

NIH Workshop Report: sensory nutrition and disease

Danielle R Reed,¹ Amber L Alhadeff,^{1,2} Gary K Beauchamp,¹ Nirupa Chaudhari,^{3,4,5} Valerie B Duffy,⁶ Monica Dus,⁷ Alfredo Fontanini,⁸ John I Glendinning,^{9,10} Barry G Green,^{11,12} Paule V Joseph,^{13,14} George A Kyriazis,¹⁵ Mark Lyte,^{16,17} Padma Maruvada,¹⁸ John P McGann,¹⁹ John T McLaughlin,^{20,21} Timothy H Moran,²² Claire Murphy,^{23,24} Emily E Noble,²⁵ M Yanina Pepino,^{26,27} Jennifer L Pluznick,²⁸ Kristina I Rother,²⁹ Enrique Saez,³⁰ Alan C Spector,^{31,32} Catia Sternini,³³ and Richard D Mattes³⁴

¹Monell Chemical Senses Center, Philadelphia, PA, USA; ²Department of Neuroscience, University of Pennsylvania, Philadelphia, PA, USA; ³Department of Physiology and Biophysics, University of Miami Miller School of Medicine, Miami, FL, USA; ⁴Department of Otolaryngology, University of Miami Miller School of Medicine, Miami, FL, USA; ⁵Program in Neurosciences, University of Miami Miller School of Medicine, Miami, FL, USA; ⁶Department of Allied Health Sciences, University of Connecticut, Storrs, CT, USA; ⁷Department of Molecular, Cellular, and Developmental Biology, University of Michigan, Ann Arbor, MI, USA; ⁸Department of Neurobiology and Behavior, Stony Brook University, Stony Brook, NY, USA; ⁹Department of Biology, Barnard College, Columbia University, New York, NY, USA; ¹⁰Department of Neuroscience and Behavior, Barnard College, Columbia University, New York, NY, USA; ¹¹The John B Pierce Laboratory, New Haven, CT, USA; ¹²Department of Surgery (Otolaryngology), Yale School of Medicine, Yale University, New Haven, CT, USA; ¹³National Institute of Alcohol Abuse and Alcoholism, NIH, Bethesda, MD, USA; ¹⁴National Institute of Nursing, NIH, Bethesda, MD, USA; ¹⁵Department of Biological Chemistry and Pharmacology, College of Medicine, The Ohio State University, Columbus, OH, USA; ¹⁶Interdepartmental Microbiology Graduate Program, Iowa State University, Ames, IA, USA; ¹⁷Department of Veterinary Microbiology and Preventive Medicine, Iowa State University, Ames, IA, USA; ¹⁸National Institute of Diabetes and Digestive and Kidney Diseases, NIH, Bethesda, MD, USA; ¹⁹Behavioral and Systems Neuroscience, Department of Psychology, Rutgers University, Piscataway, NJ, USA; ²⁰Division of Diabetes, Endocrinology, & Gastroenterology, School of Medical Sciences, Faculty of Biology, Medicine, and Health, The University of Manchester, Manchester, United Kingdom; ²¹Department of Gastroenterology, Salford Royal NHS Foundation Trust, Salford, United Kingdom; ²²Department of Psychiatry & Behavioral Sciences, Johns Hopkins University School of Medicine, Baltimore, MD, USA; ²³Department of Psychology, San Diego State University, San Diego, CA, USA; ²⁴Department of Psychiatry, University of California, San Diego, San Diego, CA, USA; ²⁵Department of Foods and Nutrition, University of Georgia, Athens, GA, USA; ²⁶Department of Food Science and Human Nutrition, University of Illinois at Urbana-Champaign, Urbana, IL, USA; ²⁷Division of Nutritional Sciences, University of Illinois at Urbana-Champaign, Urbana, IL, USA; ²⁸Department of Physiology, Johns Hopkins University School of Medicine, Baltimore, MD, USA; ²⁹Intramural Research Program, National Institute of Diabetes and Digestive and Kidney Diseases, NIH, Bethesda, MD, USA; ³⁰Department of Molecular Medicine, The Scripps Research Institute, La Jolla, CA, USA; ³¹Department of Psychology, Florida State University, Tallahassee, FL, USA; ³²Program in Neuroscience, Florida State University, Tallahassee, FL, USA; ³³Digestive Disease Division, Departments of Medicine and Neurobiology, David Geffen School of Medicine, University of California at Los Angeles, Los Angeles, CA, USA; and ³⁴Department of Nutrition Science, Purdue University, West Lafayette, IN, USA

ABSTRACT

In November 2019, the NIH held the “Sensory Nutrition and Disease” workshop to challenge multidisciplinary researchers working at the interface of sensory science, food science, psychology, neuroscience, nutrition, and health sciences to explore how chemosensation influences dietary choice and health. This report summarizes deliberations of the workshop, as well as follow-up discussion in the wake of the current pandemic. Three topics were addressed: A) the need to optimize human chemosensory testing and assessment, B) the plasticity of chemosensory systems, and C) the interplay of chemosensory signals, cognitive signals, dietary intake, and metabolism. Several ways to advance sensory nutrition research emerged from the workshop: 1) refining methods to measure chemosensation in large cohort studies and validating measures that reflect perception of complex chemosensations relevant to dietary choice; 2) characterizing interindividual differences in chemosensory function and how they affect ingestive behaviors, health, and disease risk; 3) defining circuit-level organization and function that link and interact with gustatory, olfactory, homeostatic, visceral, and

cognitive systems; and 4) discovering new ligands for chemosensory receptors (e.g., those produced by the microbiome) and cataloging cell types expressing these receptors. Several of these priorities were made more urgent by the current pandemic because infection with sudden acute respiratory syndrome coronavirus 2 (SARS-CoV-2) and the ensuing coronavirus disease of 2019 has direct short- and perhaps long-term effects on flavor perception. There is increasing evidence of functional interactions between the chemosensory and nutritional sciences. Better characterization of this interface is expected to yield insights to promote health, mitigate disease risk, and guide nutrition policy. *Am J Clin Nutr* 2021;113:232–245.

Keywords: olfaction, sweet, food preferences, food intake, liking

Introduction

The foods and fluids a person ingests can satiate, nourish, and promote growth. They can also cause harm—either within

minutes or hours after swallowing or after longer periods of intermittent ingestion (1). The chemosensory receptors of the nose, mouth, and throat provide the brain with information about the composition of foods and fluids, which in turn influences the probability of ingestion or rejection (2). For this reason, the chemical senses of taste and smell, together with chemesthesis—the chemical sensitivity of the somatosensory system—play a role in body weight and nutritional state (3).

On 12–13 November, 2019, the NIH held the “Sensory Nutrition and Disease” Workshop (4) in Bethesda, Maryland, to engage a diverse group of basic science and clinical researchers working at the interface of sensory and nutrition sciences to explore the potential of chemosensory biology to influence food preferences, intake, and nutrition. Here, the workshop is summarized, identifying new research and approaches needed for understanding how the chemical senses ultimately influence nutrition and health. Such knowledge can be used to mitigate chronic disease risk and help develop interventions that promote healthier diets. This workshop summary also includes perspectives on priorities emerging from the coronavirus disease of 2019 (COVID-19) pandemic (5).

Three main topic areas were identified by workshop participants: A) the need to optimize human chemosensory testing and assessment, B) the plasticity of chemosensory systems, and C) the interplay of chemosensory signals, dietary intake, and metabolism. These topics highlight current overarching questions at the interface of taste, smell, and food choice. Eleven gaps and opportunities were identified in our current understanding of how chemosensory biology influences nutrition and disease, including those subsequently identified by the effects of COVID-19.

Organization of the Chemosensory Systems

Each chemosensory system provides unique information to the brain. The olfactory system responds to thousands of different types of airborne molecules, whereas the gustatory system responds to a more limited set of chemicals in food and beverages, such as salts, sugars, amino acids, alkaloids, acids, and fats. The oronasal trigeminal system responds to chemicals in either volatile, liquid, or solid form via chemically sensitive receptors of the somatosensory system—a sensitivity referred to as chemesthesis (6). The signals from these different chemosensory systems interact [e.g., (7)] to form a flavor percept. This is the term for a composite perceptual integration of taste, smell, and chemesthesis and oral somatosensation, including food texture and temperature (8, 9).

The workshop was supported by the National Institute of Diabetes and Digestive and Kidney Diseases.

Where authors are identified as personnel of the NIH or other federal agencies or entities, the authors alone are responsible for the views expressed in this article, and they do not necessarily represent the decisions, policy, or views of the NIH or other entities.

Address correspondence to DRR (e-mail: reed@monell.org).

Abbreviations used: CCK, cholecystokinin; COVID-19, coronavirus disease of 2019; GLP-1, glucagon-like peptide 1; RNAseq, RNA sequencing.

Received June 11, 2020. Accepted for publication September 30, 2020.

First published online December 9, 2020; doi: <https://doi.org/10.1093/ajcn/nqaa302>.

Although many factors influence dietary choice and consumption, flavor is a primary driver of the amount and type of food a person or other animal chooses to eat. Sensory nutrition is studied in controlled environments with model organisms, in laboratory settings with human participants and direct or indirect measures of intake and health, and eventually translated into community and population-based studies to test the generalizability of laboratory-based findings.

Topics in Sensory Nutrition

Although there has been progress in understanding smell, taste, and chemesthesis, distillation of the presentations and discussion from the “Sensory Nutrition and Disease” Workshop highlighted 3 interconnected topic areas that warrant additional research (see **Table 1**): 1) optimizing chemosensory testing with whole foods to complement research of simpler taste and smell stimuli; 2) plasticity in smell, taste, and chemesthesis systems and effects on sensory and hedonic behaviors and responses; and 3) the interplay of chemosensory signals, dietary intake, microbiome, and metabolism.

Three consistent themes emerged from all 3 topic areas. One theme was the historic reliance on simple chemosensory stimuli, such as single odorants or taste compounds dissolved in water, to study complex behavioral and physiological responses. These stimuli bear little relation to consumption of real-world foods, which integrate taste and smell and other sensory inputs into a compositive flavor experience. Another consistent theme was how the ability to taste and smell (especially in the experience of pleasure) changes with life events, including development and aging; illnesses such as bacterial or viral illness, including COVID-19 (10); age-related conditions (e.g., chronic health conditions, neurodegenerative disorders, polypharmacy); and diet [e.g., (11)]. The third involves interactions between sensation, hedonic value, and feeding state, for example, how the hedonic value of a food increases when a person or animal is hungry. These themes link each of the 3 topic areas discussed below.

