



DPP-IV as a potential candidate in anti-obesity and obesity-related diseases treatment

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ABSTRACT

Along with social development and lifestyle changes, the number of overweight and obese patients worldwide is rising annually. Obesity is a chronic metabolic disease with complex etiology. Dipeptidyl peptidase IV (DPP-IV) is a novel adipokine with significantly elevated expression in the visceral fat of obese patients. DPP-IV is a molecule that regulates metabolic homeostasis and inflammatory processes. Through its enzymatic activity, it plays a significant part in achieving hypoglycemic and weight loss effects through various pathways. DPP-IV and DPP-IV inhibitors also have pleiotropic effects in modulating obesity-related diseases by reducing obesity-related inflammation, ameliorating inflammatory bowel disease (IBD), improving hepatic steatosis and lowering cardiovascular risk, and even decreasing the risk of novel coronavirus disease-19 (COVID-19). This paper reviews the mechanisms of action based on DPP-IV targets in obesity and metabolic homeostasis, as well as their active role in the treatment of chronic diseases associated with obesity.

1. Introduction

The latest research report published by The Lancet shows that the global population with obesity reached over 1 billion in 2022. Since 1990, the number of obese adults worldwide has more than doubled, while the number of obese children and adolescents (ages 5–19) has tripled. The data also indicates that in 2022, 43 % of adults were overweight[1]. Obesity is caused by a variety of factors, including the environment (environmental pollution and urbanization), genetics (metabolic rate and fat distribution), neuropsychiatric disorders (psychological stress and exercise psychology), endocrine issues (insulin imbalance and hypothyroidism), abnormal brown adipose tissue (insufficient brown adipose tissue), and others. It is a complex metabolic disorder defined as a body mass index (BMI) ≥ 30 kg/m²[2] (Fig. 1).

Obesity is associated with many chronic diseases such as type II diabetes (T2DM), hypertension, dyslipidemia, cardiovascular disease, inflammatory bowel disease (IBD)[3], non-alcoholic fatty liver disease

(NAFLD), obstructive sleep apnea syndrome, higher chance of hospitalization[4,5], and poorer prognosis in patients with coronavirus-19 (COVID-19)[6]. Recent studies[7,8] have shown that obesity is associated with increased cancer risk, recurrence, and poor prognosis. With the global prevalence of overweight and obesity gradually rising, obesity has become one of the greatest global health challenges and a major basis for the development of metabolic diseases.

The novel adipokine known as dipeptidyl peptidase IV (DPP-IV, EC 3.4.14.5), which is secreted by differentiated human adipocytes and strongly expressed in visceral fat of obese patients, is correlated with adipocyte size and markers of metabolic syndrome[9,10]. Glucagon-like peptide-1 (GLP-1), glucagon-like peptide-2 (GLP-2), gastric inhibitory polypeptide (GIP), insulin-like growth factor-1 (IGF-1), neuropeptide Y (NPY), and peptide Y (PYY) are among the DPP-IV substrates that have been investigated in vivo. GLP-1 enhances insulin secretion and inhibits glucagon secretion in a glucose-concentration-dependent manner and is capable of delaying gastric emptying[11]. PYY is expressed in the central

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nervous system, decreases appetite, and is also released by enterocytes after food intake along with GLP-1, delaying gastric emptying and reducing intestinal motility and pancreatic exocrine secretion[12]. These mechanisms are related to their roles in diabetes, obesity, and other diseases.

2. Literature search

A narrative literature search was conducted on February 20, 2023. Search terms related to obesity (“obese” OR “obesity” OR “overweight” OR “weight-loss”); DPP-IV (“DPP4” OR “DPP-IV” OR “Dipeptidyl peptidase IV”); obesity treatment (“anti-obesity drugs” OR “bariatric surgery”); obesity-related diseases (“inflammation” OR “hepatic steatosis” OR “cardiovascular disease” OR “COVID-19”); and topics of interest (“etiology” OR “mechanism” OR “comorbidity” OR “structure”).

The PubMed database was searched using a combination of these keywords, with results restricted to the years 2013 through 2023 and the language English. The articles were reviewed for relevance based on the title and abstract.

Additional references were identified from the bibliographies of retrieved articles, through additional targeted searches, and through author suggestions.

3. Research status of obesity treatment

Current treatment measures for obesity include lifestyle and behavioral interventions, surgery, and medication. Although lifestyle interventions are recommended, they are difficult to implement in primary care practice and have limited long-term efficacy, mainly due to

poor adherence[13,14]. Bariatric surgery (BSG) is frequently preferable to non-surgical therapies for groups that are extremely obese or overweight, but many patients end up with lower-than-expected results because of surgical complications, post-operative malnutrition, and weight gain[15–17]. The research and development of anti-obesity drugs (AOM) have been facing great challenges due to technical and social reasons. There have been a series of AOM that failed after regulatory approval, most of which involved cardiovascular events (sibutramine, fenfluramine, dexfenfluramine, rainbow pills), increased incidence of cancer (lorcaserin), improved risk of suicide (rimonabant), or raised likelihood of drug dependence and abuse (methamphetamine) [18,19]. The U.S. Food and Drug Administration (FDA) approved anti-obesity drugs are orlistat, phentermine, metreleptin, liraglutide, and semaglutide[20,21]. However, the effects of drugs are limited, and many approved drugs have significant side effects. Therefore, the treatment of existing AOM, BSG, or lifestyle management, cannot satisfy the requirements of global medical needs.

4. The structure and function of DPP-IV

DPP-IV, first discovered in 1966[22], is a widely expressed glycoprotein with exopeptidase activity. Its structure was found to be identical to one of the differentiation clusters 26 (CD26), which was then referred to as the activation marker of T lymphocytes, namely the T cell surface antigen CD26[23]. The gene encoding for DPP-IV is located on the long arm of chromosome 2 (2q 24.3), preceded by a promoter region regulated by transcription factors, including the nuclear factor kappa-light-chain enhancer of activated B cells (NF-κB)[24], the hepatocyte nuclear factor 1β (HFN1β)[25], the lysine acetyltransferase 5

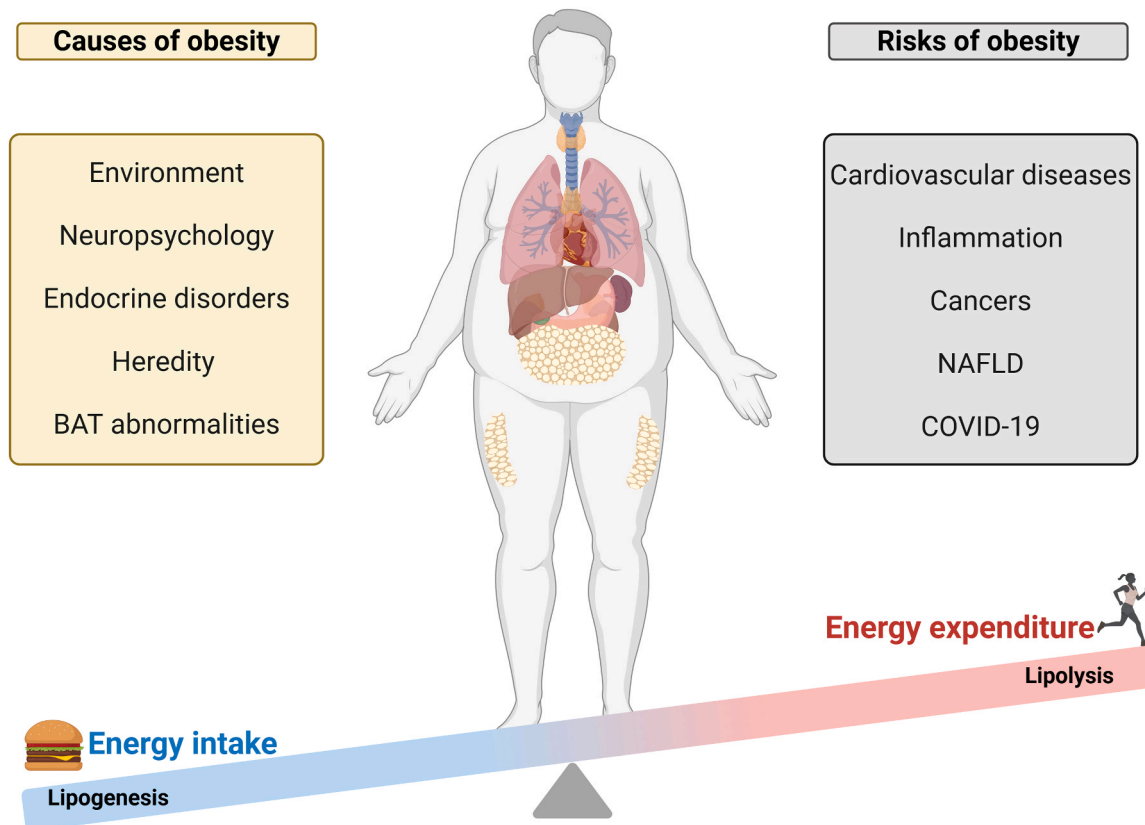


Fig. 1. Factors and Risks Contributing to Obesity. The significant rise in global obesity rates is primarily attributable to increased accessibility to high-energy foods coupled with a reduction in physical activity levels. The etiology of obesity is multifactorial, encompassing environmental influences, neuropsychological factors, endocrine disorders, genetic predispositions, and abnormalities in BAT. The associated risks encompass a wide range of health complications, including cardiovascular disease, chronic inflammation, various malignancies, NAFLD, and heightened susceptibility to COVID-19, BAT-brown adipose tissue. NAFLD- non-alcoholic fatty liver disease. COVID-19- coronavirus-19.

(KAT5)[26], and the signal transducers and activators of transcription 1 α (STAT1 α)[27], all of which are centrally involved in cell growth and proliferation, immune response and metabolism.

