell *131*, 33–45.

Garrett, W.S., Punit, S., Gallini, C.A., Michaud, M., Zhang, D., Sigrist, K.S., Lord, G.M., Glickman, J.N., and Glimcher, L.H. (2009). Colitis-associated colorectal cancer driven by T-bet deficiency in dendritic cells. Cancer Cell *16*, 208–219.

Garrett, W.S., Gallini, C.A., Yatsunenko, T., Michaud, M., DuBois, A., Delaney, M.L., Punit, S., Karlsson, M., Bry, L., Glickman, J.N., et al. (2010). Enterobacteriaceae act in concert with the gut microbiota to induce spontaneous and maternally transmitted colitis. Cell Host Microbe *8*, 292–300.

Geuking, M.B., Cahenzli, J., Lawson, M.A., Ng, D.C., Slack, E., Hapfelmeier, S., McCoy, K.D., and Macpherson, A.J. (2011). Intestinal bacterial colonization induces mutualistic regulatory T cell responses. Immunity *34*, 794–806.

Gordon, J.I., Dewey, K.G., Mills, D.A., and Medzhitov, R.M. (2012). The human gut microbiota and undernutrition. Sci. Transl. Med. *4*, 137ps112.

Grainger, J.R., Wohlfert, E.A., Fuss, I.J., Bouladoux, N., Askenase, M.H., Legrand, F., Koo, L.Y., Brenchley, J.M., Fraser, I.D., and Belkaid, Y. (2013). Inflammatory monocytes regulate pathologic responses to commensals during acute gastrointestinal infection. Nat. Med. *19*, 713–721.

Gregor, M.F., and Hotamisligil, G.S. (2011). Inflammatory mechanisms in obesity. Annu. Rev. Immunol. 29, 415–445.

Grice, E.A., Kong, H.H., Renaud, G., Young, A.C., Bouffard, G.G., Blakesley, R.W., Wolfsberg, T.G., Turner, M.L., and Segre, J.A.; NISC Comparative Sequencing Program (2008). A diversity profile of the human skin microbiota. Genome Res. *18*, 1043–1050.

Grice, E.A., Kong, H.H., Conlan, S., Deming, C.B., Davis, J., Young, A.C., Bouffard, G.G., Blakesley, R.W., Murray, P.R., Green, E.D., et al.; NISC Comparative Sequencing Program (2009). Topographical and temporal diversity of the human skin microbiome. Science *324*, 1190–1192.

Grivennikov, S.I., and Karin, M. (2010). Dangerous liaisons: STAT3 and NF-kappaB collaboration and crosstalk in cancer. Cytokine Growth Factor Rev. *21*, 11–19.

Grivennikov, S.I., Wang, K., Mucida, D., Stewart, C.A., Schnabl, B., Jauch, D., Taniguchi, K., Yu, G.Y., Osterreicher, C.H., Hung, K.E., et al. (2012). Adenomalinked barrier defects and microbial products drive IL-23/IL-17-mediated tumour growth. Nature *491*, 254–258.

Gupta, N., Limbago, B.M., Patel, J.B., and Kallen, A.J. (2011). Carbapenemresistant *Enterobacteriaceae*: Epidemiology and prevention. Clin. Infect. Dis. 53, 60–67.

Haas, A., Zimmermann, K., Graw, F., Slack, E., Rusert, P., Ledergerber, B., Bossart, W., Weber, R., Thurnheer, M.C., Battegay, M., et al.; Swiss HIV Cohort Study (2011). Systemic antibody responses to gut commensal bacteria during chronic HIV-1 infection. Gut *60*, 1506–1519.

Hall, J.A., Bouladoux, N., Sun, C.M., Wohlfert, E.A., Blank, R.B., Zhu, Q., Grigg, M.E., Berzofsky, J.A., and Belkaid, Y. (2008). Commensal DNA limits regulatory T cell conversion and is a natural adjuvant of intestinal immune responses. Immunity 29, 637–649.

Hall, J.A., Cannons, J.L., Grainger, J.R., Dos Santos, L.M., Hand, T.W., Naik, S., Wohlfert, E.A., Chou, D.B., Oldenhove, G., Robinson, M., et al. (2011a). Essential role for retinoic acid in the promotion of CD4(+) T cell effector responses via retinoic acid receptor alpha. Immunity *34*, 435–447.

Hall, J.A., Grainger, J.R., Spencer, S.P., and Belkaid, Y. (2011b). The role of retinoic acid in tolerance and immunity. Immunity 35, 13–22.

Hamada, H., Hiroi, T., Nishiyama, Y., Takahashi, H., Masunaga, Y., Hachimura, S., Kaminogawa, S., Takahashi-Iwanaga, H., Iwanaga, T., Kiyono, H., et al. (2002). Identification of multiple isolated lymphoid follicles on the antimesenteric wall of the mouse small intestine. J. Immunol. *168*, 57–64.

Hammami, R., Fernandez, B., Lacroix, C., and Fliss, I. (2013). Anti-infective properties of bacteriocins: an update. Cell. Mol. Life Sci. 70, 2947–2967.

Hand, T.W., Dos Santos, L.M., Bouladoux, N., Molloy, M.J., Pagán, A.J., Pepper, M., Maynard, C.L., Elson, C.O., 3rd, and Belkaid, Y. (2012). Acute gastrointestinal infection induces long-lived microbiota-specific T cell responses. Science 337, 1553–1556.

Hapfelmeier, S., Lawson, M.A., Slack, E., Kirundi, J.K., Stoel, M., Heikenwalder, M., Cahenzli, J., Velykoredko, Y., Balmer, M.L., Endt, K., et al. (2010). Reversible microbial colonization of germ-free mice reveals the dynamics of IgA immune responses. Science *328*, 1705–1709.

Hayes, K.S., Bancroft, A.J., Goldrick, M., Portsmouth, C., Roberts, I.S., and Grencis, R.K. (2010). Exploitation of the intestinal microflora by the parasitic nematode Trichuris muris. Science *328*, 1391–1394.

Heimesaat, M.M., Bereswill, S., Fischer, A., Fuchs, D., Struck, D., Niebergall, J., Jahn, H.K., Dunay, I.R., Moter, A., Gescher, D.M., et al. (2006). Gram-negative bacteria aggravate murine small intestinal Th1-type immunopathology following oral infection with Toxoplasma gondii. J. Immunol. *177*, 8785–8795.