Optimizing chemosensory testing in sensory nutrition paradigms

With notable exceptions [e.g., (12–14)], most human research on the contribution of oronasal sensory signals to flavor and food preferences has largely relied on psychophysical studies with limited taste stimuli (15, 16) or noncommercial stimuli that provide investigators with better experimental control but limited ecological relevance. The dearth of research using real-world foods and beverages has impeded our understanding of flavor perception (17) and highlights the need for cross-talk between sensory psychologists, nutritionists, and food scientists for new research paradigms. As an example, using orally sampled real-world foods could capture relevant orthonasal and retronasal function (18), which could be used to better understand food pleasure (19) and dietary intake (20). Likewise, the flavor quality of low-calorie sweeteners differs depending on the type of food or beverage sweetened (21). New knowledge in this area may aid efforts to reformulate foods to make them

TABLE 1 Sensory nutrition and disease: research topics and suggestions¹

Category	Description
Research topics	A) Optimizing human chemosensory testing and assessment B) Boundaries and mechanisms of chemosensory plasticity C) Interplay of chemosensory signals, food dietary intake, and metabolism
Research opportunities	1) Chemosensory and hedonic biomarkers for food intake and food-related diseases 2) Multimodal evaluation of the response to food 3) Automating and standardizing methods to measure chemosensory behaviors 4) Connecting individual differences in genetics and experience of taste, smell, and ingestive behaviors 5) Clinical research in sensory nutrition 6) Capacity building for the study of big data in the sensory nutrition realm 7) Tracing neural circuits among peripheral chemosensory cells, gut, and brain across model organisms 8) Effects of chemosensory stimuli on visceral taste receptors 9) Microbiome and sensory receptor interactions 10) Deorphanization of chemosensory receptors, especially with nontraditional ligands 11) Single-cell RNAseq to define cell types that express chemosensory receptors
COVID-19	i) Loss of taste and smell as a cardinal feature and future research directions

¹COVID-19, coronavirus disease of 2019; RNAseq, RNA sequencing.

healthier (e.g., reduce sugar) and more palatable (e.g., reduce bitterness).

New research approaches that enable systematic manipulation and measurement of sensory properties of foods and beverages are necessary to understand the contributions of flavor to the mechanisms of dietary choice. The development of defined but ecologically relevant food stimuli will pose technical challenges and complicate experimental designs and analyses, including for translation outside of laboratory settings. To meet these challenges, interdisciplinary collaborations are needed among researchers with expertise in food science and technology, the chemical senses, oral somesthesia, psychology, nutrition, and public health.

Mechanisms of chemosensory plasticity

One burgeoning theme in chemosensory neurobiology is that chemosensory systems are highly plastic. For example, experience-dependent changes can occur at all levels of the olfactory system, including the olfactory epithelium. These changes include basic gain-control functions, where the system adjusts to environments with more or fewer odors (22–25). They also include strong associative plasticity (26). In animal models, pairing odors with strong positive (liked) or negative (disliked) stimuli causes radical changes in the numbers of neurons responsive to those odors in the olfactory epithelium (27, 28). In addition, stimulus–odor pairing causes alterations in the odor-evoked activity of the olfactory nerve (29, 30) and changes in the firing of neurons in the olfactory bulb (31–33), olfactory cortices (34, 35), and beyond. Similar experiences have been confirmed to induce perceptual changes in humans (36–38).

The taste system is also plastic (39–44) and changes over time (45, 46), for example, with age (47–49), as a result of illness (50, 51), or with changes in diet (52). These changes may affect individual eating habits. For example, it is commonly believed that eating a low-sugar diet makes people more sensitive to sucrose (53), which may in turn reduce the liking for high-sugar foods. Testing this idea experimentally is topical because of the public health pressure to reduce intake of sugar (as well as

salt and fat), with the expectation that people will acclimate over time, with foods with lower salt, sugar, or fat eventually becoming more palatable or even preferred. In humans, the best-studied taste effect based on dietary change is salt reduction: when people adopt a diet lower in sodium, they gradually adjust and come to prefer lower levels of saltiness (54). The same has been shown for fat (55, 56). Similar studies to evaluate the effects of low-sugar diets are under way.

Similarly, experience-dependent changes in liking and preference for hot and spicy foods are well known (57) but not well understood. The lack of understanding no doubt is due to the involvement of multiple factors (58), ranging from desensitization of chemesthetic receptors that occurs with frequent exposure (59) to personality variables (60). In addition, the possible contribution of postingestive nutrient effects to increases in liking of hot and spicy foods over time, such as flavor-nutrient conditioning (61), has not been investigated.

When studying the plasticity of the chemosensory systems, a few methodological details warrant particular attention. First, using sodium reduction as an example, all studies carried out to date have used abrupt reductions in salt consumption, whereas a gradual reduction of salt consumption is recommended (54). Thus, dietary change studies need to consider the rate of change as well as the final magnitude of reduction. Another aspect of experimental design is choice of outcome measures. In addition to taste and the habitual aforementioned intake and choice measures, other outcome measures usually comprise body weight and/or other measures related to metabolic disease (e.g., fasting or postprandial plasma glucose concentration). However, building on animal studies, these measures should be expanded to include less obvious but equally important outcomes, such as cognitive performance (62–66). Another consideration is the period of study. For example, studies of hedonic shifts indicate they require 8–12 wk to manifest and may be reversible in similar time spans. Similarly, changes in taste preferences after bariatric surgery are a model system for human taste plasticity, but the period of measurement has been limited to several months, and it is unclear whether favorable changes postsurgery persist beyond that brief window of study (67). In contrast, viral and bacterial infections, which are among the most common causes of taste and

smell disorders, may evoke rapid changes (68), requiring methods sensitive to the detection of short-term shifts. Such infections may also result in longer-term chemosensory dysfunction, so longitudinal studies are a needed area of research.

The inability to assess the response of the chemosensory cells and nerves to stimuli has been an obstacle in understanding the mechanisms of chemosensory plasticity and its implications for food choice and intake. However, studies in animal models from fruit flies to rodents have quantified the responses of the taste buds and the sensory neurons to stimuli in animals exposed to varying diet compositions. These studies have shown that exposure to diets high in sugar and fat decreases or alters the physiological responses of the taste buds and sensory neurons to stimuli or changes their number (39–44, 69), and 2 recent studies were able to uncouple the effects of weight gain from those of diet composition (42, 44) on the taste system. Further research in model organisms will capitalize on new methods to study old questions about chemosensory plasticity and rapidly generate hypotheses to be tested in human studies.

The interplay of chemosensory signals, dietary intake, and metabolism

Chemosensory plasticity is intertwined with endocrine responses because metabolism is in constant flux in response to meals, as well as longer-term dietary and lifestyle changes. There is a bidirectional relation between metabolism and chemical sensing (70): taste, smell, and chemesthesis affect what animals choose to eat, and what animals eat may influence their chemosensory sensitivities. The nutritional status of an animal or human modulates the relevance and valence of sensory stimuli (71, 72).

The influence of feeding state on taste, smell, and eating may be a key driver of ingestive behavior. As an example, the olfactory epithelium and olfactory bulb express metabolic signaling molecules and receptors, including orexins, ghrelin, neuropeptide Y, insulin, leptin, glucagon-like peptide 1 (GLP-1), cholecystokinin (CCK), and cannabinoid receptors 1 and 2 (73). Caution in interpretation is needed, however, because although these molecules play a role in some cells and tissues as a metabolic signal, here they may only be neuromodulators involved in synaptic transmission at different brain sites underlying functions that have nothing to do with metabolism. However, insulin and leptin both increase spontaneous activity and decrease odor-evoked activity in olfactory sensory neurons in the olfactory epithelium, which points to a direct role in metabolism (74). The activity of mitral cells, a type of neuron in the olfactory bulb, is influenced by insulin, glucose, and GLP-1 (71, 72, 75, 76). The olfactory bulb receives orexin-expressing projections from the lateral hypothalamus (77, 78), whereas some mitral cells from the bulb project to the arcuate nucleus (79). The olfactory bulb and cortex are also interconnected with affective and motivational centers that can play a role in eating behavior, including the amygdala, ventral hippocampus, and orbitofrontal cortex (80–92). In free-living animals and humans, the neural signals are not linear or immutable; context may modulate each step. For example, sweet taste stimuli typically have a positive valence and trigger a feeding response, yet satiety may dampen or even eliminate feeding by modulating central pathways (93–95). In turn, changes in the central processing of sensory stimuli could

affect the early steps of the satiety cascade and result in alterations in food intake (96, 97).

The interactions between sensory systems and metabolism may also occur directly in the gut. What we eat may influence gut metabolism acting through chemoreceptors. For example, some bitter taste receptors are upregulated in gut mucosal chemoreceptor cell subtypes of people with obesity and in the gut mucosa of mice with high-fat diet-induced obesity (98, 99), suggesting an interaction with diet or diet-induced intraluminal changes. Also, the chemicals produced by the gut, especially by the microbiome, may affect the brain (100) and may even result in changes in food preferences and choice behavior. Finally, metabolic signaling pathways could act on cells autonomously to directly modulate the responsiveness of the chemosensory neurons, as has been shown in invertebrates (42, 101). Overall, understanding the ability to sense foods via chemosensory receptors in response to metabolic changes may help inform disease treatment and suggest directions for public health policy relating to food and nutrition.

Specific Suggestions to Address Gaps in Sensory Nutrition Research

This section describes specific suggestions to address current challenges and gaps in sensory nutrition research. In particular, we focus on methods or approaches that are expected to improve the tempo and nature of research in the aforementioned topic areas. These suggestions, summarized in Table 1, are organized hierarchically, starting with the study of whole animals (e.g., behavioral measures), followed by tissues and circuits, and then individual cells.

Chemosensory and hedonic biomarkers for food intake and food-related diseases

People differ in their ability to smell and taste, owing to such factors as genetics, nutritional state, and age. Effort has been made to translate time-consuming, classical psychophysical tests into brief measures to study individual differences in chemosensation (102, 103), including those designed to measure taste preferences and taste-related behaviors (104). However, chemosensory and hedonic measures are needed that are useful, feasible, and represent complex flavor sensations for population-based studies, to allow us to generalize findings from experimental and clinical studies to the general population. Summaries of a number of population-based studies that have measured smell and taste are provided in Tables 2 and 3, respectively. In the United States, brief and standardized measures of smell and taste from the NIH Toolbox were further standardized and included in the 2011–2014 NHANES for adults ≥ 40 y of age (105). NHANES is a continuous, cross-sectional, multifaceted, nationally representative assessment of the health and diet of the US population, collected via in-home visits and mobile examination centers. The 2011–2014 data provide an opportunity to examine the strength of taste and smell associations across a broad array of diseases and conditions (e.g., cardiovascular, diabetes, kidney disease, obesity, oral health, and respiratory tract), environmental exposures, and behaviors (e.g., dietary, physical activity).