The amino acid sequence of DPP-IV contains the Gly-Trp-Ser-Tyr-Gly motif, which corresponds to the common Gly-X-Ser-X-Gly sequence found in the active sites of many serine proteases. Three critical residues are essential for the enzymatic activity of DPP-IV, namely Gly-629, Ser-631, and Gly-633. Each of the two subunits of DPP-IV consists of two domains: an α/β hydrolase domain and a β -propeller domain. The catalytic site is located within a large cavity formed between the α/β hydrolase domain and the β -propeller domain. The active site can be accessed through an opening at the center of the β -propeller domain facing the membrane (37=15/22 Å) or through an opening between the hydrolase and propeller domains (20=15/22 Å)[28,29]. DPP-IV exists in two forms, the soluble form (sDPP-IV) and the membrane-bound form. sDPP-IV has enzymatic activity, its main role is to preferentially shear off the first two amino acids from the amino terminus of oligopeptides in which the second amino acid at the amino terminus is alanine or proline[30]. DPP-IV is also a cell surface serine aminopeptidase that is widely expressed on the surfaces of epithelial cells, endothelial cells, stromal cells, stem cells, and immune cells in mice and humans[31]. DPP-IV catabolizes proteins *in vivo* and has many natural substrates, including growth factors, chemokines, neuropeptides, and vasoactive peptides[32]. Due to the wide range of DPP-IV substrates, impaired sDPP-IV levels and DPP-IV activity have been reported in several metabolic abnormalities such as T2DM, NAFLD, obesity, metabolic syndrome, and polycystic ovary syndrome[33–35]. In addition, elevated levels of DPP-IV have been identified in cancer, autoimmune diseases, and neurodegenerative diseases[36,37]. DPP-IV is involved in a variety of physiological and pathological processes in the body and is associated with the development of many diseases.

5. The role of DPP-IV in metabolism and lipid homeostasis

Several mechanisms or signals[38,39] related to appetite regulation and peripheral energy absorption, storage, and expenditure are gradually being revealed with technological, pharmacological, and surgical developments. Although the mechanisms involved are not yet fully understood, these studies have helped researchers to better understand the development of obesity.

6. Anti-obesity treatment with DPP-IV and GLP-1

6.1. Peripheral signaling pathway

GLP-1 can be secreted by gut enteroendocrine cells and brain preproglucagon (PPG) neurons, defined as the peripheral and central GLP-1 systems, respectively[40]. DPP-IV inhibitors increase GLP-1 levels *in vivo* by competitively binding to the DPP-IV activation site and decreasing the catalytic activity of the enzyme. Using the protein kinase (PKA)-dependent or guanine nucleotide exchange factor 4 (Epac2) pathway, intestinal-derived GLP-1 activates the glucagon-like peptide-1 receptor (GLP-1R) to increase glucose-stimulated insulin secretion in the periphery[41]. Insulin inhibits lipolysis and fat oxidation while promoting adipogenesis, which results in the accumulation of adipose tissue and ectopic fat storage. Excessive obesity can also result in insulin resistance. Through enhancing PKA activity, lowering endoplasmic reticulum stress, and boosting β -cell function, GLP-1 reduces insulin resistance in obese diabetic mice and promotes weight reduction[42].

PKA improves insulin sensitivity in peripheral tissues by inhibiting adenylate-activated protein kinase (AMPK)-related pathways. AMPK is an important metabolic stress protein kinase that, when activated, enhances skeletal muscle insulin sensitivity by increasing the translocation of glucose transporter protein (GLUT)[43]. Hydroxymethyl glutaryl-CoA (HMG-CoA) and Acetyl-CoA carboxylase (ACC) are key enzymes for cholesterol and fatty acid synthesis, both of which are

AMPK substrates[44]. By deactivating their phosphorylation, AMPK can inhibit the accumulation of fat and cholesterol, respectively. AMPK not only acts as an energy receptor at the cellular level, but also participates in regulating energy expenditure and intake, promoting glucolipid metabolism, accelerating fatty acid oxidation, and reducing insulin resistance through hormones and cytokines[45], such as lipocalin and leptin.

PKA also improves insulin sensitivity in peripheral tissues by elevating glyoxalase (GLO)[46]. Healthy adipose tissue is characterized by high vascular density, low hypoxia, fibrosis, and M1 macrophage infiltration. Independent of BMI, it has been demonstrated that restricted adipose tissue capillarization and blood flow are correlated to hypoxia and insulin resistance. As a result, increasing angiogenesis and blood flow may be potential targets for enhancing insulin sensitivity. Down-regulation of adipose tissue expression GLO-1 activity in obese individuals' early stages of metabolic disorders shows that inhibiting it may be a potential strategy for improving the vascular and metabolic function of adipose tissue. In adipocytes, GLP-1-based therapy promotes capillarization, GLO-1 activity, and insulin sensitivity.

Finally, it is also possible to communicate with phosphoinositide 3-kinase (PI3K), mitogen-activated protein kinase (MAPK), and Wnt4/ β -catenin. GLP-1 promotes preadipocyte differentiation by upregulating peroxisome proliferator-activated receptor γ (PPAR γ) and fatty acid-binding protein 4 (FABP4) and inhibits adipogenesis in mature adipocytes by decreasing fatty acid synthase (FASN) expression[47]. By boosting the expression and activity of the coenzyme I (NAD⁺) dependent protein deacetylase (Sirt1), the mechanism of which is unclear, GLP-1 also improves white adipose (WAT) lipolysis[48]. Moreover, GLP-1 regulates thermogenesis by inhibiting signaling pathways associated with BMP4 and thereby increasing the production of thermogenic genes including mitochondrial brown lipolytic coupling protein 1 (UCP1)[49]. These mechanism diagrams are shown in Fig. 2.

6.2. Central nervous system signaling pathways

Intestinal-derived GLP-1 activates sensory afferent neurons in the nucleus solitarius through binding to capillaries, the hepatic hila, and nodular ganglion receptors in liver tissue, and through the hypothalamus, activates motor neurons in the vagus nerve, interacting with GLP-1R expressed in the vagus nerve, which transmits information upward through the vagus nerve to the central nervous system (CNS), which in turn inhibits vagal nerve activity and delays gastric emptying, thereby increasing satiety and reducing food intake[50].

Intestinal-derived GLP-1 is involved in the regulation of insulin sensitivity in a CNS-dependent manner[51]. GLP-1 activates vagal sensory neurons projecting to the brainstem in order to initiate efferent vagal reflexes via the parasympathetic branches of the autonomic nervous system. Parasympathetic cells release acetylcholine (ACh) in the islets to activate M-cholinergic receptors and stimulate calcium mobilization in β -cells, and these neurons also release pituitary adenylate cyclase-activating peptide (PACAP), which stimulates β -cell production of cyclic adenosine monophosphate (cAMP), thereby regulating insulin sensitivity.

Intestinal-derived GLP-1 is also implicated in BAT-associated thermogenesis in a CNS-dependent manner, acting through inhibition of the AMPK signaling pathway[52]. AMPK in the ventral medial nucleus of the hypothalamus (VMH) prevents high-fat diet (HFD)-induced obesity by increasing thermogenesis and subsequent energy expenditure in BAT. This effect is dependent on the AMPK α 1 isoform in steroidogenic factor 1 (SF1) neurons of VMH, as mice that selectively ablate AMPK α 1 in SF1 neurons exhibit resistance to diet-induced obesity, increased thermogenesis of BAT, browning of white adipose tissue, and improved glucose and lipid homeostasis.

Two different kinds of neurons in the arcuate nucleus of the hypothalamus (ARC) control energy metabolism: one type, agouti-related protein (AgRP), which increases appetite; the other,