Herbst, T., Sichelstiel, A., Schär, C., Yadava, K., Bürki, K., Cahenzli, J., McCoy, K., Marsland, B.J., and Harris, N.L. (2011). Dysregulation of allergic airway inflammation in the absence of microbial colonization. Am. J. Respir. Crit. Care Med. *184*, 198–205.

Hilty, M., Burke, C., Pedro, H., Cardenas, P., Bush, A., Bossley, C., Davies, J., Ervine, A., Poulter, L., Pachter, L., et al. (2010). Disordered microbial communities in asthmatic airways. PLoS ONE 5, e8578.

Homann, D., Teyton, L., and Oldstone, M.B. (2001). Differential regulation of antiviral T-cell immunity results in stable CD8+ but declining CD4+ T-cell memory. Nat. Med. 7, 913–919.

Hooper, L.V., and Macpherson, A.J. (2010). Immune adaptations that maintain homeostasis with the intestinal microbiota. Nat. Rev. Immunol. 10, 159–169.

Hooper, L.V., Wong, M.H., Thelin, A., Hansson, L., Falk, P.G., and Gordon, J.I. (2001). Molecular analysis of commensal host-microbial relationships in the intestine. Science *291*, 881–884.

Hornquist, E., and Lycke, N. (1995). Cholera toxin increases T lymphocyte responses to unrelated antigens. Adv. Exp. Med. Biol. *371B*, 1507–1512.

Hu, B., Elinav, E., Huber, S., Strowig, T., Hao, L., Hafemann, A., Jin, C., Wunderlich, C., Wunderlich, T., Eisenbarth, S.C., and Flavell, R.A. (2013). Microbiota-induced activation of epithelial IL-6 signaling links inflammasome-driven inflammation with transmissible cancer. Proc. Natl. Acad. Sci. USA *110*, 9862–9867.

Ichinohe, T., Pang, I.K., Kumamoto, Y., Peaper, D.R., Ho, J.H., Murray, T.S., and Iwasaki, A. (2011). Microbiota regulates immune defense against respiratory tract influenza A virus infection. Proc. Natl. Acad. Sci. USA *108*, 5354–5359.

lida, N., Dzutsev, A., Stewart, C.A., Smith, L., Bouladoux, N., Weingarten, R.A., Molina, D.A., Salcedo, R., Back, T., Cramer, S., et al. (2013). Commensal bacteria control cancer response to therapy by modulating the tumor microenvironment. Science *342*, 967–970.

Iliev, I.D., Funari, V.A., Taylor, K.D., Nguyen, Q., Reyes, C.N., Strom, S.P., Brown, J., Becker, C.A., Fleshner, P.R., Dubinsky, M., et al. (2012). Interactions between commensal fungi and the C-type lectin receptor Dectin-1 influence colitis. Science 336, 1314–1317.

Ismail, A.S., Severson, K.M., Vaishnava, S., Behrendt, C.L., Yu, X., Benjamin, J.L., Ruhn, K.A., Hou, B., DeFranco, A.L., Yarovinsky, F., and Hooper, L.V. (2011). Gammadelta intraepithelial lymphocytes are essential mediators of host-microbial homeostasis at the intestinal mucosal surface. Proc. Natl. Acad. Sci. USA 108, 8743–8748.

Issa, M., Ananthakrishnan, A.N., and Binion, D.G. (2008). Clostridium difficile and inflammatory bowel disease. Inflamm. Bowel Dis. 14, 1432–1442.

Ivanov, I.I., Frutos, Rde.L., Manel, N., Yoshinaga, K., Rifkin, D.B., Sartor, R.B., Finlay, B.B., and Littman, D.R. (2008). Specific microbiota direct the differentiation of IL-17-producing T-helper cells in the mucosa of the small intestine. Cell Host Microbe *4*, 337–349.

Ivanov, I.I., Atarashi, K., Manel, N., Brodie, E.L., Shima, T., Karaoz, U., Wei, D., Goldfarb, K.C., Santee, C.A., Lynch, S.V., et al. (2009). Induction of intestinal Th17 cells by segmented filamentous bacteria. Cell *139*, 485–498.

Iwase, T., Uehara, Y., Shinji, H., Tajima, A., Seo, H., Takada, K., Agata, T., and Mizunoe, Y. (2010). Staphylococcus epidermidis Esp inhibits Staphylococcus aureus biofilm formation and nasal colonization. Nature *465*, 346–349.

Jayashree, B., Bibin, Y.S., Prabhu, D., Shanthirani, C.S., Gokulakrishnan, K., Lakshmi, B.S., Mohan, V., and Balasubramanyam, M. (2013). Increased circulatory levels of lipopolysaccharide (LPS) and zonulin signify novel biomarkers of proinflammation in patients with type 2 diabetes. Mol. Cell. Biochem. *388*, 2033–210.

Johnston, C.J., McSorley, H.J., Anderton, S.M., Wigmore, S.J., and Maizels, R.M. (2014). Helminths and immunological tolerance. Transplantation *97*, 127–132.

Josefowicz, S.Z., Niec, R.E., Kim, H.Y., Treuting, P., Chinen, T., Zheng, Y., Umetsu, D.T., and Rudensky, A.Y. (2012a). Extrathymically generated regulatory T cells control mucosal TH2 inflammation. Nature *482*, 395–399.

Jung, C., Meinzer, U., Montcuquet, N., Thachil, E., Château, D., Thiébaut, R., Roy, M., Alnabhani, Z., Berrebi, D., Dussaillant, M., et al. (2012). Yersinia pseudotuberculosis disrupts intestinal barrier integrity through hematopoietic TLR-2 signaling. J. Clin. Invest. *122*, 2239–2251.

Kamada, N., Chen, G.Y., Inohara, N., and Núñez, G. (2013). Control of pathogens and pathobionts by the gut microbiota. Nat. Immunol. *14*, 685–690.

Kane, M., Case, L.K., Kopaskie, K., Kozlova, A., MacDearmid, C., Chervonsky, A.V., and Golovkina, T.V. (2011). Successful transmission of a retrovirus depends on the commensal microbiota. Science *334*, 245–249.

Karimi, K., Inman, M.D., Bienenstock, J., and Forsythe, P. (2009). *Lactobacillus reuteri*-induced regulatory T cells protect against an allergic airway response in mice. Am. J. Respir. Crit. Care Med. *179*, 186–193.

Karlsson, F.H., Tremaroli, V., Nookaew, I., Bergström, G., Behre, C.J., Fagerberg, B., Nielsen, J., and Bäckhed, F. (2013). Gut metagenome in European women with normal, impaired and diabetic glucose control. Nature *498*, 99–103.