TABLE 2 Major population-based studies with olfactory data [modified from Yang and Pinto (106)]¹

Name	Country	Years	Design	Sample	Measure	Brief description of outcome	Reference
Betula Honolulu-Asia Aging	Sweden	1988–2014 ^a	Long	Healthy adults	ID	Dysfunction correlated with cognition	(107)
	USA	1991–2012	Long	Older men	ID	75% dysfunction; dysfunction as a risk factor for future PD	(108)
Rush Memory & Aging Project	USA	1997–2014	Long	Cohort of older adults	ID	55% olfactory impairment; dysfunction correlated with AD	(109)
Epidemiology of Hearing Loss Study, Beaver Dam	USA	1998–2000	Long	Older adults in 1 US community	ID	24.5% impairment; increased risk with sinonasal disease, smoking, respiratory infection	(110)
Skövde population-based study	Sweden	2001	CS	Nationally representative adults	ID	19.1% dysfunction; increased risk with nasal polyps	(111)
Blue Mountain Eye Study	Australia	2002–2004	CS	Sydney residents	ID	Dysfunction related to BMI (in kg/m ²), cognition	(112)
Washington Heights/Inwood Columbia Aging Project	USA	2004–2010 ^b	Long	Older adults in 1 US community	ID	Dysfunction as biomarker of cognitive decline and AD dementia	(113)
National Social Life, Health & Aging Longitudinal Kailuan study	USA	2005 ^c	Long	Nationally representative older adults	ID, Thsh	3% severe dysfunction; dysfunction strong predictor of mortality	(114)
KNHANES	China	2006	Long	Adults 18–98 y old; subset	SR	Alteration correlated with larger increases in BP	(115)
	South Korea	2009–2011	CS	Nationally representative	SR	5% dysfunction; dysfunction correlated with mental health	(116)
US NHANES	USA	2011–2014	CS	Nationally representative of adults ≥40 y old	ID, SR	12.4% dysfunction, 6.5% phantom odors	(105, 117)

¹ AD, Alzheimer disease; BP, blood pressure; CS, cross-sectional; ID, measured identification of odorants; KNHANES, Korean National Health and Nutrition Examination Survey; Long, longitudinal; PD, Parkinson disease; SR, self-report; Thsh, odorant threshold.

^aMeasured in the third wave; ^bmeasured in the first wave, continuing; ^cmeasured in the fourth wave, continuing.

TABLE 3 Sampling of population-based studies with taste data¹

Study	Country	Design	Sample	Measure	Brief description of outcome	Reference
Beaver Dam Offspring Study	USA	Long	1981 adults	Taste/papillae	Taste correlated with body weight changes	(118, 119)
National Social Life, Health and Aging Project	USA	Long	Nationally representative older adults	Taste strip identification	74% gustatory deficit	(120, 121)
Swedish 1942 birth cohort	Sweden	20-y cohort	6346 adults	Reported taste distortion	7%; distortions correlated with poor oral and overall health	(122)
SONIC	Japan	Long	621 older adults	Recognition threshold	Gender and cognitive effects on function	(123)
Dallas Heart Study/Dallas Biobank	USA	CS	> 10,000	PTC taste	Race and PTC effects on smoking	(124)
Genetically isolated village of Carliantino	Italy	CS	540 healthy inhabitants	PROP taste	Nontasters heavier than tasters; interaction with dietary restraint	(125)
UK Women's Cohort Study	United Kingdom	Long	5500 adult women	PTC taste	Tasters correlated with cancer risk	(126)
Silk Road	Asia	CS	496 adults across 6 countries	PROP taste	PROP related to food preference	(127)
Brisbane Longitudinal Twin Study	Australia	Long	1576 adolescents	Sweet intensity	Sweet-BMI association	(128)
IDEFICS cohort	Europe	PCS	1861 children	Taste/papilla/ preference	Preferences associated with body weight	(129)
Generation R Study	Netherlands	Birth cohort	5585 children	PROP taste	Few PROP-diet/food associations; PROP-body weight associations	(130)
US NHANES	USA	CS	Nationally representative adults ≥40 y old	Bitter/salt intensity—whole mouth and regional	5.24% dysgeusia	(49, 105)

¹CS, cross-sectional; IDEFICS, Identification and Prevention of Dietary- and Lifestyle-Induced Health Effects in Children and Infants; Long, longitudinal; PCS, prospective cohort study; PROP, propylthiouracil; PTC, phenylthiocarbamide; SONIC, Septuagenarians, Octogenarians, Nonagenarians Investigation with Centenarians.

The majority of smell tests measure odorant identification (e.g., amyl acetate smells like a banana), which often relates to cognitive function and may not be a sensitive method to understand how smell function affects food intake. In contrast, many taste tests measure intensity responses to a single compound or quality, such as bitterness evoked by propylthiouracil or phenylthiocarbamide. These compounds are often used because of the wide range of responses they evoke (131, 132), but broadening taste testing to other compounds representing other taste qualities and mixtures of compounds to reflect the flavor percept would enrich knowledge of sensory nutrition as it relates to short-term (e.g., viral infection) and chronic (e.g., diabetes) disease. Of particular importance is work on hedonic responses to tastants, because this facet of chemosensory function is probably the strongest determinant of food choice (133).

There are several benefits to developing validated chemosensory measurement tools that are faster and more reliable than current methods as well as standardizing current methods so that results of large-scale studies can be compared directly. In addition, methods are needed to facilitate collection and analysis of large amounts of data [e.g., the All of Us precision medicine initiative and related efforts (105, 134)] to enrich hypothesis generation.

Overall, there is a critical need to optimize chemosensory testing that is feasible for population-based studies and that has relevance to dietary behaviors. Such olfactory and taste methods could produce measures that become biomarkers of long-term dietary behaviors, which, in turn, would enhance understanding of the connections between diet and chronic disease (135).

Multimodal evaluation of the response to food

Taste and smell have most often been studied in isolation, but their interaction is essential to understand how flavor affects consumption (136), especially hyperpalatability (137) or the idea that some foods [e.g., combinations of fat and sugar (138)] are so hedonically rewarding that overconsumption is inevitable. Moreover, chemesthesis and somatosensory (texture) qualities of flavor also play a role in palatability, food selection, and consumption.

Thus, sensory testing needs to integrate the myriad sensory qualities of foods and beverages. Filling this gap in the field will require interdisciplinary research that considers taste, smell, chemesthesis, vision, audition, and somatosensation not as separate contributors but as essential components of the flavor gestalt that guides food selection and intake.

Automating and standardizing methods to measure chemosensory behaviors

Animals and humans alike have characteristic behaviors that are objective signs of food and drink enjoyment or rejection (139), but more effort is needed to connect common methods of measuring these behaviors in animals (licking rate, facial expressions, sniffing) with corresponding human behaviors. Facial expressions are an example of how to forge these connections [e.g., (139)], but most human studies rely on food intake and verbal reports of liking, preference, or intensity. Thus, validated and reliable behavioral measures that have direct

correlates between humans and animals would enhance research in this area.

Ideal methods for monitoring and automatically classifying behavior, such as oral movements (licking and swallowing) and feeding behavioral assays, will be practical and compatible with other types of measurement, such as brain function using functional MRI and optical neurophysiological methods (140–142). In humans, neuroimaging methods, particularly functional and structural MRI, have the potential to reveal nutritional effects on the neural substrates of taste, smell, and ingestion. Application of neuroimaging methods to understanding brain mechanisms underlying eating behavior in populations at risk of earlier morbidity and mortality, such as those people who have obesity or diabetes, will be of particular interest (143, 144). The development of these methods should serve as a translational bridge, logically linking findings between animal models and humans.

Connecting individual differences in genetics and experience to taste, smell, and ingestive behaviors

People differ in how they perceive and respond to the sight, smell, and taste of food. For example, there are well-studied person-to-person differences in bitter perception (145), salivary composition (146), and cephalic-phase insulin responses to the sensory properties of food (147). Studies are needed to determine whether this variability reflects heritable differences across subjects, experience-induced plasticity, or their interaction and whether these effects are larger for certain flavors and tastes such as sweetness and bitterness.

Historically, the extent of genetic compared with other influences was determined by comparing biological relatives—often twins, in the case of humans (148), and inbred, selectively bred, and hybrid strains in animal models (149, 150). More recently, genome-wide association studies, which use large numbers of unrelated people, have inferred heritability by the association of a trait (e.g., the consumption of particular foods or foods with a specific taste quality) with a genetic variant (151, 152). As our knowledge of associations between food-related behaviors expands (153), genotype becomes a standard variable to consider (similarly to age or sex) when assessing taste, smell, chemesthesis, and somatosensation and their relations to food intake.

Clinical research in sensory nutrition

More than 200,000 people visit a doctor each year for problems with their chemical senses (154). Furthermore, just over 1 in 5 adults in the United States with a smell or taste disorder sought medical care for this disorder according to data from the 2014–2016 NHANES and the baseline for one of the HealthyPeople 2030 goals (155). Clinical studies of the chemesthetic senses are needed because impairments in these senses have important implications for health and quality of life. Few clinical studies have assessed chemosensory measures or clinical guidelines for treatments, and, as aforementioned, we need to optimize rapid, standardized measures for the clinical setting that could extend to nonclinical settings. In addition, existing treatment strategies for taste and smell disorders are limited and often ineffective

(156). More studies are also needed to connect chemosensory alterations to other clinical disorders such as diabetes, obesity, and cancer, and we need increased support for chemosensory clinician-scientists to foster interdisciplinary collaborations.

Building capacity to facilitate the use of big data in the sensory nutrition realm

Very large data sets are being generated that could expand our understanding of sensory–nutrition interactions. Some examples are the UK Biobank (157), the Million Veteran Program (158), and All of Us (159), which are large-scale efforts to study hundreds of thousands to millions of people through health records, surveys, biological samples, and other physiological indexes, as well as genetic data. Studies based on these data sources could further inform us about how flavor affects food choice and how variation in taste and smell receptor genes affects human health.

Thus, new large-scale collaborative initiatives and training programs in artificial intelligence and data science are needed for chemosensory scientists in the biomedical workforce. It will also be important to offer retraining programs for established scientists hoping to gain skills in these new computational areas. Finally, support for interdisciplinary teams is needed to fully mine the chemosensory data and direct analyses of chemosensory-related genes.

Tracing neural circuits among peripheral chemosensory cells, gut, and brain across model organisms

The interplay of taste, smell, chemesthesis, somatosensation, and digestion arises from neural connections between the nose, oral cavity, gut, and brain. One example of this interplay is the cephalic-phase response, which arises when food-related sensory inputs stimulate centers in the brainstem (e.g., the dorsal motor vagal nucleus), which in turn initiate parasympathetic and/or sympathetic response pathways. Another example of this interplay is between nutrient-sensing responses in the gut and the brain–gut axis connections. However, the nature of the effective stimuli, afferent pathways to the brainstem, integrating centers in the brainstem, and efferent pathways to the periphery remain largely unknown. Thus, knowledge about the interaction between the cephalic chemosensory systems, the viscera, and the gustatory system would close this knowledge gap concerning neural circuits. A better understanding of this neural circuitry in different model organisms would inform the interplay of chemosensation, food intake, and metabolism.