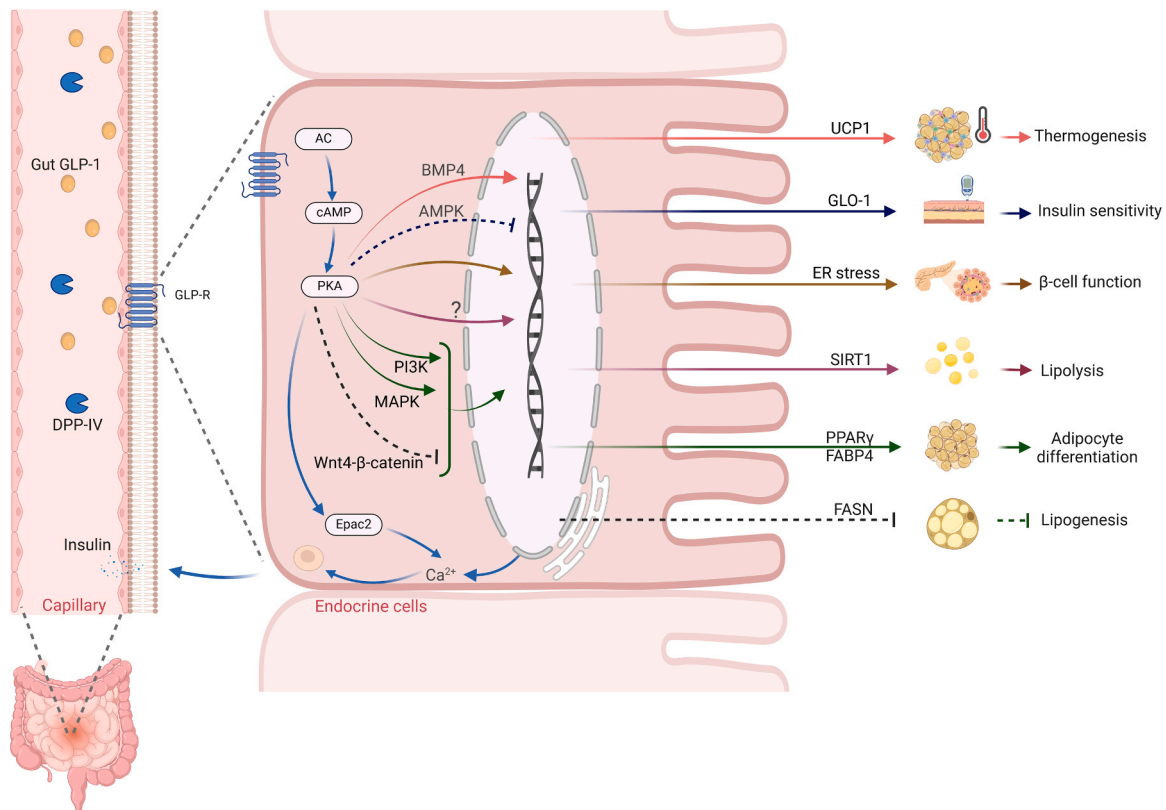


Fig. 2. Peripheral signaling pathway, In the periphery, activation of GLP-R by gut-derived GLP-1 enhances glucose-stimulated insulin secretion via the PKA-dependent or Epac2 pathway. In obese diabetic mice, GLP-1 decreased ER stress, improved β -cell function, and decreased insulin resistance by increasing PKA activity. Also, it increased GLO-1 levels and enhanced insulin sensitivity in peripheral tissues via inhibiting AMPK-related pathways. GLP-1 encourages preadipocyte development by upregulating PPAR γ and FABP4 via interacting with several signaling pathways, such as PI3K, MAPK, and Wnt4/ β -catenin pathways but inhibits adipogenesis in mature adipocytes by decreasing fatty acid synthase expression. Additionally, GLP-1 enhances lipolysis in WAT by strengthening Sirt1 expression and activity via an unidentified mechanism. Furthermore, by inhibiting the BMP4-related signaling pathway, GLP-1 contributes to the regulation of thermogenesis through raising the expression of thermogenic genes like UCP1, GLP-1- glucagon-like peptide-1. GLP-R- glucagon-like peptide-receptor. PKA- protein kinase A. Epac2-guanine nucleotide exchange factor 4. ER- endoplasmic reticulum. GLO-1- glyoxalase-1. AMPK- adenylate-activated protein kinase. PPAR γ - peroxisome proliferator-activated receptor γ . FABP4- fatty acid-binding protein 4. FASN- fatty acid synthase. PI3K- phosphoinositide 3-kinase. MAPK- mitogen-activated protein kinase. WAT- white adipose. BMP4- Bone morphogenetic protein 4. Sirt1- coenzyme I (NAD $^{+}$) dependent protein deacetylase. UCP1-mitochondrial brown lipolytic coupling protein 1.

proopiomelanocortin (POMC), which decreases appetite. The hypothalamus receives the central GLP-1 generated by the caudal neurons of the medulla oblongata, which inhibits AgRP/NPY neurons and activates POMC neurons to decrease appetite[53,54]. The activation of AgRP neurons promotes obesity by stimulating appetite and inhibiting energy expenditure, while inhibiting their activity leads to anorexia and weight loss. In contrast, chronic activation of POMC neurons inhibits food intake and increases metabolic rate, and in addition, the absence of functional mutations in the POMC gene leads to overfeeding and obesity in rodents and humans. POMC neurons prevent obesity by releasing alpha-melanocyte-stimulating hormone (α -MSH), which on the one hand suppresses appetite and on the other hand influences the body's energy dissipation by regulating the excitability of the sympathetic nervous system. These mechanism diagrams are shown in Fig. 3.

7. DPP-IV and adipose tissue distribution

Daniela et al.[55] used a proteomic approach to identify DPP-IV as a novel adipokine released by fully differentiated human adipocytes. The amount of DPP-IV secreted considerably increases as adipocytes differentiate. Comparisons with preadipocytes and macrophages indicate that adipocytes from adipose tissue are the primary source of DPP-IV release from intact organs into the circulation.

Research study[9] in both in vitro and in vivo have shown that the expression of DPP-IV in subcutaneous adipose tissue (SAT) and visceral

adipose tissue (VAT) is positively correlated with BMI, and consistently higher than SAT in VAT. DPP-IV was positively correlated with the number of VAT, adipocyte size, and adipose tissue inflammation. DPP-IV inhibitors increase the rate of oxygen consumption in BAT and inguinal white adipose tissue (iWAT) and enhance BAT function by inhibiting extracellular regulated protein kinases (ERKs) signaling pathway activation of β -agonist induced UCP1 expression[56]. DPP-IV inhibitor therapy prevents obesity by activating BAT function.

8. DPP-IV and ectopic fat deposits

Obesity initiates DPP-IV synthesis and secretion by activating the calcium-calmodulin-dependent protein kinase II-transcriptional activator 4 recombinant protein (Ca-CaMKII-ATF4) signaling pathway in mouse hepatocytes[57]. The interleukin 1 receptor-associated kinase 1 recombinant protein-transforming growth factor kinase 1 (IRAK1--TAK1) signaling pathway is activated when sDPP-IV enters adipose tissue through blood circulation and is transported to the core of adipocytes via caveolin-1 (CAV-1). While this is happening, plasma coagulation Factor Xa in the blood activates RAF1 via protease-activated receptor 2 (PAR2), and TAK1 and RAF1 together cause ERK1/2 and NF- κ B mediated inflammation, causing macrophage infiltration into visceral adipose tissue and further causing insulin resistance in VAT. It was also found that silencing DPP-IV expression in hepatocytes suppressed VAT inflammation and insulin resistance, which was not seen

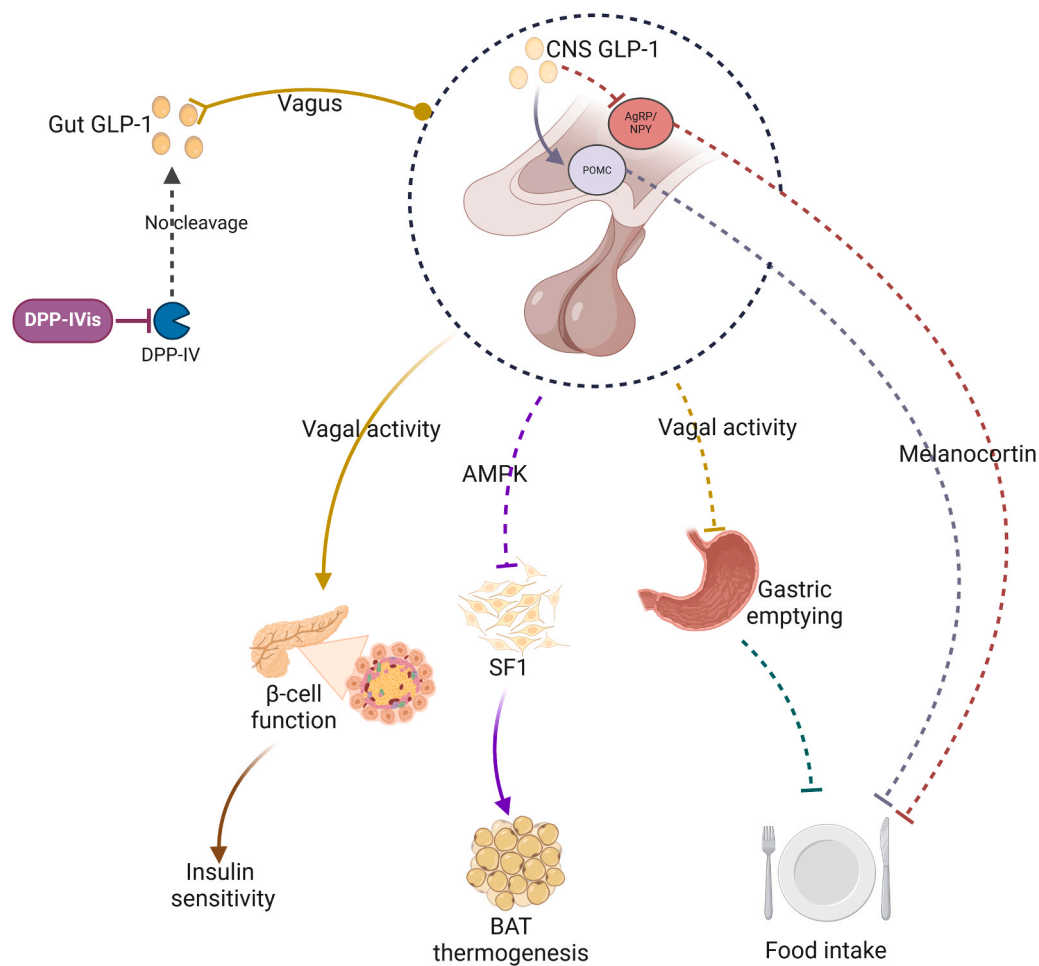


Fig. 3. Central nervous system signaling pathways. Gut-derived GLP-1 and vagus-expressed GLP-R combine to transmit information to the CNS, which in turn reduces vagal activity and stomach emptying to induce satiety and decrease food intake. In addition, gut-derived GLP-1 regulates insulin sensitivity and BAT-related thermogenesis by inhibiting the AMPK signaling pathway in a CNS-dependent manner. GLP-1 activates vagal sensory neurons projecting to the brainstem to initiate efferent vagal reflexes via the parasympathetic branches of the autonomic nervous system, stimulating β -cell production of cAMP and thus regulating insulin sensitivity. By activating POMC neurons while suppressing AgRP/ NPY neurons in the hypothalamus, central GLP-1, which is made by neurons in the caudal medulla, reduces food intake. CNS- central nervous system. cAMP- cyclic adenosine monophosphate. PACAP -pituitary adenylate cyclase-activating peptide. VMH -ventral medial nucleus of the hypothalamus. HFD-high-fat diet. SF1-steroidogenic factor 1. ARC-arcuate nucleus of the hypothalamus. AgRP-agouti-related protein. POMC-proopiomelanocortin. α -MSH-alpha-melanocyte-stimulating hormone.

with oral DPP-IV inhibitors.

sDPP-IV is recommended as a biomarker for NAFLD and has been proven to be a valid indicator of hepatocyte apoptosis and fibrosis[58]. Liver-specific DPP-IV overexpression is associated with higher plasma DPP-IV activity and lower GLP-1 levels. Adiposity and inflammation of adipose tissue increased in response to elevated hepatic DPP-IV activity. Elevated hepatic DPP-IV enhanced peroxisome proliferator-activated receptor- γ (PPAR γ) and fatty acid receptor (CD36) expression in the liver. In both people and mice, hepatocyte-specific expression of PPAR γ is associated with fatty liver and induces hepatic steatosis. CD36 and monoacylglycerol O-acyltransferase 1 (Mogat1), both of which are associated with fatty liver disease, are the primary targets of PPAR γ in the liver.