Kelsall, B. (2008). Recent progress in understanding the phenotype and function of intestinal dendritic cells and macrophages. Mucosal Immunol. *1*, 460–469.

Kinnebrew, M.A., Buffie, C.G., Diehl, G.E., Zenewicz, L.A., Leiner, I., Hohl, T.M., Flavell, R.A., Littman, D.R., and Pamer, E.G. (2012). Interleukin 23 production by intestinal CD103(+)CD11b(+) dendritic cells in response to bacterial flagellin enhances mucosal innate immune defense. Immunity *36*, 276–287.

Kiss, E.A., Vonarbourg, C., Kopfmann, S., Hobeika, E., Finke, D., Esser, C., and Diefenbach, A. (2011). Natural aryl hydrocarbon receptor ligands control organogenesis of intestinal lymphoid follicles. Science *334*, 1561–1565.

Kiyono, H., McGhee, J.R., Wannemuehler, M.J., and Michalek, S.M. (1982). Lack of oral tolerance in C3H/HeJ mice. J. Exp. Med. *155*, 605–610.

Klatt, N.R., Funderburg, N.T., and Brenchley, J.M. (2013). Microbial translocation, immune activation, and HIV disease. Trends Microbiol. 21, 6–13.

Klebanoff, C.A., Spencer, S.P., Torabi-Parizi, P., Grainger, J.R., Roychoudhuri, R., Ji, Y., Sukumar, M., Muranski, P., Scott, C.D., Hall, J.A., et al. (2013). Retinoic acid controls the homeostasis of pre-cDC-derived splenic and intestinal dendritic cells. J. Exp. Med. *210*, 1961–1976.

Kollmann, T.R., Levy, O., Montgomery, R.R., and Goriely, S. (2012). Innate immune function by Toll-like receptors: distinct responses in newborns and the elderly. Immunity *37*, 771–783.

Kong, H.H., Oh, J., Deming, C., Conlan, S., Grice, E.A., Beatson, M.A., Nomicos, E., Polley, E.C., Komarow, H.D., Murray, P.R., et al.; NISC Comparative Sequence Program (2012). Temporal shifts in the skin microbiome associated with disease flares and treatment in children with atopic dermatitis. Genome Res. 22, 850–859.

Korpe, P.S., and Petri, W.A., Jr. (2012). Environmental enteropathy: critical implications of a poorly understood condition. Trends Mol. Med. 18, 328–336.

Kosek, M., Bern, C., and Guerrant, R.L. (2003). The global burden of diarrhoeal disease, as estimated from studies published between 1992 and 2000. Bull. World Health Organ. *81*, 197–204.

Kostic, A.D., Gevers, D., Pedamallu, C.S., Michaud, M., Duke, F., Earl, A.M., Ojesina, A.I., Jung, J., Bass, A.J., Tabernero, J., et al. (2012). Genomic analysis identifies association of Fusobacterium with colorectal carcinoma. Genome Res. *22*, 292–298.

Kostic, A.D., Chun, E., Robertson, L., Glickman, J.N., Gallini, C.A., Michaud, M., Clancy, T.E., Chung, D.C., Lochhead, P., Hold, G.L., et al. (2013). Fusobacterium nucleatum potentiates intestinal tumorigenesis and modulates the tumor-immune microenvironment. Cell Host Microbe *14*, 207–215.

Kriegel, M.A., Sefik, E., Hill, J.A., Wu, H.J., Benoist, C., and Mathis, D. (2011). Naturally transmitted segmented filamentous bacteria segregate with diabetes protection in nonobese diabetic mice. Proc. Natl. Acad. Sci. USA *108*, 11548– 11553.

Kullberg, M.C., Jankovic, D., Gorelick, P.L., Caspar, P., Letterio, J.J., Cheever, A.W., and Sher, A. (2002). Bacteria-triggered CD4(+) T regulatory cells suppress Helicobacter hepaticus-induced colitis. J. Exp. Med. *196*, 505–515.

Kuss, S.K., Best, G.T., Etheredge, C.A., Pruijssers, A.J., Frierson, J.M., Hooper, L.V., Dermody, T.S., and Pfeiffer, J.K. (2011). Intestinal microbiota promote enteric virus replication and systemic pathogenesis. Science *334*, 249–252.

Lai, Y., Di Nardo, A., Nakatsuji, T., Leichtle, A., Yang, Y., Cogen, A.L., Wu, Z.R., Hooper, L.V., Schmidt, R.R., von Aulock, S., et al. (2009). Commensal bacteria regulate Toll-like receptor 3-dependent inflammation after skin injury. Nat. Med. *15*, 1377–1382.

Lamps, L.W., Madhusudhan, K.T., Havens, J.M., Greenson, J.K., Bronner, M.P., Chiles, M.C., Dean, P.J., and Scott, M.A. (2003). Pathogenic Yersinia DNA is detected in bowel and mesenteric lymph nodes from patients with Crohn's disease. Am. J. Surg. Pathol. *27*, 220–227.

Lathrop, S.K., Bloom, S.M., Rao, S.M., Nutsch, K., Lio, C.W., Santacruz, N., Peterson, D.A., Stappenbeck, T.S., and Hsieh, C.S. (2011). Peripheral education of the immune system by colonic commensal microbiota. Nature 478, 250–254.

Lee, Y.K., Menezes, J.S., Umesaki, Y., and Mazmanian, S.K. (2011). Proinflammatory T-cell responses to gut microbiota promote experimental autoimmune encephalomyelitis. Proc. Natl. Acad. Sci. USA *108* (*Suppl 1*), 4615–4622.

Ley, R.E., Peterson, D.A., and Gordon, J.I. (2006a). Ecological and evolutionary forces shaping microbial diversity in the human intestine. Cell *124*, 837–848.

Ley, R.E., Turnbaugh, P.J., Klein, S., and Gordon, J.I. (2006b). Microbial ecology: human gut microbes associated with obesity. Nature 444, 1022–1023.

Li, Y., Innocentin, S., Withers, D.R., Roberts, N.A., Gallagher, A.R., Grigorieva, E.F., Wilhelm, C., and Veldhoen, M. (2011). Exogenous stimuli maintain intraepithelial lymphocytes via aryl hydrocarbon receptor activation. Cell *147*, 629–640.

Lodes, M.J., Cong, Y., Elson, C.O., Mohamath, R., Landers, C.J., Targan, S.R., Fort, M., and Hershberg, R.M. (2004). Bacterial flagellin is a dominant antigen in Crohn disease. J. Clin. Invest. *113*, 1296–1306.