One challenge to the study of chemosensory brain circuitry in mammals is its unfavorable anatomy. For instance, gustatory areas pose unique challenges to their study in rodent models. Subcortical and cortical regions are small, cytoarchitecturally heterogeneous, and located in inaccessible anatomical positions (95). Thus, the anatomy of the taste system makes it hard to measure the responses of specific types of taste cells to stimuli in awake, behaving animals. Development of multisite neural recording methods and stimulation approaches suitable for the taste-related brain areas is critical to delineate the complex interconnectivity of the gustatory system with central circuits. Further, it will be important to continue leveraging the advances in knowledge brought by studies in invertebrate model organisms,

with research informed by hypotheses relevant to and formulated from human and preclinical models.

Effects of chemosensory stimuli on visceral taste receptors

Many chemosensory receptors are expressed in the gut and viscera, and their function has been only partially characterized. Current data suggest that these sensory pathways contribute to nutrient digestion and metabolism, and ultimately to obesity, diabetes, and other diet-related diseases. For example, hyperglycemia downregulates sweet taste receptor expression in β -cells, leading to compensatory alterations in insulin secretion in obese and diabetic mouse models (160). Likewise, in patients with type 2 diabetes, intestinal sweet taste receptor mRNA levels are inversely correlated with fasting glycemia (161). These sweet taste receptors are dysregulated in response to high luminal glucose (162) and by acute high-sucrose feeding (163). These observations suggest that taste receptors may be, in part, responsible for adaptive responses to imbalances in nutrient availability (e.g., diabetes and refined carbohydrate intake, coronary artery disease, and high saturated fat intake) and signal the need for additional research to explore the function of chemosensory receptors in the viscera.

Microbiome and sensory receptor interactions

The role of the microbiota in health and disease is ever-expanding, with its application to sensory nutrition inevitable at least in part because the microbiota in the upper airway and gastrointestinal tract, starting with the tongue, are a rich source of chemical signals. One new research avenue is the study of sometimes subtle changes in the diet and how they affect the gut microbiome [e.g., (164, 165)], perhaps reshaping it over generations. For example, rats selectively bred for high sweet preference reliably differ in the pattern of their gut microbiota compared with rats bred for low sweet preference (150). Another avenue of research is to study microbes that can produce chemicals that influence behavior (166–169) and to determine whether chemosensory bitter taste receptors detect these signals in the gut (as they do in the nose) (170). These chemical signals could be similar to or the same as neurochemicals produced by the host and affect host behavior (100, 171), for example, by getting the host to eat more of the type of food that members of the gut microbiota can easily metabolize (172). The nose and tongue also contain a diverse microbiome (173, 174), which may affect taste and smell directly. Currently, most studies of the microbiome are correlative, but the challenge ahead is to test these microbiome–host interactions experimentally. Another challenge is to have hypotheses in advance of the analysis of the very large data sets generated by most microbiome sequencing studies and to rapidly incorporate new knowledge about how to translate raw DNA sequencing data into more accurate descriptions of the microbiome [e.g., (175)].

Deorphanization of chemosensory receptors, especially with nontraditional ligands

Taste and smell receptors are part of a large family of G-protein-coupled receptors that sense extracellular chemicals and

initiate responses that result in conscious perception. However, many of these receptor proteins selectively respond to ligands that are not traditional taste or smell stimuli, including those produced in the host's own body, such as metabolites, bacterial metabolites, hormones, and neurotransmitters. Likewise, proteins identified in other tissues because of their role in metabolite sensing (e.g., sugar transporters) can act as noncanonical taste sensors (176).

Ecnotopic taste and olfactory receptors continue to be identified, but their significance often remains unknown. For example, a mouse bitter receptor has been reported to abide in gastric parietal and chief cells (177), and a human bitter taste receptor has been described in subtypes of enteroendocrine cells of the colon, including GLP-1, peptide YY, and CCK cells (98). Another bitter taste receptor has been observed in epithelial cells of the rodent small intestine (178), and yet another bitter receptor has been described in mouse Paneth cells and goblet cells (179, 180). Their functional roles in these locations have yet to be characterized.

This broad distribution of receptors in various cell types across the gut supports the concept that these chemosensory receptors serve as modulators of several functions, such as glucose homeostasis, gut motility and secretion, nutrient sensing, and secretion of hormones with purported appetitive properties (181, 182). Identifying the ligands of these receptors and sensors may uncover novel therapeutic opportunities. For instance, treatment with selective bitter agonists might target intestinal receptors and constitute a nonsurgical approach with the same benefits as bariatric surgery. However, these types of therapeutic opportunities will be easier to accomplish if the entire receptive range of these receptors is known, rather than focusing on classic taste and smell ligands (183, 184). Studies to identify this broad range of ligands also need to include those for receptors of model organisms, especially rodents, because there is often limited conservation of receptors between humans and model organisms (185). For preclinical animal studies to predict effects in humans, a functional human counterpart—in both its response to the specific compound of interest and its pattern of tissue expression—needs to be identified.

Thus, research programs to match both canonical and non-canonical chemosensory receptors to ligands should expand to include ligands beyond classic taste and smell stimuli (e.g., metabolites, hormones) and to include model organisms (e.g., fly, worm, mouse) in addition to human receptors. This new knowledge would pave the way for potent and selective pharmacological tools to study these receptors and their potentially broad role in human health and disease.

Single-cell RNA sequencing to define cell types that express chemosensory receptors

Chemosensory receptors were originally identified and characterized in taste and olfactory receptor cells, but how many additional cell types contain these receptors and their function in these different cell types are unknown. As is often the case with receptor proteins, the low expression of chemosensory receptors or their expression in a few rare cell types within a tissue hinders their detection, and they are seldom found in global RNA sequencing (RNAseq) analyses (186) or in proteomic studies (187).

Single-cell RNAseq methods provide a more sensitive approach to identify the cell-specific expression of taste and

olfactory receptors throughout the body. Analysis of even a single tissue using these methods reveals many more cell types than previously identified [e.g., (188)] and allows investigators to construct more informed hypotheses to test their function. Large-scale efforts to examine the cellular composition of common tissues are under way (189), but taste, smell, and chemesthetic tissues are often ignored by large-scale projects [for an exception, see (190)]. These single-cell studies would be most useful with a complementary effort to get validated antibodies to confirm the presence of certain markers in particular sensory cell types. Olfactory receptor proteins are hard to measure (187) and validated antibodies for chemosensory cells are often lacking (191). The goal of defining the cell types that express chemosensory receptors is made more urgent by an emerging appreciation that receptor functions can differ based on the number and type of chemosensory receptors expressed in a given cell type (192). Therefore, methods that define and validate taste and olfactory receptor cell types are a current priority.

COVID-19 and Sensory Nutrition: Gaps and Opportunities

Several months after the NIH conference was held, the US CDC declared an outbreak of SARS-CoV-2 (i.e., sudden acute respiratory syndrome coronavirus 2), which leads to COVID-19, a pandemic (5). Abrupt loss of taste and smell is among the most reliable predictors of COVID-19 (193, 194), which highlights the urgent need for practical tests of chemosensory loss that are standardized, valid, and reliable (195). Taste and smell loss with COVID-19 is fully or partially regained, but it is currently unclear what the long-term consequences will be on flavor perception, food preferences, food intake, and broader disease risks. Especially relevant are the high rates of parosmia (distortions of smell) or phantosmia (smelling something that is not present) in those with COVID-19 (10), which may have long-term consequences for nutritional health.

Summary

Understanding the determinants and consequences of an unhealthy diet is critical because it is the root of many chronic diseases, including obesity, diabetes, and heart disease. Similarly, disorders affecting chemosensory functions may compromise diet quality, predisposing individuals to health complications. To understand these complex relations, the study of sensory influences on nutrition will require the collective efforts of teams comprising diverse expertise in a wide range of fields, such as food scientists, nutritionists, psychologists, neuroscientists, geneticists, molecular biologists, microbiologists, statisticians, and computer scientists, working on multiple model organisms. Such multidisciplinary research teams will move the field forward by expanding markers, inputs, and methods to use in chemosensory and metabolism research. Establishing connections in big data, neural circuitry, and individual genetics and behaviors to identify the chemosensory stimuli, receptors, and cell types that contribute to food choice and metabolism will jumpstart this effort. Collecting robust, reproducible data will facilitate better prediction and mitigation of disease risk and guide sound nutrition policy.

We thank Christopher Lynch and the National Institute of Diabetes and Digestive and Kidney Diseases Office of Nutrition Research for organizing the Sensory Nutrition and Disease Workshop, and Kimberly Barch for her administrative support. We also thank John Hayes, Ann-Marie Torregrossa, Alissa Nolden, Stephen Roper, Jenifer Trachtman, Robert Margolskee, Susan Sullivan, Coryse St. Hillaire-Clarke, Nancy Rawson, and Dolly Al Koborssy for their input and participation during the workshop. Vicente Ramirez assisted in the revision of the manuscript.

The authors' responsibilities were as follows—all authors: contributed to writing and editing of the manuscript. GKB receives no personal funds, including speaker fees, from any commercial entity. Ajinomoto provides a consulting fee to the Monell Chemical Senses Center that is used to support a small portion of his research. His research on sweetness is supported by a competitive grant from the NIH. GKB is a member of the Board of Directors of International Life Sciences Institute North America. All other authors report no conflicts of interest.