8.1. DPP-IV and bariatric surgery

BSG is considered a safe and effective alternative therapy for severely obese or overweight individuals with related comorbidities[59]. The mainstream surgical methods are vertical sleeve gastrectomy (VSG) and gastric bypass surgery (RYGB)[60]. A study[61] on glucose metabolism, intestinal hormones, and inflammation after BSG and weight loss medications showed that both groups of patients lost weight. Three months

after BSG, there were significant changes in inflammation (decreased fasting hypersensitive C-reactive protein (hs-CRP)), intestinal peptides (decreased fasting GLP-1 levels, increased levels at 30 and 60 minutes postprandial, increased GIP and glucagon levels at 30 minutes postprandial, and increased gastric hunger levels at 60 minutes postprandial), and glucose metabolism (increased glucose and insulin levels at 30 minutes postprandial) concerning hormone levels. BSG did not substantially change plasma DPP-IV activity, and this finding was unrelated to how much weight each group lost.

Other study[62] examined the changes in circulating sDPP-IV during different weight loss interventions and found that sDPP-IV increased with age and weight, was positively associated with insulin resistance and hypertriglyceridemia, but decreased in T2DM. Additionally, it was discovered that after weight reduction surgery and non-surgical interventions over a 6-month period in obese patients with elevated sDPP-IV levels, serum concentrations of sDPP-IV were not changed by weight loss.

Although the BSG is typically preferable to non-surgery, many patients experience less favorable outcomes than anticipated because of postoperative weight regain (WR)[63]. WR occurs in about 11~22 % of patients after BSG[64]. Numerous studies[15] have been conducted on patients who have undergone BSG that demonstrate the utility of AOM

for inadequate weight reduction or weight regain. However, the findings may vary due to the heterogeneity of the study population and the study itself, as well as the fact that these studies primarily involve retrospective analysis and lack prospective analysis.

8.2. DPP-IV and DPP-IV Inhibitors Improve Obesity-Related Diseases

Numerous studies have revealed that DPP-IV and DPP-IV inhibitors regulate obesity-related diseases in multiple effects, including lowering obesity-related inflammation, enhancing liver steatosis, and reducing cardiovascular risk.

8.2.1. Improving obesity-related inflammation

Individuals with obesity cause the hepatocytes to synthesize and secrete DPP-IV because of the accumulation of excess body fat. DPP-IV and plasma Factor-Xa interact to cause adipose tissue macrophage inflammation. Targeted suppression of DPP-IV expression in hepatocytes reduces inflammation and insulin resistance in VAT[57]. Additionally, by controlling the M1/M2 state of macrophages, DPP-IV contributes significantly to the inflammation and insulin resistance brought on by obesity[65]. DPP-IV is primarily produced by M1 macrophages. By causing macrophages in adipose tissue and the liver to polarize to an anti-inflammatory phenotype, DPP-IV antagonists reduce the inflammation and insulin resistance brought on by obesity.

8.2.2. Modulating inflammatory bowel disease

Inflammatory bowel disease (IBD) is a chronic immune-mediated gastrointestinal disorder, with obesity identified as a significant risk factor associated with its progression[3]. Visceral fat is considered a key contributor to adverse outcomes in patients with IBD. Several prospective studies have evaluated the expression of DPP-IV in small intestine and colon biopsies, as well as surgical specimens from IBD patients and healthy volunteers, demonstrating that DPP-IV may serve as a biomarker for treatment escalation and response to biologic therapy in IBD patients [3,66]. Vilda is a potent DPP-IV inhibitor that has shown promising colonic protective and anti-ulcer properties in studies involving albino rats, effectively countering oxidant-induced ulcerative colitis (UC)[67]. This effect is achieved through the reduction of colonic inflammation, alleviation of oxidative and endoplasmic reticulum stress, improvement of mitochondrial function, and promotion of colonic autophagy via the modulation of the PI3K/AKT/mTOR and AMPK/Nrf2 signaling pathways. These findings suggest that Vilda may represent a promising therapeutic option for UC.

8.2.3. Improving hepatic steatosis

NAFLD has become a metabolic liver disease that affects about a quarter of the world's population[68]. Obesity is an independent risk factor for NAFLD[69]. The degree of obesity and fat distribution affect the prevalence of NAFLD. In animal studies[34,70], increased levels of hepatic DPP-IV expression promote the development of NAFLD. Several studies[71,72] also have reported the therapeutic potential of DPP-IV inhibitors for the treatment of non-alcoholic steatohepatitis (NASH) in animals and humans. DPP-IV inhibitors ameliorate the pathogenesis of NASH, inhibit adipose apoptosis, and downregulate hepatic DPP-IV expression. In addition, pro-inflammatory and pro-fibrogenic factors are reduced by DPP-IV inhibitors both in vivo and in vitro, indicating that DPP-IV inhibitors alter the progression of NASH. Furthermore[73], hematopoietic stem cell modulation can also contribute to the protective effect of DPP-IV inhibitors against NASH. Another study[74] has reported that DPP-IV inhibitors have been proven to induce autophagy, which is generally considered to prevent fatty liver, degrade damaged cell components in liver cells, and improve liver steatosis.

8.2.4. Reducing the risk of cardiovascular disease

Being overweight causes the accumulation of visceral fat. Compression of the viscera and blood arteries will result in an increase in blood

pressure, cholesterol, triglycerides, and low-density lipoprotein (LDL) levels. Chronic hypertension and hyperlipidemia are risk factors for atherosclerosis, coronary heart disease, and stroke. DPP-IV was found to be widely expressed in the vascular system, including endothelial cells, macrophages, cardiomyocytes, smooth muscle cells, valvular interstitial cells, and other cell types, suggesting its possible involvement in the development and progression of cardiovascular disease[75]. Five large-scale clinical trials[76–80] have been completed to demonstrate the cardiovascular safety of DPP-IV inhibitors. More and more studies [81–83] have found that DPP-IV inhibitors also have protective effects on a variety of cardiovascular diseases, such as hyperlipidemia, hypertension, aortic valve calcification, coronary atherosclerosis, and heart failure. A randomized placebo-controlled mechanistic study known as the "Vildagliptin in Ventricular Dysfunction Diabetes Trial" (VIVID) demonstrated that the DPP-IV inhibitor vildagliptin significantly improved outcomes in 254 patients with T2DM, heart failure, and systolic dysfunction when used in conjunction with standard glycemic and heart failure therapies[84]. In another study[85], researchers from the UK, Greece, and Germany identified a method for treating insulin-induced vascular injury associated with heart disease. They utilized DPP-IV inhibitors to treat both mice and human patients, successfully reducing oxidative stress responses in the vasculature and reversing vascular damage caused by a high-fat diet in both humans and mice. Wang et al.[86] found that DUSP26 contributes to the treatment of calcific aortic valve disease (CAVD) by inhibiting DPP-IV ubiquitination and degradation induced by Mouse Double Minute 2 (MDM2) in human valvular interstitial cells (hVICs). Katia et al.[87] discovered that the DPP-IV inhibitor sitagliptin stimulates resident cardiac stem cells and promotes neovascularization in patients with diabetic cardiomyopathy, reducing cardiac remodeling, improving myocardial function, and enhancing survival rates following myocardial infarction.

8.3. DPP-IV and DPP-IV Inhibitors Reduce the Risk of COVID-19

Patients with chronic underlying diseases are more likely to be infected with COVID-19 during the recent worldwide pandemic of COVID-19. According to studies[88,89], being overweight and obese may also be risk factors for COVID-19. COVID-19 is caused by Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2). sDPP-IV can bind SARS-CoV-2 and prevent the virus from attaching to membrane-bound DPP-IV in host cells, thus preventing virus transmission. A German study[90] shows reduced circulating levels of sDPP-IV in patients with severe neo-coronary pneumonia. Naveen et al. [91] predicted the molecular model of SARS-CoV-2, they found that the S1 domain of spike glycoprotein of SARS-CoV-2 may directly interact with DPP-IV in host cells, indicating that DPP-IV may be a common receptor for SARS-CoV-2 virus entry. In various clinical chronic diseases such as diabetes, obesity, and metabolic syndrome, serum sDPP-IV levels are also changed and related to insulin resistance[92]. These studies may also contribute to greater susceptibility to SARS-CoV-2 infection in diabetes, obesity, and the elderly. By raising sDPP-IV levels to improve the sequestration of viral particles in circulation, DPP-IV inhibitors can prevent viral entry into the body and reduce viral growth in humans [93]. The immunomodulatory effect of DPP-IV inhibitor can prevent cytokine storms in patients with COVID-19, thereby reducing the severity of the disease[94]. The importance of DPP-IV and DPP-IV inhibitors in COVID-19 has gradually been recognized.

9. Conclusion

In 1948[95], obesity was first included in the International Classification of Diseases, but did not attract much attention. In 1997, the World Health Organization (WHO) listed obesity as a chronic disease. In 2013[96], the American Medical Association passed a motion describing obesity as a "disease state with multiple pathophysiological aspects". In 2021[97], the European Commission issued a brief defining obesity as "a

chronic recurrent disease and thus a gateway to a range of other non-communicable diseases". These fully emphasize that obesity is not only a risk factor for various diseases but that obesity itself is an independent chronic disease. At present, the global prevalence of obesity continues to increase. WHO[98] estimates that by 2025, one in five adults worldwide will have obesity.

One environmental factor contributing to the rapid rise in obesity worldwide is increased access to high-energy foods and a decline in physical activity (Fig. 1). Antiepileptic and psychotropic drugs affect the regulation of appetite by the central nervous system. Chronic stress, sleep deprivation, and misaligned circadian cycles can all cause the body's endocrine system to become dysregulated. These factors may also contribute to obesity. The emergence of simple obesity in people also has a genetic basis. Obesity is a metabolic disease associated with multiple factors.