Lotz, M., Gütle, D., Walther, S., Ménard, S., Bogdan, C., and Hornef, M.W. (2006). Postnatal acquisition of endotoxin tolerance in intestinal epithelial cells. J. Exp. Med. *203*, 973–984.

Lowe, E.L., Crother, T.R., Rabizadeh, S., Hu, B., Wang, H., Chen, S., Shimada, K., Wong, M.H., Michelsen, K.S., and Arditi, M. (2010). Toll-like receptor 2 signaling protects mice from tumor development in a mouse model of colitis-induced cancer. PLoS ONE *5*, e13027.

Lozupone, C.A., Stombaugh, J.I., Gordon, J.I., Jansson, J.K., and Knight, R. (2012). Diversity, stability and resilience of the human gut microbiota. Nature 489, 220–230.

Lupp, C., Robertson, M.L., Wickham, M.E., Sekirov, I., Champion, O.L., Gaynor, E.C., and Finlay, B.B. (2007). Host-mediated inflammation disrupts the intestinal microbiota and promotes the overgrowth of Enterobacteriaceae. Cell Host Microbe 2, 204.

Macpherson, A.J., and Harris, N.L. (2004). Interactions between commensal intestinal bacteria and the immune system. Nat. Rev. Immunol. *4*, 478–485.

Macpherson, A.J., and Uhr, T. (2004). Induction of protective IgA by intestinal dendritic cells carrying commensal bacteria. Science *303*, 1662–1665.

Macpherson, A., Khoo, U.Y., Forgacs, I., Philpott-Howard, J., and Bjarnason, I. (1996). Mucosal antibodies in inflammatory bowel disease are directed against intestinal bacteria. Gut *38*, 365–375.

Macpherson, A.J., Hunziker, L., McCoy, K., and Lamarre, A. (2001). IgA responses in the intestinal mucosa against pathogenic and non-pathogenic microorganisms. Microbes Infect. *3*, 1021–1035.

Macpherson, A.J., Slack, E., Geuking, M.B., and McCoy, K.D. (2009). The mucosal firewalls against commensal intestinal microbes. Semin. Immunopathol. *31*, 145–149.

Maloy, K.J., and Powrie, F. (2011). Intestinal homeostasis and its breakdown in inflammatory bowel disease. Nature 474, 298–306.

Marcobal, A., and Sonnenburg, J.L. (2012). Human milk oligosaccharide consumption by intestinal microbiota. Clin. Microbiol. Infect. 18 (Suppl 4), 12–15.

Marcobal, A., Barboza, M., Froehlich, J.W., Block, D.E., German, J.B., Lebrilla, C.B., and Mills, D.A. (2010). Consumption of human milk oligosaccharides by gut-related microbes. J. Agric. Food Chem. *58*, 5334–5340.

Markle, J.G., Frank, D.N., Mortin-Toth, S., Robertson, C.E., Feazel, L.M., Rolle-Kampczyk, U., von Bergen, M., McCoy, K.D., Macpherson, A.J., and Danska, J.S. (2013). Sex differences in the gut microbiome drive hormone-dependent regulation of autoimmunity. Science *339*, 1084–1088.

Marshall, B.J., and Warren, J.R. (1984). Unidentified curved bacilli in the stomach of patients with gastritis and peptic ulceration. Lancet *1*, 1311–1315.

Martinoli, C., Chiavelli, A., and Rescigno, M. (2007). Entry route of Salmonella typhimurium directs the type of induced immune response. Immunity *27*, 975–984.

Maslowski, K.M., Vieira, A.T., Ng, A., Kranich, J., Sierro, F., Yu, D., Schilter, H.C., Rolph, M.S., Mackay, F., Artis, D., et al. (2009). Regulation of inflammatory responses by gut microbiota and chemoattractant receptor GPR43. Nature *461*, 1282–1286.

Mazmanian, S.K., Liu, C.H., Tzianabos, A.O., and Kasper, D.L. (2005). An immunomodulatory molecule of symbiotic bacteria directs maturation of the host immune system. Cell *122*, 107–118.

Mazmanian, S.K., Round, J.L., and Kasper, D.L. (2008). A microbial symbiosis factor prevents intestinal inflammatory disease. Nature 453, 620–625.

McGuckin, M.A., Lindén, S.K., Sutton, P., and Florin, T.H. (2011). Mucin dynamics and enteric pathogens. Nat. Rev. Microbiol. 9, 265–278.

Meijer, K., de Vos, P., and Priebe, M.G. (2010). Butyrate and other short-chain fatty acids as modulators of immunity: what relevance for health? Curr. Opin. Clin. Nutr. Metab. Care *13*, 715–721.

Meinzer, U., Barreau, F., Esmiol-Welterlin, S., Jung, C., Villard, C., Léger, T., Ben-Mkaddem, S., Berrebi, D., Dussaillant, M., Alnabhani, Z., et al. (2012). Yersinia pseudotuberculosis effector YopJ subverts the Nod2/RICK/TAK1 pathway and activates caspase-1 to induce intestinal barrier dysfunction. Cell Host Microbe *11*, 337–351.

Molloy, M.J., Bouladoux, N., and Belkaid, Y. (2012). Intestinal microbiota: shaping local and systemic immune responses. Semin. Immunol. 24, 58–66.

Molloy, M.J., Grainger, J.R., Bouladoux, N., Hand, T.W., Koo, L.Y., Naik, S., Quinones, M., Dzutsev, A.K., Gao, J.L., Trinchieri, G., et al. (2013). Intraluminal containment of commensal outgrowth in the gut during infection-induced dysbiosis. Cell Host Microbe *14*, 318–328.

Monira, S., Nakamura, S., Gotoh, K., Izutsu, K., Watanabe, H., Alam, N.H., Endtz, H.P., Cravioto, A., Ali, S.I., Nakaya, T., et al. (2011). Gut microbiota of healthy and malnourished children in bangladesh. Front. Microbiol. *2*, 228.

Mora, J.R., Iwata, M., and von Andrian, U.H. (2008). Vitamin effects on the immune system: vitamins A and D take centre stage. Nat. Rev. Immunol. *8*, 685–698.

Mucida, D., Kutchukhidze, N., Erazo, A., Russo, M., Lafaille, J.J., and Curotto de Lafaille, M.A. (2005). Oral tolerance in the absence of naturally occurring Tregs. J. Clin. Invest. *115*, 1923–1933.