References

1. Reed DR, Knaapila A. Genetics of taste and smell: poisons and pleasures. *Prog Mol Biol Transl Sci* 2010;94:213–40.
2. Breslin PA, Spector AC. Mammalian taste perception. *Curr Biol* 2008;18(4):R148–55.
3. Pfaffmann C. The sensory and motivating properties of the sense of taste. In: Jones MR, editor. *Nebraska Symposium on Motivation*. Lincoln, NE: University of Nebraska Press; 1961:71–110.
4. National Institute of Diabetes and Digestive and Kidney Diseases (NIDDK) Health Information Center. Sensory Nutrition and Disease Workshop [Internet]. Bethesda, MD: NIDDK; 2019 [cited 24 March, 2020]. Available from: <https://www.niddk.nih.gov/news/meetings-workshops/2019/sensory-nutrition-disease-workshop>.
5. CDC. Symptoms of coronavirus [Internet]. Atlanta, GA: CDC; 2020 [cited 5 June, 2020]. Available from: <https://www.cdc.gov/coronavirus/2019-ncov/symptoms-testing/symptoms.html>.
6. Green BG. Chemesthesis and the chemical senses as components of a “chemofensor complex”. *Chem Senses* 2012;37(3):201–6.
7. Guichard E, Barba C, Thomas-Danguin T, Tromelin A. Multivariate statistical analysis and odor–taste network to reveal odor–taste associations. *J Agric Food Chem* 2020;68(38):10318–28.
8. Small DM, Prescott J. Odor/taste integration and the perception of flavor. *Exp Brain Res* 2005;166(3–4):345–57.
9. Prescott J, Stevenson RJ. Chemosensory integration and the perception of flavor. In: Doty RL, editor. *Handbook of olfaction and gustation*. Hoboken, NJ: John Wiley & Sons; 2015:1005–25.
10. Parma V, Ohla K, Veldhuizen MG, Niv MY, Kelly CE, Bakke AJ, Cooper KW, Bouysset C, Pirastu N, Dibattista M, et al. More than smell. COVID-19 is associated with severe impairment of smell, taste, and chemesthesis. *medRxiv* 2020:2020.05.04.20090902.
11. Ebbeling CB, Feldman HA, Steltz SK, Quinn NL, Robinson LM, Ludwig DS. Effects of sugar-sweetened, artificially sweetened, and unsweetened beverages on cardiometabolic risk factors, body composition, and sweet taste preference: a randomized controlled trial. *J Am Heart Assoc* 2020;9(15):e015668.
12. Mattes RD, Mela DJ. Relationships between and among selected measures of sweet-taste preference and dietary intake. *Chem Senses* 1986;11(4):523–39.
13. Mattes RD. Gustation as a determinant of ingestion: methodological issues. *Am J Clin Nutr* 1985;41(4):672–83.
14. Dinehart ME, Hayes JE, Bartoshuk LM, Lanier SL, Duffy VB. Bitter taste markers explain variability in vegetable sweetness, bitterness, and intake. *Physiol Behav* 2006;87(2):304–13.
15. Drewnowski A. Taste preferences and food intake. *Annu Rev Nutr* 1997;17:237–53.
16. Grinker J. Obesity and sweet taste. *Am J Clin Nutr* 1978;31(6):1078–87.
17. Mattes RD. Flavor and feeding: introduction to an international conference. *Physiol Behav* 2012;107(4):467–8.
18. Boesveldt S, de Graaf K. The differential role of smell and taste for eating behavior. *Perception* 2017;46(3–4):307–19.
19. Lim J, Padmanabhan A. Retronasal olfaction in vegetable liking and disliking. *Chem Senses* 2013;38(1):45–55.

20. Duffy VB, Hayes JE, Sharafi M. Interactions between retronasal olfaction and taste influence vegetable liking and consumption: a psychophysical investigation. *J Agric Food Res* 2020;2:100044.
21. Tan VWK, Wee MSM, Tomic O, Forde CG. Rate-All-That-Apply (RATA) comparison of taste profiles for different sweeteners in black tea, chocolate milk, and natural yogurt. *J Food Sci* 2020;85(2):486–92.
22. Kass MD, Guang SA, Moberly AH, McGann JP. Changes in olfactory sensory neuron physiology and olfactory perceptual learning after odorant exposure in adult mice. *Chem Senses* 2016;41(2):123–33.
23. Kass MD, Moberly AH, Rosenthal MC, Guang SA, McGann JP. Odor-specific, olfactory marker protein-mediated sparsening of primary olfactory input to the brain after odor exposure. *J Neurosci* 2013;33(15):6594–602.
24. Brunjes PC. Unilateral naris closure and olfactory system development. *Brain Res Rev* 1994;19(1):146–60.
25. Stone DM, Wessel T, Joh TH, Baker H. Decrease in tyrosine hydroxylase, but not aromatic L-amino acid decarboxylase, messenger RNA in rat olfactory bulb following neonatal, unilateral odor deprivation. *Mol Brain Res* 1990;8(4):291–300.
26. McGann JP. Associative learning and sensory neuroplasticity: how does it happen and what is it good for? *Learn Mem* 2015;22(11):567–76.
27. Jones SV, Choi DC, Davis M, Ressler KJ. Learning-dependent structural plasticity in the adult olfactory pathway. *J Neurosci* 2008;28(49):13106–11.
28. Morrison FG, Dias BG, Ressler KJ. Extinction reverses olfactory fear-conditioned increases in neuron number and glomerular size. *Proc Natl Acad Sci U S A* 2015;112(41):12846–51.
29. Kass MD, Rosenthal MC, Pottackal J, McGann JP. Fear learning enhances neural responses to threat-predictive sensory stimuli. *Science* 2013;342(6164):1389–92.
30. Bhattarai JP, Schreck M, Moberly AH, Luo W, Ma M. Aversive learning increases release probability of olfactory sensory neurons. *Curr Biol* 2020;30(1):31–41.
31. Fletcher ML. Olfactory aversive conditioning alters olfactory bulb mitral/tufted cell glomerular odor responses. *Front Syst Neurosci* 2012;6:16.
32. Ross JM, Fletcher ML. Learning-dependent and -independent enhancement of mitral/tufted cell glomerular odor responses following olfactory fear conditioning in awake mice. *J Neurosci* 2018;38(20):4623–40.
33. Kass MD, McGann JP. Persistent, generalized hypersensitivity of olfactory bulb interneurons after olfactory fear generalization. *Neurobiol Learn Mem* 2017;146:47–57.
34. Funk D, Amir S. Enhanced Fos expression within the primary olfactory and limbic pathways induced by an aversive conditioned odor stimulus. *Neuroscience* 2000;98(3):403–6.
35. Chen CF, Barnes DC, Wilson DA. Generalized vs. stimulus-specific learned fear differentially modifies stimulus encoding in primary sensory cortex of awake rats. *J Neurophysiol* 2011;106(6):3136–44.
36. Li W, Howard JD, Parrish TB, Gottfried JA. Aversive learning enhances perceptual and cortical discrimination of indiscriminable odor cues. *Science* 2008;319(5871):1842–5.
37. Parma V, Ferraro S, Miller SS, Ahs F, Lundstrom JN. Enhancement of odor sensitivity following repeated odor and visual fear conditioning. *Chem Senses* 2015;40:497–506.
38. Ahs F, Miller SS, Gordon AR, Lundstrom JN. Aversive learning increases sensory detection sensitivity. *Biol Psychol* 2013;92(2):135–41.
39. Chen K, Yan J, Suo Y, Li J, Wang Q, Lv B. Nutritional status alters saccharin intake and sweet receptor mRNA expression in rat taste buds. *Brain Res* 2010;1325:53–62.
40. Maliphol AB, Garth DJ, Medler KF. Diet-induced obesity reduces the responsiveness of the peripheral taste receptor cells. *PLoS One* 2013;8(11):e79403.
41. Kaufman A, Choo E, Koh A, Dando R. Inflammation arising from obesity reduces taste bud abundance and inhibits renewal. *PLoS Biol* 2018;16(3):e2001959.
42. May CE, Vaziri A, Lin YQ, Grushko O, Khabiri M, Wang Q-P, Holme KJ, Pletcher SD, Freddolino PL, Neely GG, et al. High dietary sugar reshapes sweet taste to promote feeding behavior in *Drosophila melanogaster*. *Cell Rep* 2019;27(6):1675–85.e7.
43. Weiss MS, Hajnal A, Czaja K, Di Lorenzo PM. Taste responses in the nucleus of the solitary tract of awake obese rats are blunted compared with those in lean rats. *Front Integr Neurosci* 2019;13:35.
44. Ahart ZC, Martin LE, Kemp BR, Dutta Banik D, Roberts SGE, Torregrossa AM, Medler KF. Differential effects of diet and weight on taste responses in diet-induced obese mice. *Obesity* 2020;28(2):284–92.
45. Sylvestsky AC, Conway EM, Malhotra S, Rother KI. Development of sweet taste perception: implications for artificial sweetener use. *Endocr Dev* 2017;32:87–99.
46. Mennella JA, Bobowski NK, Reed DR. The development of sweet taste: from biology to hedonics. *Rev Endocr Metab Disord* 2016;17(2):171–8.
47. Desor JA, Beauchamp GK. Longitudinal changes in sweet preferences in humans. *Physiol Behav* 1987;39(5):639–41.
48. Murphy C, Schubert CR, Cruickshanks KJ, Klein BE, Klein R, Nondahl DM. Prevalence of olfactory impairment in older adults. *JAMA* 2002;288(18):2307–12.
49. Hoffman HJ, Bartoshuk LM, Losonczy KG, Rawal S, Hayes JE, Li C-M, Duffy VB. Dysgeusia, hypogeusia and hypergeusia among US adults aged ≥ 40 years: the National Health and Nutrition Examination Survey (NHANES), 2011–2014 (abstr). *Chem Senses* 2019;44(7):e8.
50. Garcia J, Hankins WG, Rusiniak KW. Behavioral regulation of the milieu interne in man and rat. *Science* 1974;185(4154):824–31.
51. Murphy C. Olfactory and other sensory impairments in Alzheimer disease. *Nat Rev Neurol* 2019;15(1):11–24.
52. Bertino M, Beauchamp GK, Engelman K. Long-term reduction in dietary sodium alters the taste of salt. *Am J Clin Nutr* 1982;36(6):1134–44.
53. Wise PM, Nattress L, Flammer LJ, Beauchamp GK. Reduced dietary intake of simple sugars alters perceived sweet taste intensity but not perceived pleasantness. *Am J Clin Nutr* 2016;103(1):50–60.
54. Henney JE, Taylor CL, Boon CS. Strategies to reduce sodium intake in the United States. Washington (DC): National Academies Press; 2010.
55. Mattes RD. Fat preference and adherence to a reduced-fat diet. *Am J Clin Nutr* 1993;57(3):373–81.
56. Costanzo A, Orellana L, Nowson C, Duesing K, Keast R. Fat taste sensitivity is associated with short-term and habitual fat intake. *Nutrients* 2017;9(7):781.
57. Rozin P, Schiller D. The nature and acquisition of a preference for chili pepper by humans. *Motiv Emot* 1980;4:77–101.
58. Abdel-Salam OME. Preference for hot pepper: a complex interplay of personal, cultural, and pharmacological effects. *Temperature* 2016;3(1):39–40.
59. Dessirier JM, Simons CT, Sudo M, Sudo S, Carstens E. Sensitization, desensitization and stimulus-induced recovery of trigeminal neuronal responses to oral capsaicin and nicotine. *J Neurophysiol* 2000;84(4):1851–62.
60. Byrnes NK, Hayes JE. Personality factors predict spicy food liking and intake. *Food Qual Preference* 2013;28(1):213–21.
61. Yeomans MR. Flavour–nutrient learning in humans: an elusive phenomenon? *Physiol Behav* 2012;106(3):345–55.
62. Noble EE, Kanoski SE. Early life exposure to obesogenic diets and learning and memory dysfunction. *Curr Opin Behav Sci* 2016;9:7–14.
63. Hsu TM, Konanur VR, Taing L, Usui R, Kayser BD, Goran MI, Kanoski SE. Effects of sucrose and high fructose corn syrup consumption on spatial memory function and hippocampal neuroinflammation in adolescent rats. *Hippocampus* 2015;25(2):227–39.
64. Kendig MD, Boakes RA, Rooney KB, Corbit LH. Chronic restricted access to 10% sucrose solution in adolescent and young adult rats impairs spatial memory and alters sensitivity to outcome devaluation. *Physiol Behav* 2013;120:164–72.
65. Reichelt AC, Killcross S, Hambly LD, Morris MJ, Westbrook RF. Impact of adolescent sucrose access on cognitive control, recognition memory, and parvalbumin immunoreactivity. *Learn Mem* 2015;22(4):215–24.
66. Noble EE, Hsu TM, Liang J, Kanoski SE. Early-life sugar consumption has long-term negative effects on memory function in male rats. *Nutr Neurosci* 2019;22(4):273–83.
67. Pepino MY, Bradley D, Eagon JC, Sullivan S, Abumrad NA, Klein S. Changes in taste perception and eating behavior after bariatric surgery-induced weight loss in women. *Obesity* 2014;22(5):E13–E20.