The management of AOM has a long history, full of hopes and disappointments. Comorbid conditions and the increased risk of vascular disease in obese individuals make it more difficult to evaluate the safety of AOM. Furthermore, it is very challenging to carry out extensive, long-term clinical studies on individuals who have heterologous obesity. Through its enzyme activity, the molecule DPP-IV is crucial in controlling metabolic balance and inflammatory responses. Targeting DPP-IV, it treats obesity by enhancing the activity of the substrate GLP-1, stimulating insulin release, delaying gastric emptying, and increasing satiety, among other mechanisms. In addition, by improving adipose tissue inflammation, promoting adipose tissue remodeling, and reducing ectopic adipose tissue deposition, it also regulates adipose tissue's function in metabolism and lipid homeostasis. Since BSG results in altered gastrointestinal hormone secretion, but not circulating DPP-IV levels, the possibility of combining treatment targeting DPP-IV after bariatric surgery to enhance the anti-obesity effect requires long-term, large-scale, multicenter clinical studies to evaluate its safety and efficacy. The mechanisms of action based on DPP-IV targets in obesity and metabolic balance are reviewed in this study, as well as their practical application in the management of chronic disorders linked to obesity.

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Author contributions

All the authors participated in writing the manuscript and in drawing the figures. All authors read and approved the final manuscript.

CRedit authorship contribution statement

Huolun Feng: Writing – review & editing, Validation, Supervision, Methodology, Investigation. **Jiabin Zheng:** Writing – review & editing, Validation, Supervision, Project administration. **Liyang Cai:** Writing – review & editing, Writing – original draft, Visualization, Validation, Methodology, Investigation. **Yong Li:** Writing – review & editing, Supervision, Project administration, Methodology, Funding acquisition, Formal analysis. **Xin Guo:** Writing – review & editing, Writing – original draft, Visualization, Validation, Project administration, Methodology, Investigation.

Declaration of Competing Interest

The authors declare no competing interests.

Data availability

No data was used for the research described in the article.

References

- [1] NCD Risk Factor Collaboration (NCD-RisC), Worldwide trends in underweight and obesity from 1990 to 2022: a pooled analysis of 3663 population-representative studies with 222 million children, adolescents, and adults, *Lancet* 403 (2024) 1027–1050. [https://doi.org/10.1016/S0140-6736\(23\)02750-2](https://doi.org/10.1016/S0140-6736(23)02750-2).
- [2] N. Alamuddin, Z. Bakizada, T.A. Wadden, Management of obesity, *J. Clin. Oncol.* 34 (2016) 4295–4305. <https://doi.org/10.1200/JCO.2016.66.8806>.
- [3] P. Sehgal, S. Su, J. Zech, Y. Nobel, L. Luk, I. Economou, B. Shen, J.D. Lewis, D. E. Freedberg, Visceral Adiposity Independently Predicts Time to Flare in Inflammatory Bowel Disease but Body Mass Index Does Not, *Inflamm. Bowel Dis.* 30 (2024) 594–601. <https://doi.org/10.1093/ibd/izad111>.
- [4] K. Si, Y. Hu, M. Wang, C.M. Apovian, J.E. Chavarro, Q. Sun, Weight loss strategies, weight change, and type 2 diabetes in US health professionals: a cohort study, *PLoS Med* 19 (2022) e1004094. <https://doi.org/10.1371/journal.pmed.1004094>.
- [5] R.G. Bianchetti, C.J. Lavie, F. Lopez-Jimenez, Challenges in cardiovascular evaluation and management of obese patients: JACC state-of-the-art review, *J. Am. Coll. Cardiol.* 81 (2023) 490–504. <https://doi.org/10.1016/j.jacc.2022.11.031>.
- [6] M.J. Belanger, M.A. Hill, A.M. Angelidi, M. Dalamaga, J.R. Sowers, C.S. Mantzoros, Covid-19 and disparities in nutrition and obesity, *N. Engl. J. Med* 383 (2020) e69. <https://doi.org/10.1056/NEJMp2021264>.
- [7] N. Murphy, C.C. Newton, M. Song, N. Papadimitriou, M. Hoffmeister, A.I. Phipps, T.A. Harrison, P.A. Newcomb, E.K. Aglago, S.I. Berndt, H. Brenner, D.D. Buchanan, Y. Cao, A.T. Chan, X. Chen, I. Cheng, J. Chang-Claude, N. Dimou, D. Drew, A. B. Farris, A.J. French, S. Gallinger, P. Georgeson, M. Giannakis, G.G. Giles, S. B. Gruber, S. Harlid, L. Hsu, W.-Y. Huang, M.A. Jenkins, R.S. Laskar, L. Le Marchand, P. Limburg, Y. Lin, M. Mandic, J.A. Nowak, M. Obón-Santacana, S. Ogino, C. Qu, L.C. Sakoda, R.E. Schoen, M.C. Southey, Z.K. Stadler, R. S. Steinfeld, W. Sun, S.N. Thibodeau, A.E. Toland, Q.M. Trinh, K.K. Tsilidis, T. Ugai, B. Van Guelpen, X. Wang, M.O. Woods, S.H. Zaidi, M.J. Gunter, U. Peters, P.T. Campbell, Body mass index and molecular subtypes of colorectal cancer, *J. Natl. Cancer Inst.* 115 (2023) 165–173. <https://doi.org/10.1093/jnci/djac215>.
- [8] M. Sun, J. Fritz, C. Häggström, T. Bjørge, G. Nagel, J. Manjer, A. Engeland, E. Zitt, B. van Guelpen, P. Stattin, H. Ulmer, T. Stocks, Metabolically (un)healthy obesity and risk of obesity-related cancers: a pooled study, *J. Natl. Cancer Inst.* (2023) djad008. <https://doi.org/10.1093/jnci/djad008>.
- [9] H. Sell, M. Blüher, N. Klötting, R. Schlich, M. Willems, F. Ruppe, W.T. Knoefel, A. Dietrich, B.A. Fielding, P. Arner, K.N. Frayn, J. Eckel, Adipose dipeptidyl peptidase-4 and obesity: correlation with insulin resistance and depot-specific release from adipose tissue in vivo and in vitro, *Diabetes Care* 36 (2013) 4083–4090. <https://doi.org/10.2337/dc13-0496>.
- [10] I. Barchetta, F.A. Cimmini, S. Dule, M.G. Cavallo, Dipeptidyl Peptidase 4 (DPP4) as a novel adipokine: role in metabolism and fat homeostasis, *Biomedicines* 10 (2022) 2306. <https://doi.org/10.3390/biomedicines10092306>.
- [11] A.L. João, F. Reis, R. Fernandes, The incretin system ABCs in obesity and diabetes - novel therapeutic strategies for weight loss and beyond, *Obes. Rev.* 17 (2016) 553–572. <https://doi.org/10.1111/obr.12421>.
- [12] B. Crooks, N.S. Stamataki, J.T. McLaughlin, Appetite, the enteroendocrine system, gastrointestinal disease and obesity, *Proc. Nutr. Soc.* 80 (2021) 50–58. <https://doi.org/10.1017/S0029665120006965>.
- [13] M.M. Conley, C.M. McFarlane, D.W. Johnson, J.T. Kelly, K.L. Campbell, H. L. MacLaughlin, Interventions for weight loss in people with chronic kidney disease who are overweight or obese, *Cochrane Database Syst. Rev.* 3 (2021) CD013119. <https://doi.org/10.1002/14651858.CD013119.pub2>.
- [14] H. Shaikh, P. Bradhurst, L.X. Ma, S.Y.C. Tan, S.J. Egger, J.L. Vardy, Body weight management in overweight and obese breast cancer survivors, *Cochrane Database Syst. Rev.* 12 (2020) CD012110. <https://doi.org/10.1002/14651858.CD012110.pub2>.
- [15] I.P. Redmond, A.P. Shukla, L.J. Aronne, Use of weight loss medications in patients after bariatric surgery, *Curr. Obes. Rep.* 10 (2021) 81–89. <https://doi.org/10.1007/s13679-021-00425-1>.
- [16] R. Howard, G.F. Chao, J. Yang, J. Thumma, K. Chhabra, D.E. Arterburn, A. Ryan, D.A. Telem, J.B. Dimick, Comparative safety of sleeve gastrectomy and gastric bypass up to 5 years after surgery in patients with severe obesity, *JAMA Surg.* 156 (2021) 1160–1169. <https://doi.org/10.1001/jamasurg.2021.4981>.
- [17] K. Johansson, P.-A. Svensson, J. Söderling, M. Peltonen, M. Neovius, L.M. S. Carlsson, K. Sjöholm, Long-term risk of anaemia after bariatric surgery: results from the Swedish Obese Subjects study, *Lancet Diabetes Endocrinol.* 9 (2021) 515–524. [https://doi.org/10.1016/S2213-8587\(21\)00141-8](https://doi.org/10.1016/S2213-8587(21)00141-8).
- [18] Q. Shi, Y. Wang, Q. Hao, P.O. Vandvik, G. Guyatt, J. Li, Z. Chen, S. Xu, Y. Shen, L. Ge, F. Sun, L. Li, J. Yu, K. Nong, X. Zou, S. Zhu, C. Wang, S. Zhang, Z. Qiao, Z. Jian, Y. Li, X. Zhang, K. Chen, F. Qu, Y. Wu, Y. He, H. Tian, S. Li, Pharmacotherapy for adults with overweight and obesity: a systematic review and network meta-analysis of randomised controlled trials, *Lancet* 399 (2022) 259–269. [https://doi.org/10.1016/S0140-6736\(21\)01640-8](https://doi.org/10.1016/S0140-6736(21)01640-8).
- [19] Y.J. Tak, S.Y. Lee, Anti-obesity drugs: long-term efficacy and safety: an updated review, *World J. Mens. Health* 39 (2021) 208–221. <https://doi.org/10.5534/wjmh.200010>.
- [20] E. Piliitsi, O.M. Farr, S.A. Polyzos, N. Perakakis, E. Nolen-Doerr, A.-E. Papanasiou, C.S. Mantzoros, Pharmacotherapy of obesity: available medications and drugs under investigation, *Metabolism* 92 (2019) 170–192. <https://doi.org/10.1016/j.metabol.2018.10.010>.
- [21] R. Aggarwal, M. Vaduganathan, N. Chiu, D.L. Bhatt, Potential implications of the FDA approval of semaglutide for overweight and obese adults in the United States, *Prog. Cardiovasc. Dis.* 68 (2021) 97–98. <https://doi.org/10.1016/j.pcad.2021.09.007>.