Mucida, D., Park, Y., Kim, G., Turovskaya, O., Scott, I., Kronenberg, M., and Cheroutre, H. (2007). Reciprocal TH17 and regulatory T cell differentiation mediated by retinoic acid. Science *317*, 256–260.

Mukherjee, S., Partch, C.L., Lehotzky, R.E., Whitham, C.V., Chu, H., Bevins, C.L., Gardner, K.H., and Hooper, L.V. (2009). Regulation of C-type lectin antimicrobial activity by a flexible N-terminal prosegment. J. Biol. Chem. 284, 4881–4888.

Murray, B.E. (2000). Vancomycin-resistant enterococcal infections. N. Engl. J. Med. 342, 710–721.

Naik, S., Bouladoux, N., Wilhelm, C., Molloy, M.J., Salcedo, R., Kastenmuller, W., Deming, C., Quinones, M., Koo, L., Conlan, S., et al. (2012). Compartmentalized control of skin immunity by resident commensals. Science *337*, 1115– 1119.

Ng, K.M., Ferreyra, J.A., Higginbottom, S.K., Lynch, J.B., Kashyap, P.C., Gopinath, S., Naidu, N., Choudhury, B., Weimer, B.C., Monack, D.M., and Sonnenburg, J.L. (2013). Microbiota-liberated host sugars facilitate post-antibiotic expansion of enteric pathogens. Nature *502*, 96–99.

Niess, J.H., and Adler, G. (2010). Enteric flora expands gut lamina propria CX3CR1+ dendritic cells supporting inflammatory immune responses under normal and inflammatory conditions. J. Immunol. *184*, 2026–2037.

O'Mahony, C., Scully, P., O'Mahony, D., Murphy, S., O'Brien, F., Lyons, A., Sherlock, G., MacSharry, J., Kiely, B., Shanahan, F., and O'Mahony, L. (2008). Commensal-induced regulatory T cells mediate protection against pathogen-stimulated NF-kappaB activation. PLoS Pathog. *4*, e1000112.

Ochoa-Repáraz, J., Mielcarz, D.W., Ditrio, L.E., Burroughs, A.R., Foureau, D.M., Haque-Begum, S., and Kasper, L.H. (2009). Role of gut commensal microflora in the development of experimental autoimmune encephalomyelitis. J. Immunol. *183*, 6041–6050.

Ochoa-Repáraz, J., Mielcarz, D.W., Ditrio, L.E., Burroughs, A.R., Begum-Haque, S., Dasgupta, S., Kasper, D.L., and Kasper, L.H. (2010a). Central nervous system demyelinating disease protection by the human commensal Bacteroides fragilis depends on polysaccharide A expression. J. Immunol. *185*, 4101–4108. Ochoa-Repáraz, J., Mielcarz, D.W., Wang, Y., Begum-Haque, S., Dasgupta, S., Kasper, D.L., and Kasper, L.H. (2010b). A polysaccharide from the human commensal Bacteroides fragilis protects against CNS demyelinating disease. Mucosal Immunol. 3, 487–495.

Ohnmacht, C., Marques, R., Presley, L., Sawa, S., Lochner, M., and Eberl, G. (2011). Intestinal microbiota, evolution of the immune system and the bad reputation of pro-inflammatory immunity. Cell. Microbiol. *13*, 653–659.

Olszak, T., An, D., Zeissig, S., Vera, M.P., Richter, J., Franke, A., Glickman, J.N., Siebert, R., Baron, R.M., Kasper, D.L., and Blumberg, R.S. (2012). Microbial exposure during early life has persistent effects on natural killer T cell function. Science 336, 489–493.

Osborn, O., and Olefsky, J.M. (2012). The cellular and signaling networks linking the immune system and metabolism in disease. Nat. Med. *18*, 363–374.

Pacheco, A.R., Curtis, M.M., Ritchie, J.M., Munera, D., Waldor, M.K., Moreira, C.G., and Sperandio, V. (2012). Fucose sensing regulates bacterial intestinal colonization. Nature *492*, 113–117.

Packey, C.D., and Sartor, R.B. (2008). Interplay of commensal and pathogenic bacteria, genetic mutations, and immunoregulatory defects in the pathogenesis of inflammatory bowel diseases. J. Intern. Med. *263*, 597–606.

Paulos, C.M., Wrzesinski, C., Kaiser, A., Hinrichs, C.S., Chieppa, M., Cassard, L., Palmer, D.C., Boni, A., Muranski, P., Yu, Z., et al. (2007). Microbial translocation augments the function of adoptively transferred self/tumor-specific CD8+ T cells via TLR4 signaling. J. Clin. Invest. *117*, 2197–2204.

Pepper, M., Linehan, J.L., Pagán, A.J., Zell, T., Dileepan, T., Cleary, P.P., and Jenkins, M.K. (2010). Different routes of bacterial infection induce long-lived TH1 memory cells and short-lived TH17 cells. Nat. Immunol. *11*, 83–89.

Perez, P.F., Doré, J., Leclerc, M., Levenez, F., Benyacoub, J., Serrant, P., Segura-Roggero, I., Schiffrin, E.J., and Donnet-Hughes, A. (2007). Bacterial imprinting of the neonatal immune system: lessons from maternal cells? Pediatrics *119*, e724–e732.

Peterson, D.A., McNulty, N.P., Guruge, J.L., and Gordon, J.I. (2007). IgA response to symbiotic bacteria as a mediator of gut homeostasis. Cell Host Microbe 2, 328–339.

Powell, N., Walker, A.W., Stolarczyk, E., Canavan, J.B., Gökmen, M.R., Marks, E., Jackson, I., Hashim, A., Curtis, M.A., Jenner, R.G., et al. (2012). The transcription factor T-bet regulates intestinal inflammation mediated by interleukin-7 receptor+ innate lymphoid cells. Immunity *37*, 674–684.

Pozzilli, P., Signore, A., Williams, A.J., and Beales, P.E. (1993). NOD mouse colonies around the world-recent facts and figures. Immunol. Today 14, 193–196.

PrabhuDas, M., Adkins, B., Gans, H., King, C., Levy, O., Ramilo, O., and Siegrist, C.A. (2011). Challenges in infant immunity: implications for responses to infection and vaccines. Nat. Immunol. *12*, 189–194.

Pryde, S.E., Duncan, S.H., Hold, G.L., Stewart, C.S., and Flint, H.J. (2002). The microbiology of butyrate formation in the human colon. FEMS Microbiol. Lett. *217*, 133–139.