68. Pellegrino R, Walliczek-Dworschak U, Winter G, Hull D, Hummel T. Investigation of chemosensitivity during and after an acute cold. *Int Forum Allergy Rhinol* 2017;7(2):185–91.
69. McCluskey LP, He L, Dong G, Harris R. Chronic exposure to liquid sucrose and dry sucrose diet have differential effects on peripheral taste responses in female rats. *Appetite* 2020;145:104499.
70. McIntyre JC, Thiebaud N, McGann JP, Komiyama T, Rothermel M. Neuromodulation in chemosensory pathways. *Chem Senses* 2017;42(5):375–9.
71. Fadool DA, Tucker K, Pedarzani P. Mitral cells of the olfactory bulb perform metabolic sensing and are disrupted by obesity at the level of the Kv1.3 ion channel. *PLoS One* 2011;6(9):e24921.
72. Thiebaud N, Johnson MC, Butler JL, Bell GA, Ferguson KL, Fadool AR, Fadool JC, Gale AM, Gale DS, Fadool DA. Hyperlipidemic diet causes loss of olfactory sensory neurons, reduces olfactory discrimination, and disrupts odor-reversal learning. *J Neurosci* 2014;34(20):6970–84.
73. Palouzier-Paulignan B, Lacroix MC, Aime P, Baly C, Caillol M, Congar P, Julliard AK, Tucker K, Fadool DA. Olfaction under metabolic influences. *Chem Senses* 2012;37(9):769–97.
74. Savigner A, Duchamp-Viret P, Grosmaître X, Chaput M, Garcia S, Ma M, Palouzier-Paulignan B. Modulation of spontaneous and odorant-evoked activity of rat olfactory sensory neurons by two anorectic peptides, insulin and leptin. *J Neurophysiol* 2009;101(6):2898–906.
75. Thiebaud N, Gribble F, Reimann F, Trapp S, Fadool DA. A unique olfactory bulb microcircuit driven by neurons expressing the precursor to glucagon-like peptide 1. *Sci Rep* 2019;9(1):15542.
76. Tucker K, Cho S, Thiebaud N, Henderson MX, Fadool DA. Glucose sensitivity of mouse olfactory bulb neurons is conveyed by a voltage-gated potassium channel. *J Physiol* 2013;591(10):2541–61.
77. Peyron C, Tighe DK, van den Pol AN, de Lecea L, Heller HC, Sutcliffe JG, Kilduff TS. Neurons containing hypocretin (orexin) project to multiple neuronal systems. *J Neurosci* 1998;18(23):9996–10015.
78. Gascuel J, Lemoine A, Rigault C, Datiche F, Benani A, Penicaud L, Lopez-Mascaraque L. Hypothalamus-olfactory system crosstalk: orexin A immunostaining in mice. *Front Neuroanat* 2012;6:44.
79. Russo C, Russo A, Pellitteri R, Stanzani S. Ghrelin-containing neurons in the olfactory bulb send collateralized projections into medial amygdaloid and arcuate hypothalamic nuclei: neuroanatomical study. *Exp Brain Res* 2018;236(8):2223–9.
80. In 't Zandt EE, Cansler HL, Denson HB, Wesson DW. Centrifugal innervation of the olfactory bulb: a reappraisal. *eNeuro* 2019;6(1):ENEURO.0390–18.2019.
81. Padmanabhan K, Osakada F, Tarabrina A, Kizer E, Callaway EM, Gage FH, Sejnowski TJ. Centrifugal inputs to the main olfactory bulb revealed through whole brain circuit-mapping. *Front Neuroanat* 2019;12:115.
82. Boyd AM, Sturgill JF, Poo C, Isaacson JS. Cortical feedback control of olfactory bulb circuits. *Neuron* 2012;76(6):1161–74.
83. Brunet D, Tsuno Y, Rothermel M, Shipley MT, Wachowiak M. Cell-type-specific modulation of sensory responses in olfactory bulb circuits by serotonergic projections from the raphe nuclei. *J Neurosci* 2016;36(25):6820–35.
84. Davis BJ, Macrides F. The organization of centrifugal projections from the anterior olfactory nucleus, ventral hippocampal rudiment, and piriform cortex to the main olfactory bulb in the hamster: an autoradiographic study. *J Comp Neurol* 1981;203(3):475–93.
85. Illig KR. Projections from orbitofrontal cortex to anterior piriform cortex in the rat suggest a role in olfactory information processing. *J Comp Neurol* 2005;488(2):224–31.
86. Mazo C, Lepousez G, Nissant A, Valley MT, Lledo PM. GABAB receptors tune cortical feedback to the olfactory bulb. *J Neurosci* 2016;36(32):8289–304.
87. McLean JH, Shipley MT, Nickell WT, Aston-Jones G, Reyher CK. Chemoanatomical organization of the noradrenergic input from locus coeruleus to the olfactory bulb of the adult rat. *J Comp Neurol* 1989;285(3):339–49.
88. Mouly AM, Di Scala G. Entorhinal cortex stimulation modulates amygdala and piriform cortex responses to olfactory bulb inputs in the rat. *Neuroscience* 2006;137(4):1131–41.
89. Nagayama S, Enerva A, Fletcher ML, Masurkar AV, Igarashi KM, Mori K, Chen WR. Differential axonal projection of mitral and tufted cells in the mouse main olfactory system. *Front Neural Circuits* 2010;4:120.
90. Rothermel M, Carey RM, Puche A, Shipley MT, Wachowiak M. Cholinergic inputs from basal forebrain add an excitatory bias to odor coding in the olfactory bulb. *J Neurosci* 2014;34(13):4654–64.
91. Steinfeld R, Herb JT, Sprengel R, Schaefer AT, Fukunaga I. Divergent innervation of the olfactory bulb by distinct raphe nuclei. *J Comp Neurol* 2015;523(5):805–13.
92. Fast CD, McGann JP. Amygdalar gating of early sensory processing through interactions with locus coeruleus. *J Neurosci* 2017;37(11):3085–101.
93. Sternson SM, Betley JN, Cao ZFH. Neural circuits and motivational processes for hunger. *Curr Opin Neurobiol* 2013;23(3):353–60.
94. Sun X, Veldhuizen MG, Wray AE, de Araujo IE, Sherwin RS, Sinha R, Small DM. The neural signature of satiation is associated with ghrelin response and triglyceride metabolism. *Physiol Behav* 2014;136:63–73.
95. Vincis R, Fontanini A. Central taste anatomy and physiology. *Handb Clin Neurol* 2019;164:187–204.
96. Chambers L, McCrickerd K, Yeomans MR. Optimising foods for satiety. *Trends Food Sci Technol* 2015;41(2):149–60.
97. Blundell J, Rogers P, Hill A. Evaluating the satiating power of foods: implications for acceptance and consumption. In: Colms J, Booth D, Pangborn R, Raunhardt O, editors. *Food acceptance and nutrition*. London: Academic Press; 1987. p. 205–19.
98. Latorre R, Huynh J, Mazzoni M, Gupta A, Bonora E, Clavenzani P, Chang L, Mayer EA, De Giorgio R, Sternini C. Expression of the bitter taste receptor, T2R38, in enteroendocrine cells of the colonic mucosa of overweight/obese vs. lean subjects. *PLoS One* 2016;11(2):e0147468.
99. Vegezzi G, Anselmi L, Huynh J, Barocelli E, Rozengurt E, Raybould H, Sternini C. Diet-induced regulation of bitter taste receptor subtypes in the mouse gastrointestinal tract. *PLoS One* 2014;9(9):e107732.
100. Lyte M. Microbial endocrinology: host-microbiota neuroendocrine interactions influencing brain and behavior. *Gut Microbes* 2014;5(3):381–9.
101. Ghosh DD, Sanders T, Hong S, McCurdy LY, Chase DL, Cohen N, Koelle MR, Nitabach MN. Neural architecture of hunger-dependent multisensory decision making in *C. elegans*. *Neuron* 2016;92(5):1049–62.
102. Coldwell SE, Mennella JA, Duffy VB, Pelchat ML, Griffith JW, Smutzer G, Cowart BJ, Breslin PA, Bartoshuk LM, Hastings L, et al. Gustation assessment using the NIH Toolbox. *Neurology* 2013;80(11 Suppl 3):S20–4.
103. Dalton P, Doty RL, Murphy C, Frank R, Hoffman HJ, Maute C, Kallen MA, Slotkin J. Olfactory assessment using the NIH Toolbox. *Neurology* 2013;80(11 Suppl 3):S32–6.
104. Mennella JA, Lukasewycz LD, Griffith JW, Beauchamp GK. Evaluation of the Monell forced-choice, paired-comparison tracking procedure for determining sweet taste preferences across the lifespan. *Chem Senses* 2011;36(4):345–55.
105. Hoffman HJ, Rawal S, Li C-M, Duffy VB. New chemosensory component in the U.S. National Health and Nutrition Examination Survey (NHANES): first-year results for measured olfactory dysfunction. *Rev Endocr Metab Disord* 2016;17(2):221–40.
106. Yang J, Pinto JM. The epidemiology of olfactory disorders. *Curr Otorhinolaryngol Rep* 2016;4(2):130–41.
107. Ekström I, Sjölund S, Nordin S, Nordin Adolfsson A, Adolfsson R, Nilsson L-G, Larsson M, Olofsson JK. Smell loss predicts mortality risk regardless of dementia conversion. *J Am Geriatr Soc* 2017;65(6):1238–43.
108. Ross GW, Petrovitch H, Abbott RD, Tanner CM, Popper J, Masaki K, Launer L, White LR. Association of olfactory dysfunction with risk for future Parkinson's disease. *Ann Neurol* 2008;63(2):167–73.
109. Dintica CS, Marseglia A, Rizzuto D, Wang R, Seubert J, Arfanakis K, Bennett DA, Xu W. Impaired olfaction is associated with cognitive decline and neurodegeneration in the brain. *Neurology* 2019;92(7):e700–e9.
110. Murphy C, Schubert CR, Cruickshanks KJ, Klein BE, Klein R, Nondahl DM. Prevalence of olfactory impairment in older adults. *JAMA* 2002;288(18):2307–12.