- [22] V.K. Hopsu-Havu, G.G. Glenner, A new dipeptide naphthylamidase hydrolyzing glycyl-prolyl-beta-naphthylamide, *Histochemie* 7 (1966) 197–201, <https://doi.org/10.1007/BF00577838>.
- [23] A.J. Ulmer, T. Mattern, A.C. Feller, E. Heymann, H.D. Flad, CD26 antigen is a surface dipeptidyl peptidase IV (DPPIV) as characterized by monoclonal antibodies clone TII-19-4-7 and 4ELIC7, *Scand. J. Immunol.* 31 (1990) 429–435, <https://doi.org/10.1111/j.1365-3083.1990.tb02789.x>.
- [24] P.M.-K. Tang, Y.-Y. Zhang, J.S.-C. Hung, J.Y.-F. Chung, X.-R. Huang, K.-F. To, H.-Y. Lan, DPP4/CD32b/NF- κ B Circuit: a novel druggable target for inhibiting CRP-driven diabetic nephropathy, *Mol. Ther.* 29 (2021) 365–375, <https://doi.org/10.1016/j.yimthe.2020.08.017>.
- [25] S. Senkel, B. Lucas, L. Klein-Hitpass, G.U. Ryffel, Identification of target genes of the transcription factor HNF1beta and HNF1alpha in a human embryonic kidney cell line, *Biochim Biophys. Acta* 1731 (2005) 179–190, <https://doi.org/10.1016/j.bbexp.2005.10.003>.
- [26] J. Du, L. Fu, F. Ji, C. Wang, S. Liu, X. Qiu, FosB recruits KAT5 to potentiate the growth and metastasis of papillary thyroid cancer in a DPP4-dependent manner, *Life Sci.* 259 (2020) 118374, <https://doi.org/10.1016/j.lfs.2020.118374>.
- [27] B. Bauvois, M. Djavaheri-Mergny, D. Rouillard, J. Dumont, J. Wietzerbin, Regulation of CD26/DPPIV gene expression by interferons and retinoic acid in tumor B cells, *Oncogene* 19 (2000) 265–272, <https://doi.org/10.1038/sj.onc.1203292>.
- [28] W.A. Weihofen, J. Liu, W. Reutter, W. Saenger, H. Fan, Crystal structure of CD26/dipeptidyl-peptidase IV in complex with adenosine deaminase reveals a highly amphiphilic interface, *J. Biol. Chem.* 279 (2004) 43330–43335, <https://doi.org/10.1074/jbc.M405001200>.
- [29] H.B. Rasmussen, S. Branner, F.C. Wiberg, N. Wagtmann, Crystal structure of human dipeptidyl peptidase IV/CD26 in complex with a substrate analog, *Nat. Struct. Biol.* 10 (2003) 19–25, <https://doi.org/10.1038/nsb882>.
- [30] A. Mortier, M. Gouwy, J. Van Damme, P. Proost, S. Struyf, CD26/dipeptidylpeptidase IV-chemokine interactions: double-edged regulation of inflammation and tumor biology, *J. Leukoc. Biol.* 99 (2016) 955–969, <https://doi.org/10.1189/jlb.3MR0915-401R>.
- [31] A.M. Elmansi, M.E. Awad, N.H. Eisa, D. Kondrikov, K.A. Hussein, A. Aguilar-Pérez, S. Herberg, S. Periyasamy-Thandavan, S. Fulzele, M.W. Hamrick, M.E. McGee-Lawrence, C.M. Isales, B.F. Volkman, W.D. Hill, What doesn't kill you makes you stranger: Dipeptidyl peptidase-4 (CD26) proteolysis differentially modulates the activity of many peptide hormones and cytokines generating novel cryptic bioactive ligands, *Pharm. Ther.* 198 (2019) 90–108, <https://doi.org/10.1016/j.pharmthera.2019.02.005>.
- [32] K. Ohnuma, R. Hatano, C. Morimoto, DPP4 in anti-tumor immunity: going beyond the enzyme, *Nat. Immunol.* 16 (2015) 791–792, <https://doi.org/10.1038/ni.3210>.
- [33] S. Anoop, A. Misra, S.P. Bhatt, S. Gulati, R.M. Pandey, H. Mahajan, High circulating plasma dipeptidyl peptidase-4 levels in non-obese Asian Indians with type 2 diabetes correlate with fasting insulin and LDL-C levels, triceps skinfolds, total intra-abdominal adipose tissue volume and presence of diabetes: a case-control study, *BMJ Open Diabetes Res Care* 5 (2017) e000393, <https://doi.org/10.1136/bmjopen-2017-000393>.
- [34] M. Lee, E. Shin, J. Bae, Y. Cho, J.-Y. Lee, Y.-H. Lee, B.-W. Lee, E.S. Kang, B.-S. Cha, Dipeptidyl peptidase-4 inhibitor protects against non-alcoholic steatohepatitis in mice by targeting TRAIL receptor-mediated lipooapoptosis via modulating hepatic dipeptidyl peptidase-4 expression, *Sci. Rep.* 10 (2020) 19429, <https://doi.org/10.1038/s41598-020-75288-y>.
- [35] S. Blauschmidt, T. Greither, K. Lampe, S. Köller, P. Kaltwaßer, H.M. Behre, Dipeptidyl peptidase 4 serum activity and concentration are increased in women with polycystic ovary syndrome, *Clin. Endocrinol. (Oxf.)* 87 (2017) 741–747, <https://doi.org/10.1111/cen.13444>.
- [36] X. Hu, S. Chen, C. Xie, Z. Li, Z. Wu, Z. You, DPP4 gene silencing inhibits proliferation and epithelial-mesenchymal transition of papillary thyroid carcinoma cells through suppression of the MAPK pathway, *J. Endocrinol. Invest* 44 (2021) 1609–1623, <https://doi.org/10.1007/s40618-020-01455-7>.
- [37] J. Huang, X. Liu, Y. Wei, X. Li, S. Gao, L. Dong, X. Rao, J. Zhong, Emerging role of dipeptidyl peptidase-4 in autoimmune disease, *Front Immunol.* 13 (2022) 830863, <https://doi.org/10.3389/fimmu.2022.830863>.
- [38] S.N. Chaudhari, D.A. Harris, H. Aliakbarian, J.N. Luo, M.T. Henke, R. Subramaniam, A.H. Vernon, A. Tavakkoli, E.G. Sheu, A.S. Devlin, Bariatric surgery reveals a gut-restricted TGR5 agonist with anti-diabetic effects, *Nat. Chem. Biol.* 17 (2021) 20–29, <https://doi.org/10.1038/s41589-020-0604-z>.
- [39] X. Wen, B. Zhang, B. Wu, H. Xiao, Z. Li, R. Li, X. Xu, T. Li, Signaling pathways in obesity: mechanisms and therapeutic interventions, *Signal Transduct. Target Ther.* 7 (2022) 298, <https://doi.org/10.1038/s41392-022-01149-x>.
- [40] D.I. Brierley, M.K. Holt, A. Singh, A. de Araujo, M. McDougle, M. Vergara, M. H. Afaghani, S.J. Lee, K. Scott, C. Maske, W. Langhans, E. Krause, A. de Kloet, F. M. Gribble, F. Reimann, L. Rinaman, G. de Lartigue, S. Trapp, Central and peripheral GLP-1 systems independently suppress eating, *Nat. Metab.* 3 (2021) 258–273, <https://doi.org/10.1038/s42255-021-00344-4>.
- [41] L.L. Baggio, D.J. Drucker, Biology of incretins: GLP-1 and GIP, *Gastroenterology* 132 (2007) 2131–2157, <https://doi.org/10.1053/j.gastro.2007.03.054>.
- [42] J. Yong, V.S. Parekh, S.M. Reilly, J. Nayak, Z. Chen, C. Lebeaupin, I. Jang, J. Zhang, T.P. Prakash, H. Sun, S. Murray, S. Guo, J.E. Ayala, L.S. Satin, A. R. Sattler, R.J. Kaufman, Chop/Ddit3 depletion in β cells alleviates ER stress and corrects hepatic steatosis in mice, *Sci. Transl. Med.* 13 (2021) eba9796, <https://doi.org/10.1126/scitranslmed.aba9796>.
- [43] R. Burcelin, V. Crivelli, C. Perrin, A.Da Costa, J. Mu, B.B. Kahn, M.J. Birnbaum, C. R. Kahn, P. Vollenweider, B. Thorens, GLUT4, AMP kinase, but not the insulin receptor, are required for hepatportal glucose sensor-stimulated muscle glucose utilization, *J. Clin. Invest* 111 (2003) 1555–1562, <https://doi.org/10.1172/JCI16888>.