Qin, J., Li, Y., Cai, Z., Li, S., Zhu, J., Zhang, F., Liang, S., Zhang, W., Guan, Y., Shen, D., et al. (2012). A metagenome-wide association study of gut microbiota in type 2 diabetes. Nature *490*, 55–60.

Qiu, J., Guo, X., Chen, Z.M., He, L., Sonnenberg, G.F., Artis, D., Fu, Y.X., and Zhou, L. (2013). Group 3 innate lymphoid cells inhibit T-cell-mediated intestinal inflammation through aryl hydrocarbon receptor signaling and regulation of microflora. Immunity *39*, 386–399.

Raetz, M., Hwang, S.H., Wilhelm, C.L., Kirkland, D., Benson, A., Sturge, C.R., Mirpuri, J., Vaishnava, S., Hou, B., Defranco, A.L., et al. (2013). Parasiteinduced T(H)1 cells and intestinal dysbiosis cooperate in IFN-gamma-dependent elimination of Paneth cells. Nat. Immunol. *14*, 136–142.

Rakoff-Nahoum, S., and Medzhitov, R. (2007). Regulation of spontaneous intestinal tumorigenesis through the adaptor protein MyD88. Science *317*, 124–127.

Rakoff-Nahoum, S., Paglino, J., Eslami-Varzaneh, F., Edberg, S., and Medzhitov, R. (2004). Recognition of commensal microflora by toll-like receptors is required for intestinal homeostasis. Cell *118*, 229–241. Rivas, M.A., Beaudoin, M., Gardet, A., Stevens, C., Sharma, Y., Zhang, C.K., Boucher, G., Ripke, S., Ellinghaus, D., Burtt, N., et al.; National Institute of Diabetes and Digestive Kidney Diseases Inflammatory Bowel Disease Genetics Consortium (NIDDK IBDGC); United Kingdom Inflammatory Bowel Disease Genetics Consortium; International Inflammatory Bowel Disease Genetics Consortium (2011). Deep resequencing of GWAS loci identifies independent rare variants associated with inflammatory bowel disease. Nat. Genet. *43*, 1066–1073.

Rolhion, N., and Darfeuille-Michaud, A. (2007). Adherent-invasive Escherichia coli in inflammatory bowel disease. Inflamm. Bowel Dis. *13*, 1277–1283.

Round, J.L., Lee, S.M., Li, J., Tran, G., Jabri, B., Chatila, T.A., and Mazmanian, S.K. (2011). The Toll-like receptor 2 pathway establishes colonization by a commensal of the human microbiota. Science *332*, 974–977.

Rubinstein, M.R., Wang, X., Liu, W., Hao, Y., Cai, G., and Han, Y.W. (2013). Fusobacterium nucleatum promotes colorectal carcinogenesis by modulating E-cadherin/ β -catenin signaling via its FadA adhesin. Cell Host Microbe 14, 195–206.

Salcedo, R., Worschech, A., Cardone, M., Jones, Y., Gyulai, Z., Dai, R.M., Wang, E., Ma, W., Haines, D., O'hUigin, C., et al. (2010). MyD88-mediated signaling prevents development of adenocarcinomas of the colon: role of interleukin 18. J. Exp. Med. 207, 1625–1636.

Sansonetti, P.J., and Di Santo, J.P. (2007). Debugging how bacteria manipulate the immune response. Immunity 26, 149–161.

Sartor, R.B. (2006). Mechanisms of disease: pathogenesis of Crohn's disease and ulcerative colitis. Nat. Clin. Pract. Gastroenterol. Hepatol. *3*, 390–407.

Satoh-Takayama, N., Vosshenrich, C.A., Lesjean-Pottier, S., Sawa, S., Lochner, M., Rattis, F., Mention, J.J., Thiam, K., Cerf-Bensussan, N., Mandelboim, O., et al. (2008). Microbial flora drives interleukin 22 production in intestinal NKp46+ cells that provide innate mucosal immune defense. Immunity *29*, 958–970.

Savy, M., Edmond, K., Fine, P.E., Hall, A., Hennig, B.J., Moore, S.E., Mulholland, K., Schaible, U., and Prentice, A.M. (2009). Landscape analysis of interactions between nutrition and vaccine responses in children. J. Nutr. *139*, 2154S–2218S.

Schamberger, G.P., and Diez-Gonzalez, F. (2002). Selection of recently isolated colicinogenic Escherichia coli strains inhibitory to Escherichia coli O157:H7. J. Food Prot. *65*, 1381–1387.

Scher, J.U., Sczesnak, A., Longman, R.S., Segata, N., Ubeda, C., Bielski, C., Rostron, T., Cerundolo, V., Pamer, E.G., Abramson, S.B., et al. (2013). Expansion of intestinal Prevotella copri correlates with enhanced susceptibility to arthritis. eLife 2, e01202.

Sczesnak, A., Segata, N., Qin, X., Gevers, D., Petrosino, J.F., Huttenhower, C., Littman, D.R., and Ivanov, I.I. (2011). The genome of th17 cell-inducing segmented filamentous bacteria reveals extensive auxotrophy and adaptations to the intestinal environment. Cell Host Microbe *10*, 260–272.

Shan, M., Gentile, M., Yeiser, J.R., Walland, A.C., Bornstein, V.U., Chen, K., He, B., Cassis, L., Bigas, A., Cols, M., et al. (2013). Mucus enhances gut homeostasis and oral tolerance by delivering immunoregulatory signals. Science 342, 447–453.

Shi, C., Jia, T., Mendez-Ferrer, S., Hohl, T.M., Serbina, N.V., Lipuma, L., Leiner, I., Li, M.O., Frenette, P.S., and Pamer, E.G. (2011). Bone marrow mesenchymal stem and progenitor cells induce monocyte emigration in response to circulating toll-like receptor ligands. Immunity *34*, 590–601.

Shulzhenko, N., Morgun, A., Hsiao, W., Battle, M., Yao, M., Gavrilova, O., Orandle, M., Mayer, L., Macpherson, A.J., McCoy, K.D., et al. (2011). Crosstalk between B lymphocytes, microbiota and the intestinal epithelium governs immunity versus metabolism in the gut. Nat. Med. *17*, 1585–1593.

Siegrist, C.A. (2001). Neonatal and early life vaccinology. Vaccine 19, 3331–3346.

Smith, K., McCoy, K.D., and Macpherson, A.J. (2007). Use of axenic animals in studying the adaptation of mammals to their commensal intestinal microbiota. Semin. Immunol. *19*, 59–69.