111. Bramerson A, Johansson L, Ek L, Nordin S, Bende M. Prevalence of olfactory dysfunction: the Skövde population-based study. *Laryngoscope* 2004;114(4):733–7.
112. Karpa MJ, Gopinath B, Rochtchina E, Wang JJ, Cumming RG, Sue CM, Mitchell P. Prevalence and neurodegenerative or other associations with olfactory impairment in an older community. *J Aging Health* 2010;22(2):154–68.
113. Devanand DP, Lee S, Manly J, Andrews H, Schupf N, Doty RL, Stern Y, Zahodne LB, Louis ED, Mayeux R. Olfactory deficits predict cognitive decline and Alzheimer dementia in an urban community. *Neurology* 2015;84(2):182–9.
114. Leschak CJ, Eisenberger NI. The role of social relationships in the link between olfactory dysfunction and mortality. *PLoS One* 2018;13(5):e0196708.
115. Liu Y-H, Huang Z, Vaidya A, Li J, Curhan GC, Wu S, Gao X. A longitudinal study of altered taste and smell perception and change in blood pressure. *Nutr Metab Cardiovasc Dis* 2018;28(9):877–83.
116. Joo Y-H, Hwang S-H, Han K-D, Seo J-H, Kang J-M. Relationship between olfactory dysfunction and suicidal ideation: the Korea National Health and Nutrition Examination Survey. *Am J Rhinol Allergy* 2015;29(4):268–72.
117. Bainbridge KE, Byrd-Clark D, Leopold D. Factors associated with phantom odor perception among US adults: findings from the National Health and Nutrition Examination Survey. *JAMA Otolaryngol Head Neck Surg* 2018;144(9):807–14.
118. Fischer ME, Cruickshanks KJ, Schubert CR, Pinto A, Huang G-H, Klein BE, Klein R, Pankow JS. The association of taste with change in adiposity-related health measures. *J Acad Nutr Diet* 2014;114(8):1195–202.
119. Fischer ME, Cruickshanks KJ, Schubert CR, Pinto A, Klein R, Pankratz N, Pankow JS, Huang G-H. Factors related to fungiform papillae density: the Beaver Dam Offspring Study. *Chem Senses* 2013;38(8):669–77.
120. Schumm LP, McClintock M, Williams S, Leitsch S, Lundstrom J, Hummel T, Lindau ST. Assessment of sensory function in the National Social Life, Health, and Aging Project. *J Gerontol B Psychol Sci Soc Sci* 2009;64(Suppl 1):i76–85.
121. Correia C, Lopez KJ, Wroblewski KE, Huisingh-Scheetz M, Kern DW, Chen RC, Schumm LP, Dale W, McClintock MK, Pinto JM. Global sensory impairment in older adults in the United States. *J Am Geriatr Soc* 2016;64(2):306–13.
122. Ekback G, Ordell S. Self-perceived taste disturbance: a 20-year prospective study of a Swedish 1942 birth cohort. *Gerodontology* 2017;34(2):180–6.
123. Uota M, Ogawa T, Ikebe K, Arai Y, Kamide K, Gondo Y, Masui Y, Ishizaki T, Inomata C, Takeshita H, et al. Factors related to taste sensitivity in elderly: cross-sectional findings from SONIC study. *J Oral Rehabil* 2016;43(12):943–52.
124. Risso DS, Kozlitina J, Sainz E, Gutierrez J, Wooding S, Getachew B, Luiselli D, Berg CJ, Drayna D. Genetic variation in the *TAS2R38* bitter taste receptor and smoking behaviors. *PLoS One* 2016;11(10):e0164157.
125. Tepper BJ, Koelliker Y, Zhao L, Ullrich NV, Lanzara C, d'Adamo P, Ferrara A, Ulivi S, Esposito L, Gasparini P. Variation in the bitter-taste receptor gene *TAS2R38*, and adiposity in a genetically isolated population in Southern Italy. *Obesity (Silver Spring)* 2008;16(10):2289–95.
126. Lambert JD, VanDusen SR, Cockroft JE, Smith EC, Greenwood DC, Cade JE. Bitter taste sensitivity, food intake, and risk of malignant cancer in the UK Women's Cohort Study. *Eur J Nutr* 2019;58(5):2111–21.
127. Robino A, Mezzavilla M, Pirastu N, Dognini M, Tepper BJ, Gasparini P. A population-based approach to study the impact of PROP perception on food liking in populations along the Silk Road. *PLoS One* 2014;9(3):e91716.
128. Hwang L-D, Breslin PA, Reed DR, Zhu G, Martin NG, Wright MJ. Is the association between sweet and bitter perception due to genetic variation? *Chem Senses* 2016;41(9):737–44.
129. Ahrens W, Bammann K, Siani A, Buchecker K, De Henauw S, Iacoviello L, Hebestreit A, Krogh V, Lissner L, Marild S, et al. The IDEFICS cohort: design, characteristics and participation in the baseline survey. *Int J Obes* 2011;35(Suppl 1):S3–15.
130. Wijtzes AI, Jansen W, Bouthoorn SH, Kieft-de Jong JC, Jansen PW, Franco OH, Jaddoe VW, Hofman A, Raat H. PROP taster status, food preferences and consumption of high-calorie snacks and sweet beverages among 6-year-old ethnically diverse children. *Matern Child Nutr* 2017;13(2):e12240.
131. Fox AL. The relationship between chemical composition and taste. *Science* 1931;74:607.
132. Bartoshuk LM, Duffy VB, Miller IJ. PTC/PROP tasting: anatomy, psychophysics, and sex effects. *Physiol Behav* 1994;56:1165–71.
133. Duffy VB, Hayes JE, Sullivan BS, Faghri P. Surveying food and beverage liking: a tool for epidemiological studies to connect chemosensation with health outcomes. *Ann N Y Acad Sci* 2009;1170:558–68.
134. Rawal S, Hoffman HJ, Chapo AK, Duffy VB. Sensitivity and specificity of self-reported olfactory function in a home-based study of independent-living, healthy older women. *Chem Percept* 2014;7(3–4):108–16.
135. Duffy VB, Lucchina LA, Bartoshuk LM. Genetic variation in taste: potential biomarker for cardiovascular disease risk? In: Prescott J, Tepper B, editors. *Sensitivity to PROP (6-n-propylthiouracil): measurement, significances and implications*. Erlangen, Germany: Merril Decker, Inc; 2004. p. 197–229.
136. Lundström JN, Boesveldt S, Albrecht J. Central processing of the chemical senses: an overview. *ACS Chem Neurosci* 2011;2:5–16.
137. Fazzino TL, Rohde K, Sullivan DK. Hyper-palatable foods: development of a quantitative definition and application to the US Food System Database. *Obesity* 2019;27(11):1761–8.
138. DiFeliceantonio AG, Coppin G, Rigoux L, Edwin Thanarajah S, Dagher A, Tittgemeyer M, Small DM. Supra-additive effects of combining fat and carbohydrate on food reward. *Cell Metab* 2018;28(1):33–44.e3.
139. Steiner JE, Glaser D, Hawilo ME, Berridge KC. Comparative expression of hedonic impact: affective reactions to taste by human infants and other primates. *Neurosci Biobehav Rev* 2001;25(1):53–74.
140. Bozza T, McGann JP, Mombaerts P, Wachowiak M. In vivo imaging of neuronal activity by targeted expression of a genetically encoded probe in the mouse. *Neuron* 2004;42(1):9–21.
141. Murthy VN. Olfactory maps in the brain. *Annu Rev Neurosci* 2011;34:233–58.
142. Wachowiak M, Cohen LB. Representation of odorants by receptor neuron input to the mouse olfactory bulb. *Neuron* 2001;32(4):723–35.
143. Boutelle KN, Wierenga CE, Bischoff-Grethe A, Melrose AJ, Grenesko-Stevens E, Paulus MP, Kaye WH. Increased brain response to appetitive tastes in the insula and amygdala in obese compared with healthy weight children when sated. *Int J Obes* 2015;39(4):620–8.
144. Green E, Jacobson A, Haase L, Murphy C. Neural correlates of taste and pleasantness evaluation in the metabolic syndrome. *Brain Res* 2015;1620:57–71.
145. Bartoshuk LM. Sweetness: history, preference, and genetic variability. *Food Technol* 1991;45(11):108–13.
146. Azen EA, Maeda N. Molecular genetics of human salivary proteins and their polymorphisms. *Adv Hum Genet* 1988;17:141–99.
147. Crystal SR, Teff KL. Tasting fat: cephalic phase hormonal responses and food intake in restrained and unrestrained eaters. *Physiol Behav* 2006;89(2):213–20.
148. Kaplan AR, Fischer R, Karras A, Griffin F, Powell W, Marsters RW, Glanville EV. Taste thresholds in twins and siblings. *Acta Genet Med Gemellol (Roma)* 1967;16(3):229–43.
149. Fuller JL. Single-locus control of saccharin preference in mice. *J Hered* 1974;65(1):33–6.
150. Lyte M, Fodor AA, Chapman CD, Martin GG, Perez-Chanona E, Jobin C, Dess NK. Gut microbiota and a selectively bred taste phenotype: a novel model of microbiome-behavior relationships. *Psychosom Med* 2016;78(5):610–19.
151. Cornelis MC, Tordoff MG, El-Sohemy A, van Dam RM. Recalled taste intensity, liking and habitual intake of commonly consumed foods. *Appetite* 2017;109:182–9.
152. Cornelis MC, Monda KL, Yu K, Paynter N, Azzato EM, Bennett SN, Berndt SI, Boerwinkle E, Chanock S, Chatterjee N, et al. Genome-wide meta-analysis identifies regions on 7p21 (*AHR*) and 15q24 (*CYP1A2*) as determinants of habitual caffeine consumption. *PLoS Genet* 2011;7(4):e1002033.
153. Cole JB, Florez JC, Hirschhorn JN. Comprehensive genomic analysis of dietary habits in UK Biobank identifies hundreds of genetic loci