- [44] S. Herzog, R.J. Shaw, AMPK: guardian of metabolism and mitochondrial homeostasis, *Nat. Rev. Mol. Cell Biol.* 19 (2018) 121–135, <https://doi.org/10.1038/nrm.2017.95>.
- [45] R.S. Ahima, Adipose tissue as an endocrine organ, *Obes. (Silver Spring)* 14 (Suppl 5) (2006) 242S–249S, <https://doi.org/10.1038/oby.2006.317>.
- [46] T. Rodrigues, P. Borges, L. Mar, D. Marques, M. Albano, H. Eickhoff, C. Carrêlo, B. Almeida, S. Pires, M. Abrantes, B. Martins, C. Uriarte, F. Botelho, P. Gomes, S. Silva, R. Seça, P. Matafofe, GLP-1 improves adipose tissue glyoxalase activity and capillarization improving insulin sensitivity in type 2 diabetes, *Pharm. Res* 161 (2020) 105198, <https://doi.org/10.1016/j.phrs.2020.105198>.
- [47] J. Chen, H. Zhao, X. Ma, Y. Zhang, S. Lu, Y. Wang, C. Zong, D. Qin, Y. Wang, Y. Yingfeng Yang, X. Wang, Y. Liu, GLP-1/GLP-1R signaling in regulation of adipocyte differentiation and lipogenesis, *Cell Physiol. Biochem* 42 (2017) 1165–1176, <https://doi.org/10.1159/000478872>.
- [48] F. Xu, B. Lin, X. Zheng, Z. Chen, H. Cao, H. Xu, H. Liang, J. Weng, GLP-1 receptor agonist promotes brown remodelling in mouse white adipose tissue through SIRT1, *Diabetologia* 59 (2016) 1059–1069, <https://doi.org/10.1007/s00125-016-3896-5>.
- [49] X. Wang, B. Ma, J. Chen, H. You, C. Sheng, P. Yang, S. Qu, Glucagon-like Peptide-1 improves fatty liver and enhances thermogenesis in brown adipose tissue via inhibiting BMP4-related signaling pathway in high-fat-diet-induced obese mice, *Int J. Endocrinol.* 2021 (2021) 6620289, <https://doi.org/10.1155/2021/6620289>.
- [50] S. Trapp, D.I. Brierley, Brain GLP-1 and the regulation of food intake: GLP-1 action in the brain and its implications for GLP-1 receptor agonists in obesity treatment, *Br. J. Pharm.* 179 (2022) 557–570, <https://doi.org/10.1111/bph.15638>.
- [51] Z. Huang, L. Liu, J. Zhang, K. Conde, J. Phansalkar, Z. Li, L. Yao, Z. Xu, W. Wang, J. Zhou, G. Bi, F. Wu, R.J. Seeley, M.M. Scott, C. Zhan, Z.P. Pang, J. Liu, Glucose-sensing glucagon-like peptide-1 receptor neurons in the dorsomedial hypothalamus regulate glucose metabolism, *Sci. Adv.* 8 (2022) eabn5345, <https://doi.org/10.1126/sciadv.abn5345>.
- [52] E. Milbank, N.R.V. Dragano, I. González-García, M.R. Garcia, V. Rivas-Limeres, L. Perdomo, G. Hilaret, F. Ruiz-Pino, P. Mallego, D.A. Morgan, R. Iglesias-Rey, C. Contreras, L. Vergori, J. Cunaño, B. Porteiro, A. Gavalda-Navarro, R. Oelkrug, A. Vidal, J. Roa, T. Sobrino, F. Villarroya, C. Diéguez, R. Nogueiras, C. García-Cáceres, M. Tena-Sempere, J. Mittag, M. Carmen Martínez, K. Rahmouni, R. Andriantsitohaina, M. López, Small extracellular vesicle-mediated targeting of hypothalamic AMPK α 1 corrects obesity through BAT activation, *Nat. Metab.* 3 (2021) 1415–1431, <https://doi.org/10.1038/s42255-021-00467-8>.
- [53] Z. He, Y. Gao, L. Lieu, S. Afrin, J. Cao, N.J. Michael, Y. Dong, J. Sun, H. Guo, K. W. Williams, Direct and indirect effects of liraglutide on hypothalamic POMC and NPY/AgRP neurons - Implications for energy balance and glucose control, *Mol. Metab.* 28 (2019) 120–134, <https://doi.org/10.1016/j.molmet.2019.07.008>.
- [54] Y. Dong, J. Carty, N. Goldstein, Z. He, E. Hwang, D. Chau, B. Wallace, A. Kabahizi, L. Lieu, Y. Peng, Y. Gao, L. Hu, J.N. Betley, K.W. Williams, Time and metabolic state-dependent effects of GLP-1R agonists on NPY/AgRP and POMC neuronal activity in vivo, *Mol. Metab.* 54 (2021) 101352, <https://doi.org/10.1016/j.molmet.2021.101352>.
- [55] D. Lamers, S. Famulla, N. Wronkowitz, S. Hartwig, S. Lehr, D.M. Ouwens, K. Eckardt, J.M. Kaufman, M. Ryden, S. Müller, F.-G. Hanisch, J. Ruige, P. Arner, H. Sell, J. Eckel, Dipeptidyl peptidase 4 is a novel adipokine potentially linking obesity to the metabolic syndrome, *Diabetes* 60 (2011) 1917–1925, <https://doi.org/10.2337/db10-1707>.
- [56] K. Takeda, H. Sawazaki, H. Takahashi, Y.-S. Yeh, H.-F. Jheng, W. Nomura, T. Ara, N. Takahashi, S. Seno, N. Osato, H. Matsuda, T. Kawada, T. Goto, The dipeptidyl peptidase-4 (DPP-4) inhibitor teneligliptin enhances brown adipose tissue function, thereby preventing obesity in mice, *FEBS Open Bio* 8 (2018) 1782–1793, <https://doi.org/10.1002/2211-5463.12498>.
- [57] G. Ds, L. O, Z. Z, N. Sm, Y. S, E. C, M. B, C. Mp, I. T, Hepatocyte-secreted DPP4 in obesity promotes adipose inflammation and insulin resistance, *Nature* 555 (2018), <https://doi.org/10.1038/nature26138>.
- [58] C. Baumeier, L. Schlüter, S. Sausenthaler, T. Laeger, M. Rödigier, S.A. Alaze, L. Fritsche, H.-U. Häring, N. Stefan, A. Fritsche, R.W. Schwenk, A. Schürmann, Elevated hepatic DPP4 activity promotes insulin resistance and non-alcoholic fatty liver disease, *Mol. Metab.* 6 (2017) 1254–1263, <https://doi.org/10.1016/j.molmet.2017.07.016>.
- [59] W. Al-Najim, N.G. Docherty, C.W. le Roux, Food intake and eating behavior after bariatric surgery, *Physiol. Rev.* 98 (2018) 1113–1141, <https://doi.org/10.1152/physrev.00021.2017>.
- [60] D.E. Arterburn, D.A. Telem, R.F. Kushner, A.P. Courcoulas, Benefits and risks of bariatric surgery in adults: a review, *JAMA* 324 (2020) 879–887, <https://doi.org/10.1001/jama.2020.12567>.
- [61] T.G. Braga, M.D. Graças Coelho de Souza, M. Menezes, J.F. Nogueira Neto, L. Dellatorre-Teixeira, E. Bouskela, C.W. le Roux, L.G. Kraemer-Aguiar, Dipeptidyl peptidase-4 activity, lipopolysaccharide, C-reactive protein, glucose metabolism, and gut peptides 3 months after bariatric surgery, *Surg. Obes. Relat. Dis.* 17 (2021) 113–120, <https://doi.org/10.1016/j.soard.2020.08.030>.
- [62] N. Rohmann, K. Schlicht, C. Geisler, T. Hollstein, C. Knappe, L. Krause, S. Hagen, A. Beckmann, A.K. Seoudy, P. Wietzke-Braun, K. Hartmann, D. Schulte, K. Türk, J. Beckmann, W. von Schönfels, F.A. Hägele, A. Bopsy-Westphal, A. Franke, S. Schreiber, M. Laudes, Circulating sDPP-4 is increased in obesity and insulin resistance but is not related to systemic metabolic inflammation, *J. Clin. Endocrinol. Metab.* 106 (2021) e592–e601, <https://doi.org/10.1210/clinem/dgaa758>.