Smith, P.M., Howitt, M.R., Panikov, N., Michaud, M., Gallini, C.A., Bohlooly-Y, M., Glickman, J.N., and Garrett, W.S. (2013). The microbial metabolites, short-chain fatty acids, regulate colonic Treg cell homeostasis. Science *341*, 569–573.

Smits, H.H., Engering, A., van der Kleij, D., de Jong, E.C., Schipper, K., van Capel, T.M., Zaat, B.A., Yazdanbakhsh, M., Wierenga, E.A., van Kooyk, Y., and Kapsenberg, M.L. (2005). Selective probiotic bacteria induce IL-10-producing regulatory T cells in vitro by modulating dendritic cell function through dendritic cell-specific intercellular adhesion molecule 3-grabbing nonintegrin. J. Allergy Clin. Immunol. *115*, 1260–1267.

Sobhani, I., Tap, J., Roudot-Thoraval, F., Roperch, J.P., Letulle, S., Langella, P., Corthier, G., Tran Van Nhieu, J., and Furet, J.P. (2011). Microbial dysbiosis in colorectal cancer (CRC) patients. PLoS ONE *6*, e16393.

Sonnenberg, G.F., Monticelli, L.A., Alenghat, T., Fung, T.C., Hutnick, N.A., Kunisawa, J., Shibata, N., Grunberg, S., Sinha, R., Zahm, A.M., et al. (2012). Innate lymphoid cells promote anatomical containment of lymphoid-resident commensal bacteria. Science *336*, 1321–1325.

Srinivasan, S., Liu, C., Mitchell, C.M., Fiedler, T.L., Thomas, K.K., Agnew, K.J., Marrazzo, J.M., and Fredricks, D.N. (2010). Temporal variability of human vaginal bacteria and relationship with bacterial vaginosis. PLoS ONE 5, e10197.

Stappenbeck, T.S., Hooper, L.V., and Gordon, J.I. (2002). Developmental regulation of intestinal angiogenesis by indigenous microbes via Paneth cells. Proc. Natl. Acad. Sci. USA *99*, 15451–15455.

Stecher, B., Robbiani, R., Walker, A.W., Westendorf, A.M., Barthel, M., Kremer, M., Chaffron, S., Macpherson, A.J., Buer, J., Parkhill, J., et al. (2007). Salmonella enterica serovar typhimurium exploits inflammation to compete with the intestinal microbiota. PLoS Biol. *5*, 2177–2189.

Su, L.F., Kidd, B.A., Han, A., Kotzin, J.J., and Davis, M.M. (2013). Virus-specific CD4(+) memory-phenotype T cells are abundant in unexposed adults. Immunity *38*, 373–383.

Sudo, N., Sawamura, S., Tanaka, K., Aiba, Y., Kubo, C., and Koga, Y. (1997). The requirement of intestinal bacterial flora for the development of an IgE production system fully susceptible to oral tolerance induction. J. Immunol. *159*, 1739–1745.

Sun, C.M., Hall, J.A., Blank, R.B., Bouladoux, N., Oukka, M., Mora, J.R., and Belkaid, Y. (2007). Small intestine lamina propria dendritic cells promote de novo generation of Foxp3 T reg cells via retinoic acid. J. Exp. Med. *204*, 1775–1785.

Suzuki, K., Meek, B., Doi, Y., Muramatsu, M., Chiba, T., Honjo, T., and Fagarasan, S. (2004). Aberrant expansion of segmented filamentous bacteria in IgA-deficient gut. Proc. Natl. Acad. Sci. USA *101*, 1981–1986.

Takeda, K., Kaisho, T., and Akira, S. (2003). Toll-like receptors. Annu. Rev. Immunol. 21, 335–376.

Talham, G.L., Jiang, H.Q., Bos, N.A., and Cebra, J.J. (1999). Segmented filamentous bacteria are potent stimuli of a physiologically normal state of the murine gut mucosal immune system. Infect. Immun. *67*, 1992–2000.

Tsuji, M., Komatsu, N., Kawamoto, S., Suzuki, K., Kanagawa, O., Honjo, T., Hori, S., and Fagarasan, S. (2009). Preferential generation of follicular B helper T cells from Foxp3+ T cells in gut Peyer's patches. Science *323*, 1488–1492.

Turnbaugh, P.J., Ley, R.E., Mahowald, M.A., Magrini, V., Mardis, E.R., and Gordon, J.I. (2006). An obesity-associated gut microbiome with increased capacity for energy harvest. Nature *444*, 1027–1031.

Turovskiy, Y., Sutyak Noll, K., and Chikindas, M.L. (2011). The aetiology of bacterial vaginosis. J. Appl. Microbiol. *110*, 1105–1128.

Ubeda, C., Bucci, V., Caballero, S., Djukovic, A., Toussaint, N.C., Equinda, M., Lipuma, L., Ling, L., Gobourne, A., No, D., et al. (2013). Intestinal microbiota containing Barnesiella species cures vancomycin-resistant Enterococcus faecium colonization. Infect. Immun. *81*, 965–973.

Uematsu, S., Jang, M.H., Chevrier, N., Guo, Z., Kumagai, Y., Yamamoto, M., Kato, H., Sougawa, N., Matsui, H., Kuwata, H., et al. (2006). Detection of pathogenic intestinal bacteria by Toll-like receptor 5 on intestinal CD11c+ lamina propria cells. Nat. Immunol. *7*, 868–874. Uematsu, S., Fujimoto, K., Jang, M.H., Yang, B.G., Jung, Y.J., Nishiyama, M., Sato, S., Tsujimura, T., Yamamoto, M., Yokota, Y., et al. (2008). Regulation of humoral and cellular gut immunity by lamina propria dendritic cells expressing Toll-like receptor 5. Nat. Immunol. 9, 769–776.

Umesaki, Y., Setoyama, H., Matsumoto, S., Imaoka, A., and Itoh, K. (1999). Differential roles of segmented filamentous bacteria and clostridia in development of the intestinal immune system. Infect. Immun. 67, 3504–3511.

Vaishnava, S., Behrendt, C.L., Ismail, A.S., Eckmann, L., and Hooper, L.V. (2008). Paneth cells directly sense gut commensals and maintain homeostasis at the intestinal host-microbial interface. Proc. Natl. Acad. Sci. USA *105*, 20858–20863.