- and establishes causal relationships between educational attainment and healthy eating. *Nat Commun* 2020;11:Article number 1467.
154. National Institute on Deafness and Other Communication Disorders. Taste disorders [Internet]. Bethesda, MD: NIDCD Information Clearinghouse; 2017 [accessed 6 November, 2020]. Available from: <https://www.nidcd.nih.gov/health/taste-disorders>.
 155. Healthy People 2030. Increase the proportion of adults with smell or taste disorders who discuss the problem with a provider — HOSCD–12 [Internet]. Rockville, MD: Office of Disease Prevention and Health Promotion [accessed 6 November, 2020]. Available from: <https://health.gov/healthypeople/objectives-and-data/browse-objectives/sensory-or-communication-disorders/increase-proportion-adults-smell-or-taste-disorders-who-discuss-problem-provider-hoscd-12>.
 156. Mainland JD, Barlow LA, Munger SD, Millar SE, Vergara MN, Jiang P, Schwob JE, Goldstein BJ, Boye SE, Martens JR, et al. Identifying treatments for taste and smell disorders: gaps and opportunities. *Chem Senses* 2020;45(7):493–502.
 157. Sudlow C, Gallacher J, Allen N, Beral V, Burton P, Danesh J, Downey P, Elliott P, Green J, Landray M, et al. UK Biobank: an open access resource for identifying the causes of a wide range of complex diseases of middle and old age. *PLoS Med* 2015;12(3):e1001779.
 158. Gaziano JM, Concato J, Brophy M, Fiore L, Pyarajan S, Breeling J, Whitbourne S, Deen J, Shannon C, Humphries D, et al. Million Veteran Program: a mega-biobank to study genetic influences on health and disease. *J Clin Epidemiol* 2016;70:214–23.
 159. Precision Medicine Initiative (PMI) Working Group. The Precision Medicine Initiative Cohort Program – building a research foundation for 21st century medicine. Report to the Advisory Committee to the Director. Bethesda, MD: NIH; 2015.
 160. Kyriazis GA, Smith KR, Tyrberg B, Hussain T, Pratley RE. Sweet taste receptors regulate basal insulin secretion and contribute to compensatory insulin hypersecretion during the development of diabetes in male mice. *Endocrinology* 2014;155(6):2112–21.
 161. Young RL, Sutherland K, Pezos N, Brierley SM, Horowitz M, Rayner CK, Blackshaw LA. Expression of taste molecules in the upper gastrointestinal tract in humans with and without type 2 diabetes. *Gut* 2009;58(3):337–46.
 162. Young RL, Chia B, Isaacs NJ, Ma J, Khoo J, Wu T, Horowitz M, Rayner CK. Disordered control of intestinal sweet taste receptor expression and glucose absorption in type 2 diabetes. *Diabetes* 2013;62(10):3532–41.
 163. Smith K, Karimian Azari E, LaMoia TE, Hussain T, Vargova V, Karolyi K, Veldhuis PP, Arnoletti JP, de la Fuente SG, Pratley RE, et al. T1R2 receptor-mediated glucose sensing in the upper intestine potentiates glucose absorption through activation of local regulatory pathways. *Mol Metab* 2018;17:98–111.
 164. Daly K, Darby AC, Hall N, Nau A, Bravo D, Shirazi-Beechey SP. Dietary supplementation with lactose or artificial sweetener enhances swine gut *Lactobacillus* population abundance. *Br J Nutr* 2014;111(Suppl 1):S30–5.
 165. Frankenfeld CL, Sikaroodi M, Lamb E, Shoemaker S, Gillevet PM. High-intensity sweetener consumption and gut microbiome content and predicted gene function in a cross-sectional study of adults in the United States. *Ann Epidemiol* 2015;25(10):736–42.e4.
 166. Bienenstock J, Kunze WA, Forsythe P. Disruptive physiology: olfaction and the microbiome–gut–brain axis. *Biol Rev Camb Philos Soc* 2018;93(1):390–403.
 167. Ezenwa VO, Williams AE. Microbes and animal olfactory communication: where do we go from here? *Bioessays* 2014;36(9):847–54.
 168. Carthey AJR, Gillings MR, Blumstein DT. The extended genotype: microbially mediated olfactory communication. *Trends Ecol Evol* 2018;33(11):885–94.
 169. van de Wouw M, Schellekens H, Dinan TG, Cryan JF. Microbiota-gut-brain axis: modulator of host metabolism and appetite. *J Nutr* 2017;147(5):727–45.
 170. Lee RJ, Xiong G, Kofonow JM, Chen B, Lysenko A, Jiang P, Abraham V, Doghranji L, Adappa ND, Palmer JN, et al. T2R38 taste receptor polymorphisms underlie susceptibility to upper respiratory infection. *J Clin Invest* 2012;122(11):4145–59.
 171. Lyte M. Microbial endocrinology and the microbiota-gut-brain axis. *Adv Exp Med Biol* 2014;817:3–24.
 172. Lyte M. Microbial endocrinology and nutrition: a perspective on new mechanisms by which diet can influence gut-to-brain communication. *PharmaNutrition* 2013;1(1):35–9.
 173. Wilbert SA, Mark Welch JL, Borisy GG. Spatial ecology of the human tongue dorsum microbiome. *Cell Rep* 2020;30(12):4003–15.e3.
 174. Koskinen K, Reichert JL, Hoier S, Schachenreiter J, Duller S, Moissl-Eichinger C, Schopf V. The nasal microbiome mirrors and potentially shapes olfactory function. *Sci Rep* 2018;8(1):1296.
 175. Almeida A, Mitchell AL, Boland M, Forster SC, Gloor GB, Tarkowska A, Lawley TD, Finn RD. A new genomic blueprint of the human gut microbiota. *Nature* 2019;568(7753):499–504.
 176. Yee KK, Sukumaran SK, Kotha R, Gilbertson TA, Margoskee RF. Glucose transporters and ATP-gated K⁺ (K_{ATP}) metabolic sensors are present in type 1 taste receptor 3 (T1r3)-expressing taste cells. *Proc Natl Acad Sci U S A* 2011;108(13):5431–6.
 177. Kok BP, Galmozzi A, Littlejohn NK, Albert V, Godio C, Kim W, Kim SM, Bland JS, Grayson N, Fang M, et al. Intestinal bitter taste receptor activation alters hormone secretion and imparts metabolic benefits. *Mol Metab* 2018;16:76–87.
 178. Gu F, Liu X, Liang J, Chen F, Li F. Bitter taste receptor mTas2r105 is expressed in small intestinal villus and crypts. *Biochem Biophys Res Commun* 2015;463(4):934–41.
 179. Prandi S, Bromke M, Hubner S, Voigt A, Boehm U, Meyerhof W, Behrens M. A subset of mouse colonic goblet cells expresses the bitter taste receptor Tas2r131. *PLoS One* 2013;8(12):e82820.
 180. Prandi S, Voigt A, Meyerhof W, Behrens M. Expression profiling of Tas2r genes reveals a complex pattern along the mouse GI tract and the presence of Tas2r131 in a subset of intestinal Paneth cells. *Cell Mol Life Sci* 2018;75(1):49–65.
 181. Depoortere I. Taste receptors of the gut: emerging roles in health and disease. *Gut* 2014;63(1):179–90.
 182. Sternini C. Taste receptors in the gastrointestinal tract. IV. Functional implications of bitter taste receptors in gastrointestinal chemosensing. *Am J Physiol Gastrointest Liver Physiol* 2007;292(2):G457–61.
 183. Meyerhof W, Batram C, Kuhn C, Brockhoff A, Chudoba E, Bufe B, Appendino G, Behrens M. The molecular receptive ranges of human TAS2R bitter taste receptors. *Chem Senses* 2010;35(2):157–70.
 184. Saito H, Chi Q, Zhuang H, Matsunami H, Mainland JD. Odor coding by a mammalian receptor repertoire. *Sci Signal* 2009;2(60):ra9.
 185. Lossow K, Hubner S, Roudnitsky N, Slack JP, Pollastro F, Behrens M, Meyerhof W. Comprehensive analysis of mouse bitter taste receptors reveals different molecular receptive ranges for orthologous receptors in mice and humans. *J Biol Chem* 2016;291(29):15358–77.
 186. GTEx Consortium. The Genotype-Tissue Expression (GTEx) project. *Nat Genet* 2013;45(6):580–5.
 187. Baker MS, Ahn SB, Mohamedali A, Islam MT, Cantor D, Verhaert PD, Fanayan S, Nice EC, Connor M, et al. Accelerating the search for the missing proteins in the human proteome. *Nat Commun* 2017;8:14271.
 188. Chiu IM, Barrett LB, Williams EK, Strohlic DE, Lee S, Weyer AD, Lou S, Bryman GS, Roberson DP, Ghasemlou N, et al. Transcriptional profiling at whole population and single cell levels reveals somatosensory neuron molecular diversity. *Elife* 2014;3:e04660.
 189. Rozenblatt-Rosen O, Stubbington MJT, Regev A, Teichmann SA. The Human Cell Atlas: from vision to reality. *Nature* 2017;550(7677):451–3.
 190. Brann DH, Tsukahara T, Weinreb C, Logan DW, Datta SR. Non-neural expression of SARS-CoV-2 entry genes in the olfactory epithelium suggests mechanisms underlying anosmia in COVID-19 patients. *bioRxiv* 2020:2020.03.25.009084.
 191. Behrens M, Born S, Redel U, Voigt N, Schuh V, Raguse JD, Meyerhof W. Immunohistochemical detection of TAS2R38 protein in human taste cells. *PLoS One* 2012;7(7):e40304.
 192. Dweck HKM, Carlson JR. Molecular logic and evolution of bitter taste in *Drosophila*. *Curr Biol* 2020;30(1):17.
 193. Gerkin RC, Ohla K, Veldhuizen MG, Joseph PV, Kelly CE, Bakke AJ, Steele KE, Farruggia MC, Pellegrino R, Pepino MY, et al. The best COVID-19 predictor is recent smell loss: a cross-sectional study. *medRxiv* 2020:2020.07.22.20157263.

194. Menni C, Valdes A, Freydin MB, Ganesh S, El-Sayed Moustafa J, Visconti A, Hysi P, Bowyer RCE, Mangino M, Falchi M, et al. Loss of smell and taste in combination with other symptoms is a strong predictor of COVID-19 infection. *Nat Med* 2020;26:1037–40.
195. Hannum ME, Ramirez VA, Lipson SJ, Herriman RD, Toskala AK, Lin C, Joseph PV, Reed DR. Objective sensory testing methods reveal a higher prevalence of olfactory loss in COVID-19 positive patients compared to subjective methods: a systematic review and meta-analysis. *Chem Senses* 2020:bjaa064.