- [63] K. McInnis, J.L. Brown, G. Finlayson, R. Dent, É. Doucet, Appetite Changes in Weight Regain and Weight Maintenance After Roux-en-Y Gastric Bypass, *Obes. Surg.* 32 (2022) 1–12, <https://doi.org/10.1007/s11695-022-06061-5>.
- [64] S.F. Noria, R.D. Shelby, K.D. Atkins, N.T. Nguyen, K.M. Gadde, Weight regain after bariatric surgery: scope of the problem, causes, prevention, and treatment, *Curr. Diab. Rep.* 23 (2023) 31–42, <https://doi.org/10.1007/s11892-023-01498-z>.
- [65] F. Zhuge, Y. Ni, M. Nagashimada, N. Nagata, L. Xu, N. Mukaida, S. Kaneko, T. Ota, DPP-4 Inhibition by Linagliptin Attenuates Obesity-Related Inflammation and Insulin Resistance by Regulating M1/M2 Macrophage Polarization, *Diabetes* 65 (2016) 2966–2979, <https://doi.org/10.2337/db16-0317>.
- [66] P. Pinto-Lopes, J. Afonso, R. Pinto-Lopes, C. Rocha, P. Lago, R. Gonçalves, H. Tavares De Sousa, G. Macedo, C. Camila Dias, F. Magro, Serum dipeptidyl peptidase 4: a predictor of disease activity and prognosis in inflammatory bowel disease, *Inflamm. Bowel Dis.* 26 (2020) 1707–1719, <https://doi.org/10.1093/ibd/izz319>.
- [67] M.M. Awad, R.M. El-Gohary, S. Ibrahim, M.T. Abdel Ghafar, E.E. Farghal, A. Aboalsoud, R.A.A. El-Shaer, Potential mitigating impact of a dipeptidyl peptidase-IV inhibitor, vildagliptin, on oxazolone-induced ulcerative colitis: targeting the role of PI3K/AKT/mTOR and AMPK/Nrf2 signaling pathways, *Int Immunopharmacol.* 133 (2024) 112110, <https://doi.org/10.1016/j.intimp.2024.112110>.
- [68] Z.M. Younossi, P. Golabi, L. de Avila, J.M. Paik, M. Srishord, N. Fukui, Y. Qiu, L. Burns, A. Afendy, F. Nader, The global epidemiology of NAFLD and NASH in patients with type 2 diabetes: a systematic review and meta-analysis, *J. Hepatol.* 71 (2019) 793–801, <https://doi.org/10.1016/j.jhep.2019.06.021>.
- [69] E. Lee, H. Korf, A. Vidal-Puig, An adipocentric perspective on the development and progression of non-alcoholic fatty liver disease, 00077–6, *J. Hepatol.* S0168-8278 (23) (2023), <https://doi.org/10.1016/j.jhep.2023.01.024>.
- [70] J.C. Bae, DPP-4 inhibitor in type 2 diabetes mellitus patient with non-alcoholic fatty liver disease: achieving two goals at once? *Endocrinol. Metab. (Seoul.)* 37 (2022) 858–860, <https://doi.org/10.3803/EnM.2022.605>.
- [71] J.H. Oh, D.W. Jun, H.Y. Kim, S.M. Lee, E.L. Yoon, J. Hwang, J.H. Park, H. Lee, W. Kim, H. Kim, Discovery of dipeptidyl peptidase-4 inhibitor specific biomarker in non-alcoholic fatty liver disease mouse models using modified basket trial, *Clin. Mol. Hepatol.* 28 (2022) 497–509, <https://doi.org/10.3350/cmh.2022.0019>.
- [72] S. Alam, J. Ghosh, G. Mustafa, M. Kamal, N. Ahmad, Effect of sitagliptin on hepatic histological activity and fibrosis of nonalcoholic steatohepatitis patients: a 1-year randomized control trial, *Hepat. Med* 10 (2018) 23–31, <https://doi.org/10.2147/HMER.S158053>.
- [73] G. Nguyen, S.Y. Park, D.V. Do, D.-H. Choi, E.-H. Cho, Gemigliptin alleviates succinate-induced hepatic stellate cell activation by ameliorating mitochondrial dysfunction, *Endocrinol. Metab.* 37 (2022) 918–928, <https://doi.org/10.3803/EnM.2022.1530>.
- [74] G. Fan, F. Li, P. Wang, X. Jin, R. Liu, Natural-product-mediated autophagy in the treatment of various liver diseases, *Int J. Mol. Sci.* 23 (2022) 15109, <https://doi.org/10.3390/ijms232315109>.
- [75] S.-Y. Chen, X.-Q. Kong, K.-F. Zhang, S. Luo, F. Wang, J.-J. Zhang, DPP4 as a potential candidate in cardiovascular disease, *J. Inflamm. Res* 15 (2022) 5457–5469, <https://doi.org/10.2147/JIR.S380285>.
- [76] W. Wb, C. Cp, H. Sr, N. Se, B. Rm, B. Gl, P. At, F. Pr, M. Cr, S. K, C. W, C. Wc, F. Z, Alogliptin after acute coronary syndrome in patients with type 2 diabetes, *N. Engl. J. Med.* 369 (2013), <https://doi.org/10.1056/NEJMoa1305889>.
- [77] S. Bm, B. Dl, E. B, S. Pg, J. D, B. H, P. O, R. F, W. Sd, H. Eb, C. Ma, U. Ja, D. Nr, O. M, M. Dk, R. Kk, L. La, I. R, Saxagliptin and cardiovascular outcomes in patients with type 2 diabetes mellitus, *N. Engl. J. Med.* 369 (2013), <https://doi.org/10.1056/NEJMoa1307684>.
- [78] J.B. Green, M.A. Bethel, P.W. Armstrong, J.B. Buse, S.S. Engel, J. Garg, R. Josse, K. D. Kaufman, J. Koglin, S. Korn, J.M. Lachin, D.K. McGuire, M.J. Pencina, E. Standl, P.P. Stein, S. Suryawanshi, F. Van de Werf, E.D. Peterson, R.R. Holman, TECOS Study Group, Effect of sitagliptin on cardiovascular outcomes in type 2 diabetes, *N. Engl. J. Med* 373 (2015) 232–242, <https://doi.org/10.1056/NEJMoa1501352>.
- [79] J. Rosenstock, V. Perkovic, O.E. Johansen, M.E. Cooper, S.E. Kahn, N. Marx, J. H. Alexander, M. Pencina, R.D. Toto, C. Wanner, B. Zinman, H.J. Woerle, D. Baanstra, E. Pfarr, S. Schnaidt, T. Meinicke, J.T. George, M. von Eynatten, D. K. McGuire, CARMELINA Investigators, Effect of linagliptin vs placebo on major cardiovascular events in adults with type 2 diabetes and high cardiovascular and renal risk: the CARMELINA randomized clinical trial, *JAMA* 321 (2019) 69–79, <https://doi.org/10.1001/jama.2018.18269>.
- [80] J. R, K. Se, J. Oe, B. Z, E. Ma, W. Hj, E. P, A. K, M. M, D. B, T. M, G. Jt, M. von E, M. Dk, N. M, Effect of linagliptin vs glimepiride on major adverse cardiovascular outcomes in patients with type 2 diabetes: the carolina randomized clinical trial, *JAMA* 322 (2019), <https://doi.org/10.1001/jama.2019.13772>.
- [81] M.A. Nauck, J.J. Meier, M.A. Cavender, M. Abd El Aziz, D.J. Drucker, Cardiovascular actions and clinical outcomes with glucagon-like peptide-1 receptor agonists and dipeptidyl peptidase-4 inhibitors, *Circulation* 136 (2017) 849–870, <https://doi.org/10.1161/CIRCULATIONAHA.117.028136>.
- [82] N.A. Subrahmanyam, R.M. Koshy, K. Jacob, J.M. Pappachan, Efficacy and cardiovascular safety of DPP-4 inhibitors, *Curr. Drug Saf.* 16 (2021) 154–164, <https://doi.org/10.2174/1574886315999200819150544>.
- [83] J.J.V. McMurray, P. Ponikowski, G.B. Bolli, V. Lukashevich, P. Kozlovski, W. Kothny, J.D. Lewsey, H. Krum, VIVID trial committees and investigators, effects of vildagliptin on ventricular function in patients with type 2 diabetes mellitus and heart failure: a randomized placebo-controlled trial, *JACC Heart Fail* 6 (2018) 8–17, <https://doi.org/10.1016/j.jchf.2017.08.004>.
- [84] I. Akoumianakis, I. Badi, G. Douglas, S. Chuaiphichai, L. Herdman, N. Akawi, M. Margaritis, A.S. Antonopoulos, E.K. Oikonomou, C. Psarros, N. Galiatsatos, D. Tousoulis, A. Kardos, R. Sayeed, G. Krassopoulos, M. Petrou, U. Schwahn, P. Wohlfart, N. Tennagels, K.M. Channon, C. Antoniades, Insulin-induced vascular redox dysregulation in human atherosclerosis is ameliorated by dipeptidyl peptidase 4 inhibition, *Sci. Transl. Med* 12 (2020) eaav8824, <https://doi.org/10.1126/scitranslmed.aav8824>.
- [85] DUSP26 induces aortic valve calcification by antagonizing MDM2-mediated ubiquitination of DPP4 in human valvular interstitial cells | *European Heart Journal* | Oxford Academic, (n.d.). <https://academic.oup.com/eurheartj/article/42/30/2935/6310285?login=false> (accessed September 2, 2024)..
- [86] K.C. Nogueira, M. Furtado, R.T. Fukui, M.R.S. Correia, R.F. dos Santos, J. L. Andrade, M.E. Rossi da Silva, Left ventricular diastolic function in patients with type 2 diabetes treated with a dipeptidyl peptidase-4 inhibitor: a pilot study, *Diabetol. Metab. Syndr.* 6 (2014) 103, <https://doi.org/10.1186/1758-5996-6-103>.
- [87] N. S, B. Al, S. Mb, L. Ds, Obesity and impaired metabolic health in patients with COVID-19, *Nat. Rev. Endocrinol.* 16 (2020), <https://doi.org/10.1038/s41574-020-0364-6>.
- [88] N. Stefan, SARS-CoV-2 fires up inflammation in adipose tissue, *Nat. Rev. Endocrinol.* 19 (2023) 8–9, <https://doi.org/10.1038/s41574-022-00778-0>.
- [89] K. Schlicht, N. Rohmann, C. Geisler, T. Hollstein, C. Knappe, K. Hartmann, J. Schwarz, F. Tran, D. Schunk, R. Junker, T. Bahner, P. Rosenstiel, D. Schulte, K. Türk, A. Franke, S. Schreiber, M. Laudes, Correction: circulating levels of soluble dipeptidylpeptidase-4 are reduced in human subjects hospitalized for severe COVID-19 infections, *Int J. Obes. (Lond.)* 46 (2022) 243, <https://doi.org/10.1038/s41366-021-00988-y>.
- [90] N. Vankadari, J.A. Wilce, Emerging WuHan (COVID-19) coronavirus: glycan shield and structure prediction of spike glycoprotein and its interaction with human CD26, *Emerg. Microbes Infect.* 9 (2020) 601–604, <https://doi.org/10.1080/22221751.2020.1739565>.
- [91] L.L. Baggio, E.M. Varin, J.A. Koehler, X. Cao, Y. Lohknygina, S.R. Stevens, R. R. Holman, D.J. Drucker, Plasma levels of DPP4 activity and sDPP4 are dissociated from inflammation in mice and humans, *Nat. Commun.* 11 (2020) 3766, <https://doi.org/10.1038/s41467-020-17556-z>.
- [92] E.M. Varin, E.E. Mulvihill, J.L. Beaudry, G. Pujadas, S. Fuchs, J.-F. Tanti, S. Fazio, K. Kaur, X. Cao, L.L. Baggio, D. Matthews, J.E. Campbell, D.J. Drucker, Circulating levels of soluble dipeptidyl peptidase-4 are dissociated from inflammation and induced by enzymatic DPP4 inhibition, *e5, Cell Metab.* 29 (2019) 320–334, <https://doi.org/10.1016/j.cmet.2018.10.001>.
- [93] Z.D. Kifle, A.E. Woldeyohanin, C.A. Demeke, SARS-CoV-2 and diabetes: a potential therapeutic effect of dipeptidyl peptidase 4 inhibitors in diabetic patients diagnosed with COVID-19, *Metab. Open* 12 (2021) 100134, <https://doi.org/10.1016/j.metop.2021.100134>.
- [94] W.P.T. James, WHO recognition of the global obesity epidemic, *Int J. Obes. (Lond.)* 32 (Suppl 7) (2008) S120–S126, <https://doi.org/10.1038/ijo.2008.247>.
- [95] A. De Lorenzo, L. Romano, L. Di Renzo, N. Di Lorenzo, G. Cennamo, P. Gualtieri, Obesity: a preventable, treatable, but relapsing disease, *Nutrition* 71 (2020) 110615, <https://doi.org/10.1016/j.nut.2019.110615>.
- [96] T. Burki, European commission classifies obesity as a chronic disease, *Lancet Diabetes Endocrinol.* 9 (2021) 418, [https://doi.org/10.1016/S2213-8587\(21\)00145-5](https://doi.org/10.1016/S2213-8587(21)00145-5).
- [97] M.S. Mohammed, S. Sendra, J. Lloret, I. Bosch, Systems and WBANs for controlling obesity, *J. Health Eng.* 2018 (2018) 1564748, <https://doi.org/10.1155/2018/1564748>.