Vaishnava, S., Yamamoto, M., Severson, K.M., Ruhn, K.A., Yu, X., Koren, O., Ley, R., Wakeland, E.K., and Hooper, L.V. (2011). The antibacterial lectin RegIllgamma promotes the spatial segregation of microbiota and host in the intestine. Science *334*, 255–258.

van der Waaij, D., Berghuis-de Vries, J.M., and Lekkerkerk Lekkerkerk-v. (1971). Colonization resistance of the digestive tract in conventional and antibiotic-treated mice. J. Hyg. (Lond.) 69, 405–411.

van Nood, E., Vrieze, A., Nieuwdorp, M., Fuentes, S., Zoetendal, E.G., de Vos, W.M., Visser, C.E., Kuijper, E.J., Bartelsman, J.F., Tijssen, J.G., et al. (2013). Duodenal infusion of donor feces for recurrent Clostridium difficile. N. Engl. J. Med. 368, 407–415.

Vernacchio, L., Vezina, R.M., Mitchell, A.A., Lesko, S.M., Plaut, A.G., and Acheson, D.W. (2006). Diarrhea in American infants and young children in the community setting: incidence, clinical presentation and microbiology. Pediatr. Infect. Dis. J. 25, 2–7.

Viaud, S., Saccheri, F., Mignot, G., Yamazaki, T., Daillère, R., Hannani, D., Enot, D.P., Pfirschke, C., Engblom, C., Pittet, M.J., et al. (2013). The intestinal microbiota modulates the anticancer immune effects of cyclophosphamide. Science 342, 971–976.

Vijay-Kumar, M., Aitken, J.D., Carvalho, F.A., Cullender, T.C., Mwangi, S., Srinivasan, S., Sitaraman, S.V., Knight, R., Ley, R.E., and Gewirtz, A.T. (2010). Metabolic syndrome and altered gut microbiota in mice lacking Toll-like receptor 5. Science *328*, 228–231.

von Essen, M.R., Kongsbak, M., Schjerling, P., Olgaard, K., Odum, N., and Geisler, C. (2010). Vitamin D controls T cell antigen receptor signaling and activation of human T cells. Nat. Immunol. *11*, 344–349.

Wade, T.K., and Wade, W.F. (2008). Variable gene family usage of protective and non-protective anti-Vibrio cholerae O1 LPS antibody heavy chains. Microbiol. Immunol. *52*, 611–620.

Wannemuehler, M.J., Kiyono, H., Babb, J.L., Michalek, S.M., and McGhee, J.R. (1982). Lipopolysaccharide (LPS) regulation of the immune response: LPS converts germfree mice to sensitivity to oral tolerance induction. J. Immunol. *129*, 959–965.

Wei, M., Shinkura, R., Doi, Y., Maruya, M., Fagarasan, S., and Honjo, T. (2011). Mice carrying a knock-in mutation of Aicda resulting in a defect in somatic hypermutation have impaired gut homeostasis and compromised mucosal defense. Nat. Immunol. *12*, 264–270. Cell

Weiner, H.L., da Cunha, A.P., Quintana, F., and Wu, H. (2011). Oral tolerance. Immunol. Rev. 241, 241–259.

Weinstock, J.V., Summers, R., and Elliott, D.E. (2004). Helminths and harmony. Gut 53, 7-9.

Wen, L., Ley, R.E., Volchkov, P.Y., Stranges, P.B., Avanesyan, L., Stonebraker, A.C., Hu, C., Wong, F.S., Szot, G.L., Bluestone, J.A., et al. (2008). Innate immunity and intestinal microbiota in the development of Type 1 diabetes. Nature 455, 1109–1113.

Winter, S.E., Winter, M.G., Xavier, M.N., Thiennimitr, P., Poon, V., Keestra, A.M., Laughlin, R.C., Gomez, G., Wu, J., Lawhon, S.D., et al. (2013). Hostderived nitrate boosts growth of E. coli in the inflamed gut. Science *339*, 708–711.

Wolk, K., and Sabat, R. (2006). Interleukin-22: a novel T- and NK-cell derived cytokine that regulates the biology of tissue cells. Cytokine Growth Factor Rev. *17*, 367–380.

Worbs, T., Bode, U., Yan, S., Hoffmann, M.W., Hintzen, G., Bernhardt, G., Förster, R., and Pabst, O. (2006). Oral tolerance originates in the intestinal immune system and relies on antigen carriage by dendritic cells. J. Exp. Med. 203, 519–527.

Wu, S., Rhee, K.J., Albesiano, E., Rabizadeh, S., Wu, X., Yen, H.R., Huso, D.L., Brancati, F.L., Wick, E., McAllister, F., et al. (2009). A human colonic commensal promotes colon tumorigenesis via activation of T helper type 17 T cell responses. Nat. Med. *15*, 1016–1022.

Wu, H.J., Ivanov, I.I., Darce, J., Hattori, K., Shima, T., Umesaki, Y., Littman, D.R., Benoist, C., and Mathis, D. (2010). Gut-residing segmented filamentous bacteria drive autoimmune arthritis via T helper 17 cells. Immunity *32*, 815–827.

Yatsunenko, T., Rey, F.E., Manary, M.J., Trehan, I., Dominguez-Bello, M.G., Contreras, M., Magris, M., Hidalgo, G., Baldassano, R.N., Anokhin, A.P., et al. (2012). Human gut microbiome viewed across age and geography. Nature 486, 222–227.

Yoshimoto, S., Loo, T.M., Atarashi, K., Kanda, H., Sato, S., Oyadomari, S., Iwakura, Y., Oshima, K., Morita, H., Hattori, M., et al. (2013). Obesity-induced gut microbial metabolite promotes liver cancer through senescence secretome. Nature *499*, 97–101.

Yurkovetskiy, L., Burrows, M., Khan, A.A., Graham, L., Volchkov, P., Becker, L., Antonopoulos, D., Umesaki, Y., and Chervonsky, A.V. (2013). Gender bias in autoimmunity is influenced by microbiota. Immunity 39, 400–412.

Zenewicz, L.A., and Flavell, R.A. (2011). Recent advances in IL-22 biology. Int. Immunol. 23, 159–163.

Zenewicz, L.A., Yin, X., Wang, G., Elinav, E., Hao, L., Zhao, L., and Flavell, R.A. (2013). IL-22 deficiency alters colonic microbiota to be transmissible and colitogenic. J. Immunol *190*, 5306–5312.

Zoumpopoulou, G., Foligne, B., Christodoulou, K., Grangette, C., Pot, B., and Tsakalidou, E. (2008). Lactobacillus fermentum ACA-DC 179 displays probiotic potential in vitro and protects against trinitrobenzene sulfonic acid (TNBS)-induced colitis and Salmonella infection in murine models. Int. J. Food Microbiol. *121*, 18–26.